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(57) Abstract

Thermostable glycosidase enzymes derived from various Thermococcus, Staphylothermus and Pyrococcus organisms is disclosed. The enzymes are produced from native or recombinant host cells and can be utilized in the food processing industry, pharmaceutical industry and in the textile industry, detergent industry and in the baking industry.

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GLYCOSIDASE ENZYMES

BACKGROUND OF THE INVENTION

1. Field of the Inventions

This invention relates to newly identified polynucleotides, polypeptides encoded by such polynucleotides, the use of such polynucleotides and polypeptides, as well as the production and isolation of such polynucleotides and polypeptides. More particularly, the polynucleotides and polypeptides of the present invention has been putatively identified as glucosidases, α -galactosidases, β -galactosidases, β -mannosidases, β -mannanases, endoglucanases, and pullalanases.

2. Description of Related Art

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The glycosidic bond of β -galactosides can be cleaved by different classes of enzymes: (i) phospho- β -galactosidases (EC3.2.1.85) are specific for a phosphorylated substrate generated via phosphoenolpyruvate phosphotransferase system (PTS)-dependent uptake; (ii) typical β-galactosidases (EC 3.2.1.23), represented by the Escherichia coli LacZ enzyme, which are relatively specific for β -galactosides; and (iii) β -glucosidases (EC 3.2.1.21) such as the enzymes of Agrobacterium faecalis, Clostridium thermocellum, Pyrococcus furiosus or Sulfolobus solfataricus (Day, A.G. and Withers, S.G., (1986) Purification and characterization of a β -glucosidase from Alcaligenes faecalis. Can. J. Biochem. Cell. Biol. 64, 914-922; Kengen, S.W.M., et al. (1993) Eur. J. Biochem., 213, 305-312; Ait, N., Cruezet, N. and Cattaneo, J. (1982) Properties of β -glucosidase purified from Clostridium thermocellum. J. Gen. Microbiol. 128, 569-577; Grogan, D.W. (1991) Evidence that β-galactosidase of Sulfolobus solfataricus is only one of several activities of a thermostable β -D-glycodiase. Appl. Environ. Microbiol. 57, 1644-1649). Members of the latter group, although highly specific with respect to the β -anomeric configuration of the glycosidic linkage, often display a rather relaxed substrate specificity and hydrolyze β glucosides as well as β -fucosides and β -galactosides.

Generally, α -galactosidases are enzymes that catalyze the hydrolysis of galactose groups on a polysaccharide backbone or hydrolyze the cleavage of di- or oligosaccharides comprising galactose.

Generally, \(\beta\)-mannanases are enzymes that catalyze the hydrolysis of mannose groups internally on a polysaccharide backbone or hydrolyze the cleavage of di- or oligosaccaharides comprising mannose groups. \(\beta\)-mannosidases hydrolyze non-reducing, terminal mannose residues on a mannose-containing polysaccharide and the cleavage of di- or oligosaccaharides comprising mannose groups.

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Guar gum is a branched galactomannan polysaccharide composed of β -1,4 linked mannose backbone with α -1,6 linked galactose side chains. The enzymes required for the degradation of guar are β -mannanase, β -mannosidase and α -galactosidase. β -mannanase hydrolyses the mannose backbone internally and β -mannosidase hydrolyses non-reducing. terminal mannose residues. α -galactosidase hydrolyses α -linked galactose groups.

Galactomannan polysaccharides and the enzymes that degrade them have a variety of applications. Guar is commonly used as a thickening agent in food and is utilized in hydraulic fracturing in oil and gas recovery. Consequently, galactomannanases are industrially relevant for the degradation and modification of guar. Furthermore, a need exists for thermostable galactomannases that are active in extreme conditions associated with drilling and well stimulation.

There are other applications for these enzymes in various industries, such as in the beet sugar industry. 20-30% of the domestic U.S. sucrose consumption is sucrose from sugar beets. Raw beet sugar can contain a small amount of raffinose when the sugar beets are stored before processing and rotting begins to set in. Raffinose inhibits the crystallization of sucrose and also constitutes a hidden quantity of sucrose. Thus, there is merit to eliminating raffinose from raw beet sugar. α -Galactosidase has also been used as a digestive aid to break down raffinose, stachyose, and verbascose in such foods as beans and other gassy foods.

β-galactosidases which are active and stable at high temperatures appear to be superior enzymes for the production of lactose-free dietary milk products (Chaplin, M.F.

and Bucke, C. (1990) In: Enzyme Technology, pp. 159-160, Cambridge University Press, Cambridge, UK). Also, several studies have demonstrated the applicability of β-galactosidases to the enzymatic synthesis of oligosaccharides via transglycosylation reactions (Nilsson, K.G.I. (1988) Enzymatic synthesis of oligosaccharides. Trends Biotechnol. 6, 156-264; Cote, G.L. and Tao, B.Y. (1990) Oligosaccharide synthesis by enzymatic transglycosylation. Glycoconjugate J. 7, 145-162). Despite the commercial potential, only a few β-galactosidases of thermophiles have been characterized so far. Two genes reported are β-galactoside-cleaving enzymes of the hyperthermophilic bacterium Thermotoga maritima, one of the most thermophilic organotrophic eubacteria described to date (Huber, R., Langworthy, T.A., König, H., Thomm, M., Woese, C.R., Sleytr, U.B. and Stetter, K.O. (1986) T. martima sp. nov. represents a new genus of unique extremely thermophilic eubacteria growing up to 90°C, Arch. Microbiol. 144, 324-333) one of the most thermophilic organotrophic eubacteria described to date. The gene products have been identified as a β-galactosidase and a β-glucosidase.

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Pullulanase is well known as a debranching enzyme of pullulan and starch. The enzyme hydrolyzes α -1,6-glucosidic linkages on these polymers. Starch degradation for the production or sweeteners (glucose or maltose) is a very important industrial application of this enzyme. The degradation of starch is developed in two stages. The first stage involves the liquefaction of the substrate with α -amylase, and the second stage, or saccharification stage, is performed by β -amylase with pullalanase added as a debranching enzyme, to obtain better yields.

Endoglucanases can be used in a variety of industrial applications. For instance, the endoglucanases of the present invention can hydrolyze the internal β-1,4-glycosidic bonds in cellulose, which may be used for the conversion of plant biomass into fuels and chemicals. Endoglucanases also have applications in detergent formulations, the textile industry, in animal feed, in waste treatment, and in the fruit juice and brewing industry for the clarification and extraction of juices.

Brief Description of the Drawings

The following drawings are illustrative of embodiments of the invention and are not meant to limit the scope of the invention as encompassed by the claims.

Figures 1a-b are the full-length DNA and corresponding deduced amino acid sequence of M11TL of the present invention. Sequencing was performed using a 378 automated DNA sequencer for all sequences of the present invention (Applied Biosystems, Inc.).

Figure 2 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of OC1/4V-33B/G.

Figure 3 is an illustration of the full-length DNA and corresponding deduced amino acid sequence of F1-12G.

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Figures 4a-b are the full-length DNA and corresponding deduced amino acid sequence of 9N2-31B/G.

Figures 5a-b are the full-length DNA and corresponding deduced amino acid sequence of MSB8-6G.

Figure 6 is the full-length DNA and corresponding deduced amino acid sequence of AEDII12RA-18B/G.

Figures 7a-b are the full-length DNA and corresponding deduced amino acid sequence of GC74-22G.

Figures 8a-b are the full-length DNA and corresponding deduced amino acid sequence of VC1-7G1.

Figures 9a-c are the full-length DNA and corresponding deduced amino acid sequence of 37GP1.

Figures 10a-c are the full-length DNA and corresponding deduced amino acid sequence of 6GC2.

Figures 11a-d are the full-length DNA and corresponding deduced amino acid sequence of 6GP2.

Figures 12a-c are the full-length DNA and corresponding deduced amino acid sequence of 63GB1.

Figures 13a-b are the full-length DNA and corresponding deduced amino acid sequence of OC1/4V.

Figures 14a-e are the full-length DNA and corresponding deduced amino acid sequence of 6GP3.

Figures 15a-d are the full-length DNA and corresponding deduced amino acid sequence of *Thermotoga maritima* MSB8-6GP2.

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Figures 16a-c are the full-length DNA and corresponding deduced amino acid sequence of *Thermotoga maritima* MSB8-6GB4.

Figures 17a-d are the full-length DNA and corresponding deduced amino acid sequence of *Banki gouldi* 37GP4.

Figures 18a-b are the full-length DNA and corresponding deduced amino acid sequence of *Pyrococcus furiosus* VC1-7EG1.

SUMMARY OF THE INVENTION

In a preferred embodiment of the present invention, there are provided isolated nucleic acids (polynucleotides) which encode mature enzymes having the deduced amino acid sequences of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64).

In another embodiment, the invention provides a method for producing a polypeptide including culturing host cells containing the polynucleotide of Figures 1-18 and expressing from the host cell a polypeptide encoded by the polynucleotide and isolating the polypeptide.

In another embodiment, the invention provides an enzyme selected from the group consisting of an enzyme having an amino acid sequence set forth in SEQ ID NOS: 15-28 or 61-64 and an enzyme which has at least 30 consecutive amino acid residue as an enzyme having an amino acid sequence set forth in SEQ ID NOS: 15-28 or 61-64.

In yet another embodiment, the invention provides a method for generating glucose from soluble cell oligosaccharides which includes contacting a sample containing oligosaccharides with an effective amount of an enzyme selected from the group of

enzymes having the amino acid sequence set forth in SEQ ID NOS: 15-28, 61-63 and 64 such that glucose is produced

The publications discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

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Definitions

"Monosaccharide", as used herein, refers to a single polyhydroxy aldehyde or ketone unit.

"Oligosaccharide", as used herein, consist of short chains of monosaccharide units joined together by covalent bonds. Of these, the most abundant are the disaccharides, which have two monosaccharide units.

"Polysaccharide", as used herein, consists of long chains having many monosaccharide units.

The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as intervening sequences (introns) between individual coding segments (exons).

A coding sequence is "operably linked to" another coding sequence when RNA polymerase will transcribe the two coding sequences into a single mRNA, which is then translated into a single polypeptide having amino acids derived from both coding sequences. The coding sequences need not be contiguous to one another so long as the expressed sequences ultimately process to produce the desired protein.

"Recombinant" enzymes refer to enzymes produced by recombinant DNA techniques; *i.e.*, produced from cells transformed by an exogenous DNA construct encoding the desired enzyme. "Synthetic" enzymes are those prepared by chemical synthesis.

A DNA "coding sequence of" or a "nucleotide sequence encoding" a particular enzyme, is a DNA sequence which is transcribed and translated into an enzyme when placed under the control of appropriate regulatory sequences.

Detailed Description of the Invention

The polynucleotides and polypeptides of the present invention have been identified as glucosidases. α -galactosidases, β -galactosidases, β -mannosidases, β -mannanases, endoglucanases, and pullalanases as a result of their enzymatic activity.

In accordance with one aspect of the present invention, there are provided novel enzymes, as well as active fragments, analogs and derivatives thereof.

In accordance with another aspect of the present invention, there are provided isolated nucleic acid molecules encoding the enzymes of the present invention including mRNAs, cDNAs, genomic DNAs as well as active analogs and fragments of such enzymes.

In accordance with yet a further aspect of the present invention, there is provided a process for producing such polypeptides by recombinant techniques comprising culturing recombinant prokaryotic and/or eukaryotic host cells, containing a nucleic acid sequence of the present invention, under conditions promoting expression of said enzymes and subsequent recovery of said enzymes.

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In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes, or polynucleotides encoding such enzymes for hydrolyzing lactose to galactose and glucose for use in the food processing industry, the pharmaceutical industry, for example, to treat intolerance to lactose, as a diagnostic reporter molecule, in corn wet milling, in the fruit juice industry, in baking, in the textile industry and in the detergent industry.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes for hydrolyzing guar gum (a galactomannan polysaccharide) to remove non-reducing terminal mannose residues. Further polysaccharides such as galactomannan and the enzymes according to the invention that degrade them have a variety of applications. Guar gum is commonly used as a thickening agent in food and also is utilized in hydraulic fracturing in oil and gas recovery. Consequently, mannanases are industrially relevant for the degradation and modification of guar gums. Furthermore, a need exists for thermostable mannases that are active in extreme conditions associated with drilling and well stimulation.

In accordance with yet a further aspect of the present invention, there are also provided nucleic acid probes comprising nucleic acid molecules of sufficient length to specifically hybridize to a nucleic acid sequence of the present invention.

In accordance with yet a further aspect of the present invention, there is provided a process for utilizing such enzymes, or polynucleotides encoding such enzymes, for *in vitro* purposes related to scientific research, for example, to generate probes for identifying similar sequences which might encode similar enzymes from other organisms by using certain regions, *i.e.*, conserved sequence regions, of the nucleotide sequence.

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These and other aspects of the present invention should be apparent to those skilled in the art from the teachings herein.

The polynucleotides of this invention were originally recovered from genomic gene libraries derived from the following organisms:

M11TL is a new species of *Desulfurococcus* isolated from Diamond Pool in Yellowstone National Park. The organism grows optimally at 85-88°C, pH 7.0 in a low salt medium containing yeast extract, peptone, and gelatin as substrates with a N₂/CO₂ gas phase.

OC1/4V is from the genus *Thermotoga*. The organism was isolated from Yellowstone National Park. It grows optimally at 75° C in a low salt medium with cellulose as a substrate and N_2 in gas phase.

Pyrococcus furiosus VC1 and (7EG1) is from the genus Pyrococcus. VC1 was isolated from Vulcano, Italy. It grows optimally at 100°C in a high salt medium (marine) containing elemental sulfur, yeast extract, peptone and starch as substrates and N₂ in gas phase.

Staphylothermus marinus F1 is a from the genus Staphylothermus. F1 was isolated from Vulcano, Italy. It grows optimally at $85\,^{\circ}$ C, pH 6.5 in high salt medium (marine) containing elemental sulfur and yeast extract as substrates and N_2 in gas phase.

Thermococcus 9N-2 is from the genus Thermococcus 9N-2 was isolated from diffuse vent fluid in the East Pacific Rise. It is a strict anaerobe that grows optimally at 87°C.

Thermotoga maritima MSB8 and MSB8 (Clone # 6GP2 and 6GB4) is from the genus Thermotogo, and was isolated from Vulcano, Italy. MSB8 grows optimally at 85°C, pH 6.5 in a high salt medium (marine) containing starch and yeast extract as substrates and N₂ in gas phase.

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Thermococcus alcaliphilus AEDII12RA is from the genus Thermococcus. AEDII12RA grows optimally at 85° C, pH 9.5 in a high salt medium (marine) containing polysulfides and yeast extract as substrates and N_2 in gas phase.

Thermococcus chitonophagus GC74 is from the genus Thermococcus. GC74 grows optimally at 85 °C, pH 6.0 in a high salt medium (marine) containing chitin, meat extract, elemental sulfur and yeast extract as substrates and N_2 in gas phase. AEPII 1a grows optimally at 85 °C at pH 6.5 in marine medium under anaerobic conditions. It has many substrates. Bankia gouldi is from the genus Bankia.

Accordingly, the polynucleotides and enzymes encoded thereby are identified by the organism from which they were isolated, and are sometimes hereinafter referred to as "M11TL" (Figure 1 and SEQ ID NOS:1 and 15), "OC1/4V-33B/G" (Figure 2 and SEQ ID NOS:2 and 16), "F1-12G" (Figure 3 and SEQ ID NOS:3 and 17), "9N2-31B/G" (Figure 4 and SEQ ID NOS:4 and 18), "MSB8" (Figure 5 and SEQ ID NOS:5 and 19), "AEDII12RA-18B/G" (Figure 6 and SEQ ID NOS:6 and 20), "GC74-22G" (Figure 7 and SEQ ID NOS:7 and 21), "VC1-7G1" (Figure 8 and SEQ ID NOS:8 and 22), "37GP1" (Figure 9 and SEQ ID NOS: 9 and 23), "6GC2" (Figure 10 and SEQ ID NOS: 10 and 24), "6GP2" (Figure 11 and SEQ ID NOS:11 and 25), "AEPII 1a" (Figure 12 and SEQ ID NOS:12 and 26), "OC1/4V" (Figure 13 and SEQ ID NOS:13 and 27), and "6GP3" (Figure 14 and SEQ ID NOS:28), "MSB8-6GP2" (Figure 15 and SEQ ID NOS:57 and 61), "MSB8-6GB4"(Figure 16 and SEQ ID NOS:58 and 62),"VC1-7EG1"(Figure 17 and SEQ ID NOS:59 and 63), and 37GP4 (Figure 18 and SEQ ID NOS:60 and 64).

The polynucleotides and polypeptides of the present invention show identity at the nucleotide and protein level to known genes and proteins encoded thereby as shown in Table 1.

Table 1

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			Nucleic
	Gene/Protein with	Protein	Acid
Clone	Closest Homology	Identity	Identity
M11TL-29G	Sulfolobus sulfataricus	51%	55%
	DSM 1616/P1, β-		
	galactosidase		
OC1/4V-33B/G	Caldocellum	52%	57%
	saccharolyticum, β-		
	glucosidase		
Staphylothermus	Bacillus polymyxa, β-	36%	48%
marinus F1-12G	galactosidase		
Thermococcus 9N2-	Sulfolobus sulfataricus	51%	50%
31B/G	ATCC 49255/MT4, β-		
	galactosidase		
Thermotoga maritima	Clostridium thermocellum	45%	53%
MSB8-6G	bglB		
Thermococcus	Bacillus polymyxa, β-	34%	48%
AEDII12RA-18B/G	galactosidase		
Thermococcus	Sulfolobus sulfataricus	46%	54%
chitonophagus GC74-	ATCC 49255/MT4, β-		
22G	galactosidase		

Pyrococcus furiosus VC1-7G1	Sulfolobus sulfataricus/MT-4 β-galactosidase	46.4%	52.5%
Thermotoga maritima α-galactosidase (6GC2)	Pediococcus pentosaceaus α-galactosidase	49%	29%
Thermotoga maritima B-mannanase (6GP2)	Aspergillus aculeatus mannanase	56%	37%
AEPII 1a ß- mannosidase (63GB1)	Sulfolobus solfactaricus ß-galactosidase	78%	56%
OC1/4V endoglucanase (33GP1)	Clostridium thermocellum endo-1,4-ß-endoglucanase	65%	43%
Thermotoga maritima pullalanase (6GP3)	Caldocellum saccharolyticum α- destrom 6 glucanohydralase	72	53
Bankia gouldi mix Endoglucanase (37GP1)	None available		

The polynucleotides and enzymes of the present invention show homology to each other as shown in Table 2.

Table 2

Clone	Gene/Protein with Closest Homology	Protein Identity	Nucleic Acid Identity
Staphylothermus marinus F1-12G	Thermococcus AEDII12RA-18B/G, β- galactosidase, glucosidase	55%	57%
Thermococcus 9N2- 31B/G	Thermococcus chitonophagus GC74- 22G-glucosidase`	74%	66%
Pyrococcus furiosus VC1-7G1	Pyrococcus furiosus VC1- 7B/G β-galactosidase	46.4%	54%

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All the clones identified in Tables 1 and 2 encode polypeptides which have α -glycosidase or β -glycosidase activity.

This invention, in addition to the isolated nucleic acid molecules encoding the enzymes of the present invention, also provide substantially similar sequences. Isolated nucleic acid sequences are substantially similar if: (i) they are capable of hybridizing under conditions hereinafter described, to the polynucleotides of SEQ ID NOS: 1-14 and 57-60; (ii) or they encode DNA sequences which are degenerate to the polynucleotides of SEQ ID NOS: 1-14 and 57-60. Degenerate DNA sequences encode the amino acid sequences of SEQ ID NOS:15-28 and 61-64, but have variations in the nucleotide coding sequences. As used herein, substantially similar refers to the sequences having similar identity to the sequences of the instant invention. The nucleotide sequences that are substantially the same can be identified by hybridization or by sequence comparison. Enzyme sequences that are substantially the same can be identified by one or more of the following: proteolytic digestion, gel electrophoresis and/or microsequencing.

One means for isolating the nucleic acid molecules encoding the enzymes of the present invention is to probe a gene library with a natural or artificially designed probe using art recognized procedures (see, for example: Current Protocols in Molecular Biology,

Ausubel F.M. *et al.* (EDS.) Green Publishing Company Assoc. and John Wiley Interscience, New York, 1989, 1992). It is appreciated to one skilled in the art that the polynucleotides of SEQ ID NOS: 1-14 and 57-60 or fragments thereof (comprising at least 12 contiguous nucleotides), are particularly useful probes. Other particular useful probes for this purpose are hybridizable fragments to the sequences of SEQ ID NOS: 1-14 and 57-60 (*i.e.*, comprising at least 12 contiguous nucleotides).

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With respect to nucleic acid sequences which hybridize to specific nucleic acid sequences disclosed herein, hybridization may be carried out under conditions of reduced stringency, medium stringency or even stringent conditions. As an example of oligonucleotide hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at 45 °C in a solution consisting of 0.9 M NaCl, 50 mM NaH₂PO₄, pH 7.0, 5.0 mM Na₂EDTA, 0.5% SDS, 10X Denhardt's, and 0.5 mg/ml polyriboadenylic acid. Approximately 2 X 10⁷ cpm (specific activity 4-9 X 1th cpm/ug) of ³²P end-labeled oligonucleotide probe are then added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature in 1X SET (150 mM NaCl, 20 mM Tris hydrochloride, pH 7.8, 1 mM Na₂EDTA) containing 0.5% SDS, followed by a 30 minute wash in fresh 1X SET at Tm 10°C for the oligonucleotide probe. The membrane is then exposed to auto-radiographic film for detection of hybridization signals.

Stringent conditions means hybridization will occur only if there is at least 90% identity, preferably at least 95% identity and most preferably at least 97% identity between the sequences. Further, it is understood that a section of a 100 bps sequence that is 95 bps in length has 95% identity with the 1090 bps sequence from which it is obtained. See J. Sambrook et al., Molecular Cloning, A Laboratory Manual, 2d Ed., Cold Spring Harbor Laboratory (1989) which is hereby incorporated by reference in its entirety. Also, it is understood that a fragment of a 100 bps sequence that is 95 bps in length has 95% identity with the 100 bps sequence from which it is obtained.

As used herein, a first DNA (RNA) sequence is at least 70% and preferably at least 80% identical to another DNA (RNA) sequence if there is at least 70% and preferably at

least a 80% or 90% identity, respectively, between the bases of the first sequence and the bases of the another sequence, when properly aligned with each other, for example when aligned by BLASTN.

"Identity" as the term is used herein, refers to a polynucleotide sequence which comprises a percentage of the same bases as a reference polynucleotide (SEQ ID NOS:1-14 and 57-60). For example, a polynucleotide which is at least 90% identical to a reference polynucleotide, has polynucleotide bases which are identical in 90% of the bases which make up the reference polynucleotide and may have different bases in 10% of the bases which comprise that polynucleotide sequence.

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The present invention relates polynucleotides which differ from the reference polynucleotide such that the changes are silent changes, for example the change do not alter the amino acid sequence encoded by the polynucleotide. The present invention also relates to nucleotide changes which result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptide encoded by the reference polynucleotide. In a preferred aspect of the invention these polypeptides retain the same biological action as the polypeptide encoded by the reference polynucleotide.

It is also appreciated that such probes can be and are preferably labeled with an analytically detectable reagent to facilitate identification of the probe. Useful reagents include but are not limited to radioactivity, fluorescent dyes or enzymes capable of catalyzing the formation of a detectable product. The probes are thus useful to isolate complementary copies of DNA from other sources or to screen such sources for related sequences.

The polynucleotides of this invention were recovered from genomic gene libraries from the organisms listed in Table 1. For example, gene libraries can be generated in the Lambda ZAP II cloning vector (Stratagene Cloning Systems). Mass excisions can be performed on these libraries to generate libraries in the pBluescript phagemid. Libraries are thus generated and excisions performed according to the protocols/methods hereinafter described.

The excision libraries are introduced into the *E. coli* strain BW14893 F'kan1A. Expression clones are then identified using a high temperature filter assay. Expression clones encoding several glucanases and several other glycosidases are identified and repurified. The polynucleotides, and enzymes encoded thereby, of the present invention, yield the activities as described above.

The coding sequences for the enzymes of the present invention were identified by screening the genomic DNAs prepared for the clones having glucosidase or galactosidase activity.

An example of such an assay is a high temperature filter assay wherein expression clones were identified by use of high temperature filter assays using buffer Z (see recipe below) containing 1 mg/ml of the substrate 5-bromo-4-chloro-3-indolyl-β-D-glucopyranoside (XGLU) (Diagnostic Chemicals Limited or Sigma) after introducing an excision library into the *E. coli* strain BW14893 F'kan1A. Expression clones encoding XGLUases were identified and repurified from M11TL, OC1/4V, Pyrococcus furiosus VC1, Staphylothemus marinus F1, Thermococcus 9N-2, Thermotoga maritima MSB8, Thermococcus alcaliphilus AEDII12RA, and Thermococcus chitonophagus GC74.

Z-buffer: (referenced in Miller, J.H. (1992) A Short Course in Bacterial Genetics, p. 445.)

per liter:

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 $Na_2HPO_4-7H_2O$ 16.1g $NaH_2PO_4-7H_2O$ 5.5g KCl 0.75g $MgSO_4-7H_2O$ 0.246g β-mercaptoethanol 2.7ml

Adjust pH to 7.0

High Temperature Filter Assay

(1) The f factor fkan (from E. coli strain CSH118)(1) was introduced into the pho-phh-lac-strain BW14893(2). BW13893(2). The filamentous phage library was plated on the resulting strain, BW14893 Fkan. (Miller, J.H. (1992) A Short Course in

Bacterial Genetics; Lee, K.S., Metcalf, et al., (1992) Evidence for two phosphonate degradative pathways in Enterobacter Aerogenes, J. Bacteriol., 174:2501-2510.

- (2) After growth on 100 mm LB plates containing 100 μg/ml ampicillin, 80 μg/ml nethicillin and 1mM IPTG, colony lifts were performed using Millipore HATF membrane filters.
- (3) The colonies transferred to the filters were lysed with chloroform vapor in 150 mm glass petri dishes.

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- (4) The filters were transferred to 100 mm glass petri dishes containing a piece of Whatman 3MM filter paper saturated with buffer.
 - (a) when testing for galactosidase activity (XGALase), 3MM paper was saturated with Z buffer containing 1 mg/ml XGAL (ChemBridge Corporation). After transferring filter bearing lysed colonies to the glass petri dish, placed dish in oven at 80-85°C.
 - (b) when testing for glucosidase (XGLUase), 3MM paper was saturated with Z buffer containing 1 mg/ml XGLU. After transferring filter bearing lysed colonies to the glass petri dish, placed dish in oven at 80-85 °C.
- (5) Positives' were observed as blue spots on the filter membranes. Used the following filter rescue technique to retrieve plasmid from lysed positive colony. Used pasteur pipette (or glass capillary tube) to core blue spots on the filter membrane. Placed the small filter disk in an Eppendorf tube containing 20 μl water. Incubated the Eppendorf tube at 75°C for 5 minutes followed by vortexing to elute plasmid DNA off filter. This DNA was transformed into electrocompetent *E. coli* cells DH10B for Thermatoga maritima MSB8-6G, Staphylothermus marinus F1-12G, Thermococcus AEDII12RA-18B/G, Thermococcus chitonophagus GC74-22G, M11Tl and OC1/4V. Electrocompetent BW14893 F'kan1A *E. coli* were used for Thermococcus 9N2-31B/G, and *Pyrococcus furiosus* VC1-7G1. Repeated filter-lift assay on transformation plates to identify 'positives'. Return transformation plates to 37°C incubator after filter lift to regenerate colonies. Inoculate 3 ml LB liquid containing 100 μg/ml ampicillin with repurified positives and incubate at 37°C

overnight. Isolate plasmid DNA from these cultures and sequence plasmid insert. In some instances where the plates used for the initial colony lifts contained non-confluent colonies, a specific colony corresponding to a blue spot on the filter could be identified on a regenerated plate and repurified directly, instead of using the filter rescue technique.

Another example of such an assay is a variation of the high temperature filter assay wherein colony-laden filters are heat-killed at different temperatures (for example, 105°C for 20 minutes) to monitor thermostability. The 3MM paper is saturated with different buffers (i.e., 100 mM NaCl, 5 mM MgCl₂, 100 mM Tris-Cl (pH 9.5)) to determine enzyme activity under different buffer conditions.

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A β -glucosidase assay may also be employed, wherein Glcp β Np is used as an artificial substrate (aryl- β -glucosidase). The increase in absorbance at 405 nm as a result of p-nitrophenol (pNp) liberation was followed on a Hitachi U-1100 spectrophotometer, equipped with a thermostatted cuvette holder. The assays may be performed at 80°C or 90°C in closed 1-ml quartz cuvette. A standard reaction mixture contains 150 mM trisodium substrate, pH 5.0 (at 80°C), and 0.95 mM pNp derivative pNp = 0.561 mM⁻¹ cm⁻¹). The reaction mixture is allowed to reach the desired temperature, after which the reaction is started by injecting an appropriate amount of enzyme (1.06 ml final volume).

1 U β -glucosidase activity is defined as that amount required to catalyze the formation of 1.0 μ mol pNp/min. D-cellobiose may also be used as a substrate.

An ONPG assay for β -galactosidase activity is described by Miller, J.H. (1992) A Short Course in Bacterial Genetics and Mill, J.H. (1992) Experiments in Molecular Genetics, the contents of which are hereby incorporated by reference in their entirety.

A quantitative fluorometric assay for β -galactosidase specific activity is described by : Youngman P., (1987) Plasmid Vectors for Recovering and Exploiting Tn917 Transpositions in Bacillus and other Gram-Positive Bacteria. In Plasmids: A Practical approach (ed. K. Hardy) pp 79-103. IRL Press, Oxford. A description of the procedure can be found in Miller (1992) p. 75-77, the contents of which are incorporated by reference herein in their entirety.

The polynucleotides of the present invention may be in the form of DNA which DNA includes cDNA, genomic DNA, and synthetic DNA. The DNA may be double-stranded or single-stranded, and if single stranded may be the coding strand or non-coding (anti-sense) strand. The coding sequences which encodes the mature enzymes may be identical to the coding sequences shown in Figures 1-8 (SEQ ID NOS: 1-14 and 57-60) or may be a different coding sequence which coding sequence, as a result of the redundancy or degeneracy of the genetic code, encodes the same mature enzymes as the DNA of Figures 1-18 (SEQ ID NOS: 1-14 and 57-60).

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The polynucleotide which encodes for the mature enzyme of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64) may include, but is not limited to: only the coding sequence for the mature enzyme; the coding sequence for the mature enzyme and additional coding sequence such as a leader sequence or a proprotein sequence; the coding sequence for the mature enzyme (and optionally additional coding sequence) and non-coding sequence, such as introns or non-coding sequence 5' and/or 3' of the coding sequence for the mature enzyme.

Thus, the term "polynucleotide encoding an enzyme (protein)" encompasses a polynucleotide which includes only coding sequence for the enzyme as well as a polynucleotide which includes additional coding and/or non-coding sequence.

The present invention further relates to variants of the hereinabove described polynucleotides which encode for fragments, analogs and derivatives of the enzymes having the deduced amino acid sequences of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64). The variant of the polynucleotide may be a naturally occurring allelic variant of the polynucleotide or a non-naturally occurring variant of the polynucleotide.

Thus, the present invention includes polynucleotides encoding the same mature enzymes as shown in Figures 1-18 (SEQ ID NOS: 15-28 and 61-64) as well as variants of such polynucleotides which variants encode for a fragment, derivative or analog of the enzymes of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64). Such nucleotide variants include deletion variants, substitution variants and addition or insertion variants.

As hereinabove indicated, the polynucleotides may have a coding sequence which is a naturally occurring allelic variant of the coding sequences shown in Figures 1-18 (SEQ

ID NOS: 1-14 and 57-60). As known in the art, an allelic variant is an alternate form of a polynucleotide sequence which may have a substitution, deletion or addition of one or more nucleotides, which does not substantially alter the function of the encoded enzyme.

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Fragments of the full length gene of the present invention may be used as a hybridization probe for a cDNA or a genomic library to isolate the full length DNA and to isolate other DNAs which have a high sequence similarity to the gene or similar biological activity. Probes of this type preferably have at least 10, preferably at least 15, and even more preferably at least 30 bases and may contain, for example, at least 50 or more bases. The probe may also be used to identify a DNA clone corresponding to a full length transcript and a genomic clone or clones that contain the complete gene including regulatory and promotor regions, exons, and introns. An example of a screen comprises isolating the coding region of the gene by using the known DNA sequence to synthesize an oligonucleotide probe. Labeled oligonucleotides having a sequence complementary to that of the gene of the present invention are used to screen a library of genomic DNA to determine which members of the library the probe hybridizes to.

The present invention further relates to polynucleotides which hybridize to the hereinabove-described sequences if there is at least 70%, preferably at least 90%, and more preferably at least 95% identity between the sequences. The present invention particularly relates to polynucleotides which hybridize under stringent conditions to the hereinabove-described polynucleotides. As herein used, the term "stringent conditions" means hybridization will occur only if there is at least 95% and preferably at least 97% identity between the sequences. The polynucleotides which hybridize to the hereinabove described polynucleotides in a preferred embodiment encode enzymes which either retain substantially the same biological function or activity as the mature enzyme encoded by the DNA of Figures 1-18 (SEQ ID NOS: 1-14 and 57-60).

Alternatively, the polynucleotide may have at least 15 bases, preferably at least 30 bases, and more preferably at least 50 bases which hybridize to any part of a polynucleotide of the present invention and which has an identity thereto, as hereinabove described, and which may or may not retain activity. For example, such polynucleotides may be employed

as probes for the polynucleotides of SEQ ID NOS: 1-14 and 57-60, for example, for recovery of the polynucleotide or as a diagnostic probe or as a PCR primer.

Thus, the present invention is directed to polynucleotides having at least a 70% identity, preferably at least 90% identity and more preferably at least a 95% identity to a polynucleotide which encodes the enzymes of SEQ ID NOS: 15-28 and 61-64 as well as fragments thereof, which fragments have at least 15 bases, preferably at least 30 bases and most preferably at least 50 bases, which fragments are at least 90% identical, preferably at least 95% identical and most preferably at least 97% identical under stringent conditions to any portion of a polynucleotide of the present invention.

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The present invention further relates to enzymes which have the deduced amino acid sequences of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64) as well as fragments, analogs and derivatives of such enzyme.

The terms "fragment," "derivative" and "analog" when referring to the enzymes of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64) means enzymes which retain essentially the same biological function or activity as such enzymes. Thus, an analog includes a proprotein which can be activated by cleavage of the proprotein portion to produce an active mature enzyme.

The enzymes of the present invention may be a recombinant enzyme, a natural enzyme or a synthetic enzyme, preferably a recombinant enzyme.

The fragment, derivative or analog of the enzymes of Figures 1-18 (SEQ ID NOS: 15-28 and 61-64) may be (i) one in which one or more of the amino acid residues are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code, or (ii) one in which one or more of the amino acid residues includes a substituent group, or (iii) one in which the mature enzyme is fused with another compound, such as a compound to increase the half-life of the enzyme (for example, polyethylene glycol), or (iv) one in which the additional amino acids are fused to the mature enzyme, such as a leader or secretory sequence or a sequence which is employed for purification of the mature enzyme or a proprotein sequence. Such fragments, derivatives

and analogs are deemed to be within the scope of those skilled in the art from the teachings herein.

The enzymes and polynucleotides of the present invention are preferably provided in an isolated form, and preferably are purified to homogeneity.

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The term "isolated" means that the material is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or enzyme present in a living animal is not isolated, but the same polynucleotide or enzyme, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or enzymes could be part of a composition, and still be isolated in that such vector or composition is not part of its natural environment.

The enzymes of the present invention include the enzymes of SEQ ID NOS: 15-28 and 61-64 (in particular the mature enzyme) as well as enzymes which have at least 70% similarity (preferably at least 70% identity) to the enzymes of SEQ ID NOS: 15-28 and 61-64 and more preferably at least 90% similarity (more preferably at least 90% identity) to the enzymes of SEQ ID NOS: 15-28 and 61-64 and still more preferably at least 95% similarity (still more preferably at least 95% identity) to the enzymes of SEQ ID NOS: 15-28 and 61-64 and also include portions of such enzymes with such portion of the enzyme generally containing at least 30 amino acids and more preferably at least 50 amino acids.

As known in the art "similarity" between two enzymes is determined by comparing the amino acid sequence and its conserved amino acid substitutes of one enzyme to the sequence of a second enzyme.

A variant, i.e. a "fragment", "analog" or "derivative" polypeptide, and reference polypeptide may differ in amino acid sequence by one or more substitutions, additions, deletions, fusions and truncations, which may be present in any combination.

Among preferred variants are those that vary from a reference by conservative amino acid substitutions. Such substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the replacements, one for another, among the aliphatic amino acids Ala,

Val, Leu and Ile; interchange of the hydroxyl residues Ser and Thr, exchange of the acidic residues Asp and Glu, substitution between the amide residues Asn and Gln, exchange of the basic residues Lys and Arg and replacements among the aromatic residues Phe, Tyr.

Most highly preferred are variants which retain the same biological function and activity as the reference polypeptide from which it varies.

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Fragments or portions of the enzymes of the present invention may be employed for producing the corresponding full-length enzyme by peptide synthesis: therefore, the fragments may be employed as intermediates for producing the full-length enzymes. Fragments or portions of the polynucleotides of the present invention may be used to synthesize full-length polynucleotides of the present invention.

The present invention also relates to vectors which include polynucleotides of the present invention, host cells which are genetically engineered with vectors of the invention and the production of enzymes of the invention by recombinant techniques.

Host cells are genetically engineered (transduced or transformed or transfected) with the vectors of this invention which may be, for example, a cloning vector or an expression vector. The vector may be, for example, in the form of a plasmid, a viral particle, a phage, etc. The engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the genes of the present invention. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression, and will be apparent to the ordinarily skilled artisan.

The polynucleotides of the present invention may be employed for producing enzymes by recombinant techniques. Thus, for example, the polynucleotide may be included in any one of a variety of expression vectors for expressing an enzyme. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences, e.g., derivatives of SV40; bacterial plasmids; phage DNA; baculovirus; yeast plasmids; vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. However, any other vector may be used as long as it is replicable and viable in the host.

The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is inserted into an appropriate restriction endonuclease site(s) by procedures known in the art. Such procedures and others are deemed to be within the scope of those skilled in the art.

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The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct mRNA synthesis. As representative examples of such promoters, there may be mentioned: LTR or SV40 promoter, the <u>E. coli</u> lac or trp, the phage lambda P_L promoter and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. The expression vector also contains a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression.

In addition, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in <u>E. coli</u>.

The vector containing the appropriate DNA sequence as hereinabove described, as well as an appropriate promoter or control sequence, may be employed to transform an appropriate host to permit the host to express the protein.

As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as <u>E. coli</u>, <u>Streptomyces</u>, <u>Bacillus subtilis</u>; fungal cells, such as yeast; insect cells such as <u>Drosophila S2</u> and <u>Spodoptera Sf9</u>; animal cells such as CHO, COS or Bowes melanoma; adenoviruses; plant cells, etc. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

More particularly, the present invention also includes recombinant constructs comprising one or more of the sequences as broadly described above. The constructs comprise a vector, such as a plasmid or viral vector, into which a sequence of the invention has been inserted, in a forward or reverse orientation. In a preferred aspect of this embodiment, the construct further comprises regulatory sequences, including, for example, a promoter, operably linked to the sequence. Large numbers of suitable vectors and

promoters are known to those of skill in the art. and are commercially available. The following vectors are provided by way of example; Bacterial: pQE70, pQE60, pQE-9 (Qiagen), pD10, psiX174, pBluescript II KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene); ptrc99a, pKK223-3, pKK233-3, pDR540, pRIT5 (Pharmacia); Eukaryotic: pSV2CAT, pOG44, pXT1, pSG (Stratagene) pSVK3, pBPV, pMSG, pSVL (Pharmacia). However, any other plasmid or vector may be used as long as they are replicable and viable in the host.

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Promoter regions can be selected from any desired gene using CAT (chloramphenical transferase) vectors or other vectors with selectable markers. Two appropriate vectors are pKK232-8 and pCM7. Particular named bacterial promoters include lacI, lacZ, T3, T7, gpt, lambda P_R , P_L and trp. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art.

In a further embodiment, the present invention relates to host cells containing the above-described constructs. The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or the host cell can be a prokaryotic cell, such as a bacterial cell. Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis, L., Dibner, M., Battey, I., Basic Methods in Molecular Biology, (1986)).

The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Alternatively, the enzymes of the invention can be synthetically produced by conventional peptide synthesizers.

Mature proteins can be expressed in mammalian cells, yeast, bacteria, or other cells under the control of appropriate promoters. Cell-free translation systems can also be employed to produce such proteins using RNAs derived from the DNA constructs of the present invention. Appropriate cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook, et al., Molecular Cloning: A Laboratory

Manual. Second Edition, Cold Spring Harbor, N.Y., (1989), the disclosure of which is hereby incorporated by reference.

Transcription of the DNA encoding the enzymes of the present invention by higher eukaryotes is increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10 to 300 bp that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, a cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers.

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Generally, recombinant expression vectors will include origins of replication and selectable markers permitting transformation of the host cell, e.g., the ampicillin resistance gene of E. coli and S. cerevisiae TRP1 gene, and a promoter derived from a highly-expressed gene to direct transcription of a downstream structural sequence. Such promoters can be derived from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), α-factor, acid phosphatase, or heat shock proteins, among others. The heterologous structural sequence is assembled in appropriate phase with translation initiation and termination sequences, and preferably, a leader sequence capable of directing secretion of translated enzyme. Optionally, the heterologous sequence can encode a fusion enzyme including an N-terminal identification peptide imparting desired characteristics, e.g., stabilization or simplified purification of expressed recombinant product.

Useful expression vectors for bacterial use are constructed by inserting a structural DNA sequence encoding a desired protein together with suitable translation initiation and termination signals in operable reading phase with a functional promoter. The vector will comprise one or more phenotypic selectable markers and an origin of replication to ensure maintenance of the vector and to, if desirable, provide amplification within the host. Suitable prokaryotic hosts for transformation include <u>E. coli</u>, <u>Bacillus subtilis</u>, <u>Salmonella typhimurium</u> and various species within the genera Pseudomonas, Streptomyces, and Staphylococcus, although others may also be employed as a matter of choice.

As a representative but nonlimiting example, useful expression vectors for bacterial use can comprise a selectable marker and bacterial origin of replication derived from

commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017). Such commercial vectors include, for example, pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden) and GEM1 (Promega Biotec, Madison, WI, USA). These pBR322 "backbone" sections are combined with an appropriate promoter and the structural sequence to be expressed.

Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter is induced by appropriate means (e.g., temperature shift or chemical induction) and cells are cultured for an additional period.

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Cells are typically harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract retained for further purification.

Microbial cells employed in expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents, such methods are well known to those skilled in the art.

Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts, described by Gluzman, Cell, 23:175 (1981), and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa and BHK cell lines. Mammalian expression vectors will comprise an origin of replication. a suitable promoter and enhancer, and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 splice, and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

The enzyme can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing

configuration of the mature protein. Finally, high performance liquid chromatography (HPLC) can be employed for final purification steps.

The enzymes of the present invention may be a naturally purified product, or a product of chemical synthetic procedures, or produced by recombinant techniques from a prokaryotic or eukaryotic host (for example, by bacterial, yeast, higher plant, insect and mammalian cells in culture). Depending upon the host employed in a recombinant production procedure, the enzymes of the present invention may be glycosylated or may be non-glycosylated. Enzymes of the invention may or may not also include an initial methionine amino acid residue.

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 β -galactosidase hydrolyzes lactose to galactose and glucose. Accordingly, the OC1/4V, 9N2-31B/G, AEDII12RA-18B/G and F1-12G enzymes may be employed in the food processing industry for the production of low lactose content milk and for the production of galactose or glucose from lactose contained in whey obtained in a large amount as a by-product in the production of cheese. Generally, it is desired that enzymes used in food processing, such as the aforementioned β -galactosidases, be stable at elevated temperatures to help prevent microbial contamination.

These enzymes may also be employed in the pharmaceutical industry. The enzymes are used to treat intolerance to lactose. In this case, a thermostable enzyme is desired, as well. Thermostable β -galactosidases also have uses in diagnostic applications, where they are employed as reporter molecules.

Glucosidases act on soluble cellooligosaccharides from the non-reducing end to give glucose as the sole product. Glucanases (endo- and exo-) act in the depolymerization of cellulose, generating more non-reducing ends (endo-glucanases, for instance, act on internal linkages yielding cellobiose, glucose and cellooligosaccharides as products). β-glucosidases are used in applications where glucose is the desired product. Accordingly, M11TL, F1-12G, GC74-22G, MSB8-6G, OC1/4V, VC1-7G1, 9N2-31B/G and AEDII12RA18B/G may be employed in a wide variety of industrial applications, including in corn wet milling for the separation of starch and gluten, in the fruit industry for clarification and equipment maintenance, in baking for viscosity reduction, in the textile

industry for the processing of blue jeans, and in the detergent industry as an additive. For these and other applications, thermostable enzymes are desirable.

Antibodies generated against the enzymes corresponding to a sequence of the present invention can be obtained by direct injection of the enzymes into an animal or by administering the enzymes to an animal, preferably a nonhuman. The antibody so obtained will then bind the enzymes itself. In this manner, even a sequence encoding only a fragment of the enzymes can be used to generate antibodies binding the whole native enzymes. Such antibodies can then be used to isolate the enzyme from cells expressing that enzyme.

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For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, 1975, Nature, 256:495-497), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., 1983, Immunology Today 4:72), and the EBV-hybridoma technique to produce human monoclonal antibodies (Cole, et al., 1985, in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96).

Techniques described for the production of single chain antibodies (U.S. Patent 4,946,778) can be adapted to produce single chain antibodies to immunogenic enzyme products of this invention. Also, transgenic mice may be used to express humanized antibodies to immunogenic enzyme products of this invention.

Antibodies generated against the enzyme of the present invention may be used in screening for similar enzymes from other organisms and samples. Such screening techniques are known in the art, for example, one such screening assay is described in "Methods for Measuring Cellulase Activities", *Methods in enzymology*, Vol 160, pp. 87-116, which is hereby incorporated by reference in its entirety.

The present invention will be further described with reference to the following examples; however, it is to be understood that the present invention is not limited to such examples. All parts or amounts, unless otherwise specified, are by weight.

In order to facilitate understanding of the following examples certain frequently occurring methods and/or terms will be described.

"Plasmids" are designated by a lower case p preceded and/or followed by capital letters and/or numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. In addition, equivalent plasmids to those described are known in the art and will be apparent to the ordinarily skilled artisan.

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"Digestion" of DNA refers to catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinarily skilled artisan. For analytical purposes, typically 1 µg of plasmid or DNA fragment is used with about 2 units of enzyme in about 20 µl of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 µg of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer. Incubation times of about 1 hour at 37°C are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion the reaction is electrophoresed directly on a polyacrylamide gel to isolate the desired fragment.

Size separation of the cleaved fragments is performed using 8 percent polyacrylamide gel described by Goeddel, D. et al., Nucleic Acids Res., 8:4057 (1980).

"Oligonucleotides" refers to either a single stranded polydeoxynucleotide or two complementary polydeoxynucleotide strands which may be chemically synthesized. Such synthetic oligonucleotides have no 5' phosphate and thus will not ligate to another oligonucleotide without adding a phosphate with an ATP in the presence of a kinase. A synthetic oligonucleotide will ligate to a fragment that has not been dephosphorylated.

"Ligation" refers to the process of forming phosphodiester bonds between two double stranded nucleic acid fragments (Maniatis, T., et al., Id., p. 146). Unless otherwise provided, ligation may be accomplished using known buffers and conditions with 10 units of T4 DNA ligase ("ligase") per $0.5~\mu g$ of approximately equimolar amounts of the DNA fragments to be ligated.

Unless otherwise stated, transformation was performed as described in the method of Graham, F. and Van der Eb, A., Virology, 52:456-457 (1973).

Example 1

Bacterial Expression and Purification of Glycosidase Enzymes

DNA encoding the enzymes of the present invention, SEQ ID NOS: 1-14 and 57-60 were initially amplified from a pBluescript vector containing the DNA by the PCR technique using the primers noted herein. The amplified sequences were then inserted into the respective PQE vector listed beneath the primer sequences, and the enzyme was expressed according to the protocols set forth herein. The 5' and 3' primer sequences for the respective genes are as follows:

Thermococcus AEDII12RA -18B/G

5' CCGAGAATTCATTAAAGAGGAGAAATTAACTATGGTGAATGCTATGATTGTC 3' (SEQ ID NO:29)

3' CGGAAGATCTTCATAGCTCCGGAAGCCCATA 5' (SEQ ID NO:30)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Blg II.

OC1/4V-33B/G

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5' CCGAGAATTCATTAAAGAGGAGAAATTAACTATGATAAGAAGGTCCGATTTTCC 3' (SEQ ID NO:31)

3' CGGAAGATCTTTAAGATTTTAGAAATTCCTT 5' (SEQ ID NO:32)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Bgl II.

Thermococcus 9N2 - 31B/G

5' CCGAGAATTCATTAAAGAGGAGAAATTAACTATGCTACCAGAAGGCTTTCTC 3' (SEQ ID NO:33)

3' CGGAGGTACCTCACCCAAGTCCGAACTTCTC 5' (SEQ ID NO:34)

Vector: pQE30; and contains the following restriction enzyme sites 5' EcoRI and 3' Kpnl.

Staphylothermus marinus F1 - 12G

5' CCGAGAATTCATTAAAGAGGAGAAATTAACTATGATAAGGTTTCCTGATTAT 3' (SEQ ID NO:35)

3' CGGAAGATCTTTATTCGAGGTTCTTTAATCC 5' (SEQ ID NO:36)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Bgl II.

Thermococcus chitonophagus GC74 - 22G

5' CCGAGAATTCATTAAAGAGGAGAAATTAACTATGCTTCCAGGAGAACTTTCTC 3' (SEQ ID NO:37)

3' CGGAGGATCCCTACCCTCTCTAAGATCTC 5' (SEQ ID NO:38)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' BamHI.

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5' AATAATCTAGAGCATGCAATTCCCCAAAGACTTCATGATAG 3' (SEQ ID NO:39)

3' AATAAAAGCTTACTGGATCAGTGTAAGATGCT 5' (SEQ ID NO:40)

Vector: pQE70; and contains the following restriction enzyme sites 5' SphI and 3' Hind III.

Thermotoga maritima MSB8-6G

5' CCGACAATTGATTAAAGAGGAGAAATTAACTATGGAAAGGATCGATGAAATT 3' (SEQ ID NO:41) 3' CGGAGGTACCTCATGGTTTGAATCTCTTCTC 5' (SEQ ID NO:42)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' KpnI.

Pyrococcus furiosus VC1 - 7G1

5' CCGACAATTGATTAAAGAGGAGAAATTAACTATGTTCCCTGAAAAGTTCCTT 3' (SEQ ID NO:43)

3' CGGAGGTACCTCATCCCCTCAGCAATTCCTC 5' (SEQ ID NO:44)

Vector: pQE12; and contains the following restriction enzyme sites 5' EcoRI and 3' Kpn I.

Bankia gouldi endoglucanase (37GP1)

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5' AATAAGGATCCGTTTAGCGACGCTCGC 3' (SEQ ID NO:45)

3' AATAAAAGCTTCCGGGTTGTACAGCGGTAATAGGC 5' (SEQ ID NO:46)

Vector: pQE52; and contains the following restriction enzyme sites 5' Bam HI and 3' Hind III.

Thermotoga maritima α-galactosidase (6GC2)

5' TTTATTGAATTCATTAAAGAGGAGAAATTAACTATGATCTGTGTGGAAATATTCGGAAAG 3' (SEQ ID NO:47)

3' TCTATAAAGCTTTCATTCTCTCACCCTCTTCGTAGAAG 5' (SEQ ID NO:48)

Vector: pQET; and contains the following restriction enzyme sites 5' EcoRI and 3' Hind III.

Thermotoga maritima \(\beta\)-mannanase (6GP2)

5' TTTATTCAATTGATTAAAGAGGAGAAATTAACTATGGGGATTGGTGGCGACGAC 3' (SEQ ID NO:49)

3' TTTATTAAGCTTATCTTTTCATATTCACATACCTCC 5' (SEQ ID NO:50)

Vector: pQEt; and contains the following restriction enzyme sites 5' Hind III and 3' EcoRI.

AEPII 1a B-mannanase (63GB1)

5' TTTATTGAATTCATTAAAGAGGAGAAATTAACTATGCTACCAGAAGAGTTCCTATGGGGC 3' (SEQ ID NO:51)

3' TTTATTAAGCTTCTCATCAACGGCTATGGTCTTCATTTC 5' (SEQ ID NO:52)

Vector: pQEt; and contains the following restriction enzyme sites 5' Hind III and 3' EcoRI.

OCI/4V endoglucanase (33GP1)

- 5' AAAAAACAATTGAATTCATTAAAGAGGAGAAATTAACTATGGTAGAAAGACACTTCAGATATGTTCTT
- 3' (SEQ ID NO:53)
- 3' TTTTTCGGATCCAATTCTTCATTTACTCTTTGCCTG 5' (SEQ ID NO:54)

Vector: pQEt; and contains the following restriction enzyme sites 5' BamHI and 3' EcoRI.

Thermotoga maritima pullalanase (6GP3)
5' TTTTGGAATTCATTAAAGAGGAGAAATTAACTATGGAACTGATCATAGAAGGTTAC 3'
(SEQ ID NO:55)
3' ATAAGAAGCTTTTCACTCTCTGTACAGAACGTACGC 5' (SEQ ID NO:56)

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Vector: pQEt; and contains the following restriction enzyme sites 5' EcoRI and 3' Hind III.

The restriction enzyme sites indicated correspond to the restriction enzyme sites on the bacterial expression vector indicated for the respective gene (Qiagen, Inc. Chatsworth, CA). The pQE vector encodes antibiotic resistance (Amp^r), a bacterial origin of replication (ori), an IPTG-regulatable promoter operator (P/O), a ribosome binding site (RBS), a 6-His tag and restriction enzyme sites.

The pQE vector was digested with the restriction enzymes indicated. The amplified sequences were ligated into the respective pQE vector and inserted in frame with the sequence encoding for the RBS. The ligation mixture was then used to transform the E. coli strain M15/pREP4 (Qiagen, Inc.) by electroporation. M15/pREP4 contains multiple copies of the plasmid pREP4, which expresses the lacI repressor and also confers kanamycin resistance (Kan'). Transformants were identified by their ability to grow on LB plates and ampicillin/kanamycin resistant colonies were selected. Plasmid DNA was isolated and confirmed by restriction analysis. Clones containing the desired constructs were grown overnight (O/N) in liquid culture in LB media supplemented with both Amp (100 ug/ml) and Kan (25 ug/ml). The O/N culture was used to inoculate a large culture at a ratio of 1:100 to 1:250. The cells were grown to an optical density 600 (O.D.600) of between 0.4 and IPTG ("Isopropyl-B-D-thiogalacto pyranoside") was then added to a final 0.6. concentration of 1 mM. IPTG induces by inactivating the lacI repressor, clearing the P/O leading to increased gene expression. Cells were grown an extra 3 to 4 hours. Cells were then harvested by centrifugation.

The primer sequences set out above may also be employed to isolate the target gene from the deposited material by hybridization techniques described above.

Example 2

Isolation of A Selected Clone From the Deposited genomic clones

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A clone is isolated directly by screening the deposited material using the oligonucleotide primers set forth in Example 1 for the particular gene desired to be isolated. The specific oligonucleotides are synthesized using an Applied Biosystems DNA synthesizer. The oligonucleotides are labeled with ³²P--ATP using T4 polynucleotide kinase and purified according to a standard protocol (Maniatis et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Press, Cold Spring, NY, 1982). The deposited clones in the pBluescript vectors may be employed to transform bacterial hosts which are then plated on 1.5% agar plates to the density of 20,000-50,000 pfu/150 mm plate. These plates are screened using Nylon membranes according to the standard screening protocol (Stratagene, 1993). Specifically, the Nylon membrane with denatured and fixed DNA is prehybridized in 6 x SSC, 20 mM NaH₂PO₄, 0.4%SDS, 5 x Denhardt's 500 μg/ml denatured, sonicated salmon sperm DNA; and 6 x SSC, 0.1% SDS. After one hour of prehybridization, the membrane is hybridized with hybridization buffer 6xSSC, 20 mM NaH, PO₄, 0.4%SDS, 500 ug/ml denatured, sonicated salmon sperm DNA with 1x10⁶ cpm/ml ³²P-probe overnight at 42°C. The membrane is washed at 45-50°C with washing buffer 6 x SSC, 0.1% SDS for 20-30 minutes dried and exposed to Kodak X-ray film overnight. Positive clones are isolated and purified by secondary and tertiary screening. The purified clone is sequenced to verify its identity to the primer sequence.

Once the clone is isolated, the two oligonucleotide primers corresponding to the gene of interest are used to amplify the gene from the deposited material. A polymerase chain reaction is carried out in 25 μ l of reaction mixture with 0.5 ug of the DNA of the gene of interest. The reaction mixture is 1.5-5 mM MgCl₂, 0.01% (w/v) gelatin, 20 μ M each of dATP, dCTP, dGTP, dTTP, 25 pmol of each primer and 0.25 Unit of Taq

polymerase. Thirty five cycles of PCR (denaturation at 94°C for 1 min; annealing at 55°C for 1 min; elongation at 72°C for 1 min) are performed with the Perkin-Elmer Cetus automated thermal cycler. The amplified product is analyzed by agarose gel electrophoresis and the DNA band with expected molecular weight is excised and purified. The PCR product is verified to be the gene of interest by subcloning and sequencing the DNA product. The ends of the newly purified genes are nucleotide sequenced to identify full length sequences. Complete sequencing of full length genes is then performed by Exonuclease III digestion or primer walking.

Example 3

Screening for Galactosidase Activity

Screening procedures for α -galactosidase protein activity may be assayed for as follows:

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Substrate plates were provided by a standard plating procedure. Dilute XL1-Blue MRF E coli host of (Stratagene Cloning Systems, La Jolla, CA) to O.D.₆₀₀ = 1.0 with NZY media. In 15 ml tubes, inoculate 200 μ l diluted host cells with phage. Mix gently and incubate tubes at 37 °C for 15 min. Add approximately 3.5 ml LB top agarose (0.7%) containing 1mM IPTG to each tube and pour onto all NYZ plate surface. Allow to cool and incubate at 37 °C overnight. The assay plates are obtained as substrate p-Nitrophenyl α -galactosidase (Sigma) (200 mg/100 ml) (100 mM NaCl, 100 mM Potassium-Phosphate) 1% (w/v) agarose. The plaques are overlayed with nitrocellulose and incubated at 4 °C for 30 minutes whereupon the nitrocellulose is removed and overlayed onto the substrate plates. The substrate plates are then incubated at 70 °C for 20 minutes.

Example 4

Screening of Clones for Mannanase Activity

A solid phase screening assay was utilized as a primary screening method to test clones for ß-mannanase activity.

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A culture solution of the Y1090-*E. coli* host strain (Stratagene Cloning Systems, La Jolla, CA) was diluted to O.D.₆₀₀=1.0 with NZY media. The amplified library from *Thermotoga maritima* lambda gtl1 library was diluted in SM (phage dilution buffer): $5 \times 10^7 \, \text{pfu/µl}$ diluted 1:1000 then 1:100 to $5 \times 10^2 \, \text{pfu/µl}$. Then 8 µl of phage dilution ($5 \times 10^2 \, \text{pfu/µl}$) was plated in 200 µl host cells. They were then incubated in 15 ml tubes at 37 °C for 15 minutes.

Approximately 4 ml of molten, LB top agarose (0.7%) at approximately 52 °C was added to each tube and the mixture was poured onto the surface of LB agar plates. The agar plates were then incubated at 37 °C for five hours. The plates were replicated and induced with 10 mM IPTG-soaked Duralon-UVTM nylon membranes (Stratagene Cloning Systems, La Jolla, CA) overnight. The nylon membranes and plates were marked with a needle to keep their orientation and the nylon membranes were then removed and stored at 4 °C.

An Azo-galactomannan overlay was applied to the LB plates containing the lambda plaques. The overlay contains 1% agarose, 50 mM potassium-phosphate buffer pH 7, 0.4% Azocarob-galactomannan. (Megazyme, Australia). The plates were incubated at 72 °C. The Azocarob-galactomannan treated plates were observed after 4 hours then returned to incubation overnight. Putative positives were identified by clearing zones on the Azocarob-galactomannan plates. Two positive clones were observed.

The nylon membranes referred to above, which correspond to the positive clones were retrieved, oriented over the plate and the portions matching the locations of the clearing zones for positive clones were cut out. Phage was eluted from the membrane cut-out portions by soaking the individual portions in 500 μ l SM (phage dilution buffer) and 25 μ l CHCl₃.

Example 5

Screening of Clones for Mannosidase Activity

A solid phase screening assay was utilized as a primary screening method to test clones for \(\beta \)-mannosidase activity.

A culture solution of the Y1090-*E. coli* host strain (Stratagene Cloning Systems, La Jolla, CA) was diluted to O.D. $_{600}$ =1.0 with NZY media. The amplified library from AEPII 1a lambda gtl1 library was diluted in SM (phage dilution buffer): 5×10^7 pfu/µl diluted 1:1000 then 1:100 to 5×10^2 pfu/µl. Then 8 µl of phage dilution (5×10^2 pfu/µl) was plated in 200 µl host cells. They were then incubated in 15 ml tubes at 37 °C for 15 minutes.

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Approximately 4 ml of molten, LB top agarose (0.7%) at approximately 52 °C was added to each tube and the mixture was poured onto the surface of LB agar plates. The agar plates were then incubated at 37 °C for five hours. The plates were replicated and induced with 10 mM IPTG-soaked Duralon-UVTM nylon membranes (Stratagene Cloning Systems, La Jolla, CA) overnight. The nylon membranes and plates were marked with a needle to keep their orientation and the nylon membranes were then removed and stored at 4 °C.

A p-nitrophenyl-\$\beta\$-D-manno-pyranoside overlay was applied to the LB plates containing the lambda plaques. The overlay contains 1% agarose, 50 mM potassium-phosphate buffer pH 7, 0.4% p-nitrophenyl-\$\beta\$-D-manno-pyranoside. (Megazyme, Australia). The plates were incubated at 72 °C. The p-nitrophenyl-\$\beta\$-D-manno-pyranoside treated plates were observed after 4 hours then returned to incubation overnight. Putative positives were identified by clearing zones on the p-nitrophenyl-\$\beta\$-D-manno-pyranoside plates. Two positive clones were observed.

The nylon membranes referred to above, which correspond to the positive clones were retrieved, oriented over the plate and the portions matching the locations of the clearing zones for positive clones were cut out. Phage was eluted from the membrane cut-out portions by soaking the individual portions in 500 μ l SM (phage dilution buffer) and 25 μ l CHCl₃.

Example 6

Screening for Pullulanase Activity

Screening procedures for pullulanase protein activity may be assayed for as follows:

Substrate plates were provided by a standard plating procedure. Host cells are diluted to $O.D._{600} = 1.0$ with NZY or appropriate media. In 15 ml tubes, inoculate 200 μ l diluted host cells with phage. Mix gently and incubate tubes at 37 °C for 15 min. Add approximately 3.5 ml LB top agarose (0.7%) is added to each tube and the mixture is plated, allowed to cool, and incubated at 37 °C for about 28 hours. Overlays of 4.5 mls of the following substrate are poured:

100 ml total volume

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0.5g	Red Pullulan Red (Megazyme, Australia)
1.0g	Agarose
5ml	Buffer (Tris-HCL pH 7.2 @ 75 °C)
2ml	5M NaCl
5ml	CaCl ₂ (100mM)
85ml	dH ₂ O

Plates are cooled at room temperature, and thenm incubated at 75°C for 2 hours. Positives are observed as showing substrate degradation.

Example 7

Screening for Endoglucanase Activity

Screening procedures for endoglucanase protein activity may be assayed for as follows:

1. The gene library is plated onto 6 LB/GelRite/0.1% CMC/NZY agar plates (~4,800 plaque forming units/plate) in E.coli host with LB agarose as top agarose. The plates are incubated at 37°C overnight.

- 2. Plates are chilled at 4°C for one hour.
- 3. The plates are overlayed with Duralon membranes (Stratagene) at room temperature for one hour and the membranes are oriented and lifted off the plates and stored at 4°C.
- 4. The top agarose layer is removed and plates are incubated at 37°C for -3 hours.
 - 5. The plate surface is rinsed with NaCl.

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- 6. The plate is stained with 0.1% Congo Red for 15 minutes.
- 7. The plate is destained with 1M NaCl.
- 8. The putative positives identified on plate are isolated from the Duralon membrane (positives are identified by clearing zones around clones). The phage is eluted from the membrane by incubating in $500\mu l\ SM + 25\mu l\ CHCl_3$ to elute.
- 9. Insert DNA is subcloned into any appropriate cloning vector and subclones are reassayed for CMCase activity using the following protocol:
- i) Spin 1ml overnight miniprep of clone at maximum speed for 3 minutes.
- ii) Decant the supernatant and use it to fill "wells" that have been made in an LB/GelRite/0.1% CMC plate.
 - iii) Incubate at 37°C for 2 hours.
 - iv) Stain with 0.1% Congo Red for 15 minutes.
 - v) Destain with 1M NaCl for 15 minutes.
 - vi) Identify positives by clearing zone around clone.

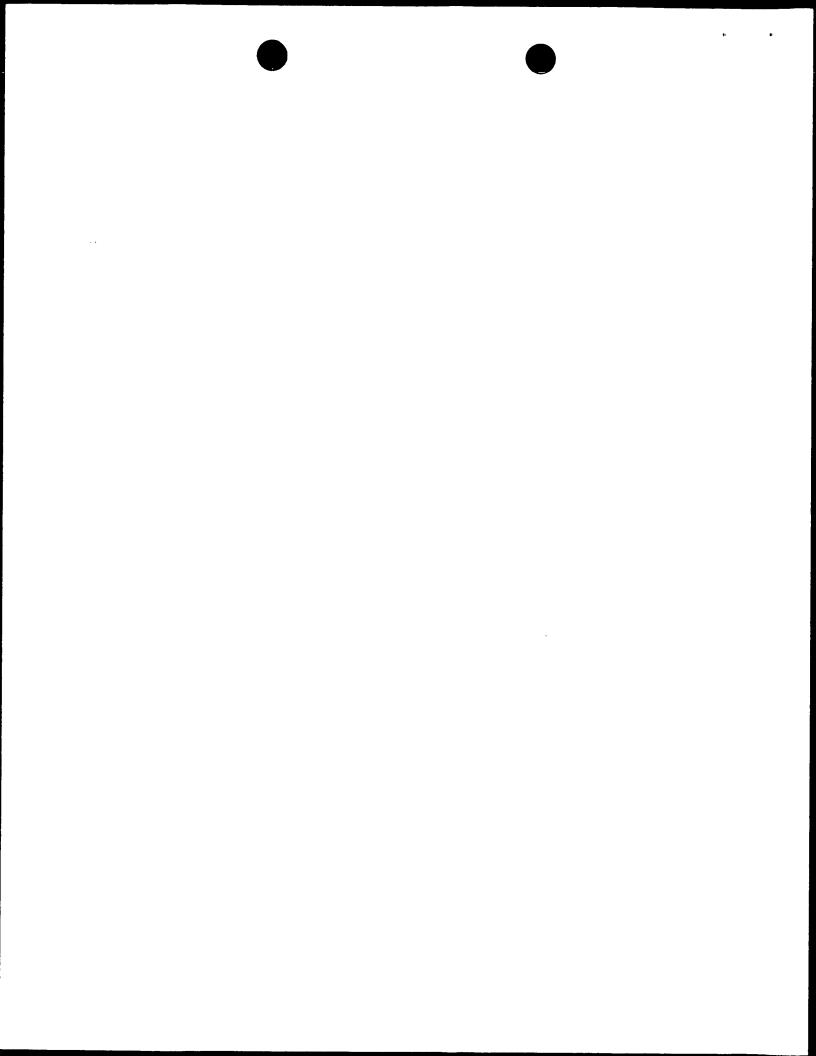
Numerous modifications and variations of the present invention are possible in light of the above teachings and, therefore, within the scope of the appended claims, the invention may be practiced otherwise than as particularly described.

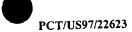
WHAT IS CLAIMED IS:

1. An isolated polynucleotide selected from the group consisting of:

- (a) SEQ ID NOS: 1-14 and 57-60;
- (b) SEQ ID NOS: 1-14 and 57-60, wherein T can also be U;
- (c) polynucleotide sequences complementary to SEQ ID NOS: 1-14 and 57-60;
- (d) polynucleotide sequences which encode an amino acid sequence as set forth in SEQ ID NOS:15-28, and 61-64; and
- (e) fragments of (a), (b), (c) or (d) that are at least 15 consecutive bases in length and that will selectively hybridize to DNA which encodes a polypeptide of SEQ ID NOS:15-28, and 61-64.
- 2. A vector comprising a polynucleotide of claim 1.
- 3. A host cell containing the vector of claim 2.
- 4. The method of claim 3, wherein the host cell is a eukaryotic cell.
- 5. The method of claim 3, wherein the host cell is a prokaryotic cell.
- 6. A method for producing a polypeptide comprising:
 - (a) culturing the host cells of claim 3;
 - (b) expressing from the host cell of claim 3 a polypeptide encoded by said polynucleotide; and
 - (c) isolating the polypeptide.

- 7. An enzyme selected from the group consisting of:
 - (a) an enzyme comprising an amino acid sequence set forth in SEQ ID NOS: 15-28 or 61-64; and
 - (b) an enzyme which comprises at least 30 consecutive amino acid residue as an enzyme of (a).
- 8. An enzyme of which at least a portion is coded for by a polynucleotide of claim 1, and which is selected from the group consisting of:
 - (a) an enzyme comprising an amino acid sequence which is at least 70% identical to an amino acid sequence selected from the group of amino acid sequences set forth in SEQ ID NOS:15-28 or 61-64; and
 - (b) an enzyme which comprises at least 30 amino acid residues to the enzyme of (a).
- 9. A method for generating glucose from soluble cell oligosaccharides comprising contacting a sample containing oligosaccharides with an effective amount of an enyzme selected from the group consisting of an enzyme having the amino acid sequence set forth in SEQ ID NOS: 15-28, 61-63 and 64 such that glucose is produced.
- 10. The method of cliam 9, wherein the sample is selected from the group consisting of dairy products, fruit juices, detergents, textiles, guar gum, animal feed, plant biomass and waste products.
- The method of claim 9, wherein the oligosaccharide is selected from the group consisting of maltose, cellobiose, lactose, sucrose, raffinose, stachyose, verbascose, cellulose, starch, amylose, glycogen, disacharrides, polysacharrides and pullulan.



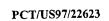


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Figure 1a.

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1441	PAG GIn	TAA	14	146	214	1111	1117.	1314	1111	Pro	λяμ	G1 _u	1,4-11	Gla	11 1 21	lant.	The	Lang	ATC 11c	1440 480
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Figure 1b(Continued)

OC1/4 GLYCOSIDASE - 31G/B COMPLETE GENE SEQUENCE - 9/95

GENE SEQUENCE - 9/95
ANY ATA AGA AGG TCC GAT TTT CCA AAA GAT TTT ATC TTC GGA ACG GCT ACG GCA GCA TAC 60
61 CAG ATT GAA GGT GCA GCA AAC GAA GAT GGC AGA GGG CCA TCA ATT TGG GAT GTC TTT TCA 120 C1 Gln Ile Glu Gly Ale Ale Agn Glu Agn Gly Arg Gly Pro Ser Ile Trp Agn Val Phe Ser 40
121 CAC ACC CCT GGC AAA ACC CCT
121 CAC ACC CCT GGC AAA ACC CTG AAC GGT GAC ACA GGA GAC GTT GCC TGT GAC CAT TAT CAC 180 181 CGA TAC AAC CAA GAA GAA GAA GAA GAA GAA GAA G
81 Ile Ser Trp Pro Arg Ile Met Pro Asp Gly Lys Asn Ile Asn Gln Lys Gly Val Asp Phe 100
301 TAC AAC AGA CTC GTT GAT GAG CTT TTG AAG AAT GAT ATC ATA CCA TTC GTA ACA CTC TAT 360 101 Tyr Asn Arg Leu Val Asp Glu Leu Leu Lys Asn Asp Ile Ile Pro Phe Val Thr Leu Tyr 120
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TOU ATT ACA CTC AAC CAA COL TOO TO
161 Trp Ile Thr Leu Arn Glu Pro Trp Cys Ser Ser Phe Ser Gly Tyr Tyr Thr Gly Glu His 180
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541 GCC CCG GGT CAT CAA AAT TTA CAA GAA GCG ATA ATC GCG GCG CAC AAC CTG TTG AGG GAA . 600 181 Ala Pro Gly His Gln Asn Leu Gln Glu Ala Ile Ile Ala Ala His Asn Leu Leu Arg Glu 200
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101 WAA GAA GCA CTT COA
781 GAA GAA GCA GTT GCA CTT TAT ACG GAA AAA GGG TTG CAA GTT CTC GAT AGC GAT ATG AAT 840 G1u Glu Glu Ala Val Ala Leu Tyr Thr Glu Lys Gly Leu Gln Val Leu Asp Ser Asp Het Asn 280
841 ATT ATT TOO ACT CO 120 ACT CO
841 ATT ATT TCG ACT CCT ATA GAC TTC TTT GGT GTG AAT TAT TAC ACA AGA ACA CTT GTT 900
The Law Val Val 200
THE GLY ASP LEU Pro Lvs Thr Chu 120
ATG GGA TGG GAA ATG TAG GOD TO THE
ASP NEC LEU VAL TYF Leu LVS Chu Arg. 340
TAT AAA CTA CCA CTT TAT AND
341 Tyr Lys Leu Pro Leu Tyr Ile Thr Glu Asn Gly Het Ala Gly Pro Asp Lys Leu Glu Asn 360
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1081 GGA AGA GTT CAT CAT AAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1081 GGA AGA GTT CAT CAT AAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1141 GAA GAA GT CAT GAT AAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1141 GAA GAA GT CAT GAT GAT GAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1141 GAA GAA GT CAT GAT GAT GAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1142 GAA GT CAT GAT GAT GAT GAT TAC CGA ATT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1142 GAA GT GAT GAT GAT GAT GAT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1143 GAA GT GAT GAT GAT GAT GAT GAA TAT TTG GAA AAG CAC TTT GAA AAA GCA CTT 1140 1144 GAA GT GAT GAT GAT GAT GAT GAT GAT GAT
out old by his phe Glu Lys Ala Leu 190
Ann Ann Ann
THE GAN TEG CCC TCC CCA TAIC TOO AND TOO
1201 CCA AAA AGG ATTA TETO AAA CAM TETO
the Lys City Phe Leu Lys Ser End 419

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STAPHYLOTHERMUS MARINUS GLYCOSIDASE - 12G COMPLETE GENE SEQUENCE 9/95

1 777 ABA ABA ABA
1 TTG ATA ACC TIT CCT GAT TAT TTC TTG TIT GGA ACA GGT AGA TGA TGG GAG CAG ATG GAG. 60
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TO CIV OUR TAT AIR COR BLA
181 CTG GGA TAT AAT CCT TAT AGG TTC TCC ATA GAG TGG AGT AGA ATA TTT CCC AGA AAA GAT 61 Leu Gly Tyr Asn Ala Tyr Arg Phe Ser Ile Glu Trp Ser Arg Ile Phe Pro Arg Lys Asp 80
241 CAT ATA CAT TAT CAC more and and an army the Pro Arg Lys Asp 80
241 CAT ATA GAT TAT GAG TCG CTT AAT AAG TAT AAG GAA ATA GTT AAT CTA CTT AGA AAA TAC 300 81 His Ile Asp Tyr Glu Ser Leu Asn Lys Tyr Lys Glu Ile Val Asn Leu Leu Arg Lys Tyr 100
301 GGG ATA GAA CCT GTA ATC ACT CTT CAC CAC TTC ACA AAC CCG CAA TGG TTT ATG AAA ATT 360
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121 AGG GGA GAA CTA CAA AGE GEO
721 AGG GGA GAA CTA GAA ACT CTC CGT GGA AAA TAC CGA GTT GAG CCC GGA AAT ATT GAT TTC 780 241 Arg Gly Glu Leu Glu Thr Leu Arg Gly Lys Tyr Arg Val Glu Pro Gly Asn Ile Asp Phe 260
781 ATA GGC ATA AAC TAT TAT TOO DO D
781 ATA GGC ATA AAC TAT TAT TCA TCA TAT ATT GTA AAA TAT ACT TGG AAT CCT TTT AAA CTA 840 261 Ile Gly Ile Asn Tyr Tyr Ser Ser Tyr Ile Val Lys Tyr Thr Trp Asn Pro Phe Lys Leu 280
841 CAT ATT 121 CTC C11 C11
841 CAT ATT ANA GTC GAA CCA TTA GAT ACA GGT CTA TGG ACA ACT ATG GGT TAC TGC ATA TAT 900 281 His Ile Lys Val Glu Pro Leu Asp Thr Gly Leu Trp Thr Thr Het Gly Tyr Cys Ile Tyr 300
THE THE GIV TVE CVE TIE THE
901 CCT AGA GGA ATA TAT GAA GTT GTA ATG AAA ACT CAT GAG AAA TAC GGC AAA GAA ATA ATC 960 101 Pro Arg Gly Ile Tyr Glu Val Val Net Lys Thr His Glu Lys Tyr Gly Lys Glu Ile Ile 120
The same of the same same same same same same same sam
961 ATT ACA GAG AAC GGT GTT GCA GTA GAA AAT GAT GAA TTA AGG ATT TTA TCC ATT ATC AGG 1020
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TOG AGE THE ATE GAT AAT THE CAS THE CASE THE CAS
of the Ash Gin Arg Phe Gly Leu Val 380
1141 CAA CTT GAT TAT AAG ACT TTT GAG AGA
of the control of the
ATA CCA CCT ACC AAG ACT ATA ACT ATA
401 The Ala Arg Thr Lys Thr The Ser Asp Glu Tyr Leu Glu Lys Tyr Gly Leu Lys Asp Leu 420
1201 GAA TAA 1266
421 Glu End 422

Figure 3

Thermococcis 9N2 Glydosidase -319/G Complete gene sequence 9/95

ATG CTA CCA GAA GGC TITE
ATG CTA CCA GAA GGC TTT CTC TGG GGC GTG TCC CAG TCC GGC TTT CAG TTC GAG ATG CGC 60 Het Leu Pro Glu Gly Phe Leu Trp Gly Val Ser Gin Sax Gly Phe Gln Phe Glu Het Gly 20
61 GAC AAG CTC AGG AGG AAC ATT GAT CUG AAC AUA GAC TGG TGG AAG TGG GTC AGG GAT CCC 120
181 GAA CTT TAC GAG AAG GAT CAC CGC CTC GCC AGA GAC CTC GCT CTG AAC GTT TAC AGG ATT 240 61 Glu Leu Tyr Glu Lys Asp XLs Arg Leu Als Arg Asp Leu Gly Leu Ash Val Tyr Arg Ile 80 241 GGA ATA GAG TRO 100
24: GGA ATA GAG TOTAL AND
81 GLY 11e GLU TEP Ser Arg 11e Phe Pro TEP FEG THE TEM PRO UN AND GLAG GTT GAG GTT GAG 300
141 Glu Leu Gly Phe Lys Val Ile Val Asn Leu Asn His Fhe Thr Leu Pro Leu Trp Leu His 160
481 GAT CCC ATA ATC CCC ACC CAC CAC CAC CTC ACC AAC GCT ACC ATT GCC TGG GTC GGC CAC 540 161 Asp Pro Ile Ile Ala Arg Clu Lys Ala Leu Thr Ann Cly Arg Ile Gly Trp Val Gly Gln 180
The same of the sa
541 GAG AGC GTG GTG GAG TTC GCC AAG TAC GCG GCG TAC ATC GCG AAC GCA CTC GGG GAC CTC 600 181 Glu Ser Val Val Glu Pho Ale Lys Tyr Ale Ale Tyr Ile Ale Asn Ale Leu Gly Asp Leu 200
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'** AM AIU ATA 110 COC OLD HO
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THE VALUE WAT ARE COME AND ADDRESS.
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281 Ala Tyr Pro Tyr Asp Ser Asn Asp Pro Lys Asp val Lys Ala Glu Asn Asp Asn Tyr 300
THE CAC AGE GGG CTC TO THE
301 TTC CAC AGE GGG CTC TTC GAC GCA ATC CAC AAG GGC AAG CTC AAC ATC GAG TTC GAC 360 301 Phe His Ser Gly Leu Phe Phe Asp Ala Ile His Lys Gly Lys Leu Asn Ile Glu Phe Asp 320
THE WAY LAY LAY LAY AND THE WAY AND THE WA
961 GGT GAG ACC TTC GTC AAR GTT CGG CAT CTC AGG GGG AAC GAC TGG ATA GGC GTT AAC TAC 1020
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CAC ACC CTG CGG CCG TAG TAG TAG TAG
1261 GAC ACC CTG CGG CCG TAC TAC CTC GCG AGC CAT GTA CGG AAG ATT GAG GAG GCG TAC GAG 1320 421 Asp Thr Leu Arg Pro Tyr Tyr Leu Ala Sur His Val Ala Lys Ils Glu Glu Ala Tyr Glu 440
440

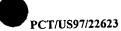
Figure 4a

6/46

WO 98/24799

321 441	VI.	61 ₁	T TAC	CA(GT(C ACC	C GG	: TX	C CT	TAC	î rec	GC	- CTC	ACC	GAC	: ***	TAC	: cuc	TCC	GCC Ala	138
) 6]	CLC.	CC1	*	٠											~ ~ ~	A.511		Clu	מוז	Ala	440
.01	Leu	Gly	? Phe) Azg	Het	Arg	Phe	GLy		TAT	Lys	GTG Val	GAT ABD	CTC	ATA	ACC	MG	CAC	AGA	Ala ACA Thr	1440
41	ccc	· ccc	CAC	·												THE	LYS	Glu	Arg	The	480
	•••	vr.A	CIU	GIT	Ser	Val	Lys	Val	Tyr	yra	Gly	Tiu	Val	GAG	AAC	AAC AAC	CCA	OIC.	LOC	MG Lys	1500
01	CAA	ATC	CCC	CAG	ANG	-10	CCA	CTT	ccc	TGA	15	30			_		ULY	ATT	ser	Ly:	500

Figure 4b(Continued)



	1	AT(GAA Glu	Ar Ar	iG Al	rc G	AT G			CTC Leu	TCT Scr	Chi				ACA Thr	GA		A AA				CTC I.eu	GTT Vat	60 20
	61 21	-	GGG Gly		T GC	ਜ c , l.e			GGA	CTT Leu	TTT Phc	GG(CAT His	TCC Ser	. AG	A (T)'(; GC	-		GCG Na	GCT	120
	21 41	GGA	GAA Glu	AC	'A CA	T CC	:c	7 (CA /	\GA	стт	GGA	AT	r co	.T (GCG	111	GT.	כדכ			•	GT	CCC	180
		•					, Va		TH /	-	Leu	Cly	lle 4.44	Pro		Ala 	Phe	Val		Λla	As,		Cly	Pro	60
	61 .	Ala	Gly	Leu	Arg	llc	Ası	1 2	ro T	hr	Arg	Glu	Asn	Ası	p C	Slu	Asa	Thr	Tyr	TA(Thr		pr CG	GCA Ala	240 80
	11 1	Phe	CCC Pro	GT: Val	GA, Glu	A AT	C AT	G C	TC C	CT :	TCT Ser	ACC Thr	TGG Trp				GAC Asp	CTT Leu	CTG Leu	GA A	GA. Glu	_	TG al	GGA Gly	300 100
30 10	1 1	-ys	GCC Ala	ATC Mei	GG/ Gly	GA. Glu	A GA Glu	A G	I A	GG (GAA Glu	TAC Tyr	GGT Gly	GT(Val			CTG CTG	CTT Leu	CTT Lev	GCA Ala	CCT Pro	G G	CG	ATG Met	360 120
36 12	I A	MC um	ATT ile	CAC His	AGA Arg	AA(Asn	Pro	CT	т то • Су	ज c			AAT Asn	TTC Phe			TAC Tyr	TAC Tyr	TCA Ser	GAA Glu	GAT Asp	CC Pro		GTC Val	420 140
42) 14)	_	77 °	TCC Ser	GGT Gly	GAA Glu	ATG Mei	GCT Ala	TC Ser	A GC				AAG Lys					TCT Ser	CAA Gla	GGG Gly	GTG Val	GG Gly		GCC Ala	480 160
48 I 16 I	- C	GC A	ITA .	AAA Lys	CAC His	TTT Phe	GTC Vai	GC: Ala	۸۸ G ۸۶۱	C A.	AC (GAA Giu				ns rea		GTA Val	GTG Val	GAC Asp	AC Thr		ATC	540 180
541 181	ت ۷	TG T	et (GAG Glu	CGA Arg	GCC Ala	CTC Leu	AG/ Arg	Glu	A A	ΓΑ Τ Τ	AT I	CTG Lev	AAA Lys	GC Gly			GAA Giu	ATT lic	GCT Ala	GTC Val	AA(-	.ys	600 200
60 I 20 I	GC Ali	A A	GA (cc TCC	TGG Trp	ACC Thr	GTG Val	ATC Met	A G	G GC	T T	AC /	AAC Asn	AAA Lys	CT(G A		GGA Gly		TAC Tyr	TGT Cys	TCA Ser		AG iin	660 220
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721 241	A G Ser	ic G	AC T	de .	TAC Tyr	GCG Ala	GGA Gly	GAC Asp	AA (CC Pro	T G	TA C		CAG Gin	CTC	٠.	AG (cc	GGA Gly	AAC Aan	GAT	ATG		тс	780 260
781 261	AT(G CO	т с • G	GG /	.ys	GCG Ala	TAT Tyr	CAG Gla	GT C	AA Asn	C A				AGA	. G	NT C	GAA	ATA	GAA	Asp GAA	McI	A.	TG	840
841 231	GA (G GC	C T	•	ug	GAG	GGA	•		ΑG	T G/	NG G	AG (717	CTC	G/	\т с	AG '	TGT	GTG	Glu AGA	lle AAC	M A	т	280 900
901					-		AAC				(C)				Leu				•	Vat AAC	Arg AAG	Asn	Ite Ge		300 960
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371	Leu	Glu	A TO	r A	AC is	GCG Ala	GAA Glu	CTC Val	GCC Ala	TAC Tyr	Gh Gh	A G			GCG Als	GA					CTT Lev	CTT Lev	GA GIO		020 340
1021 341	AAC Asn	AA S	C GO	π c y v	17 (11 ts	.Eu	CCG	TTC Phe	GAT Asp	GAA Glu	AA A	T AC			GTC Vai	CC	C G				ACC Thr	GGT Gly	CA Gli		080 060
1881 361	ATC lie	GA, Glu	A AC	r Ik	ΓΑ / : 1	AAG (GGA (GGA . Gly	ACG Thr	GGA Gly	A G Ser	T GC			ACC Thr		T C	ca A	GA 1	rac	ACG	ATC	TC	τ 1	140
1141	ATC lie			A G	ge A	TA /		JAA .	AGA	AAC	AT	G AA	KG 17	T(' (GAC	GA.	A G	AA C	-	•	Thr TCC	NCT	Ser		180 (OO
			()10	, (,)	y í	ic I	.ys (, UI	Arg.	Asn	Me	ı l.y	· Pi	ne /	Asp	Glu	Gi	lu L	eu /	Nia .	Sei	The	Tyr	•	100

Figure:.5a

1201 GAG GAG TAC ATA AAA AAG ATG AGA GAA ACA GAG GAA TAT AAA CCC AGA ACC GAC 401 Glu Glu Tyr He Lyx Lyx Mer Arg Glu Thr Glu Glu Tyr Lys Pro Arg Tr T TCC Asp Ser Τm 420 1261 GGA ACG GTC ATA ANA CCG ANA CTC CCA GAG ANT TTC CTC TCA GAA ANA GAG 421 Gly The Val lie Lya Pro Lys Leu Pro Giu Asa Phe Leu Ser ATA AAG 444 1320 Ciu Lyx He Lys 440 1321 CCT CCA AAG AAA AAC GAT GTT GCA GTT GTT GTG ATC AGT AGG ATC TCC 441 Pro Pro Lys Lya Asn Asp Val Ala Val Val Val lic Ser Arg lic CCT GAG GGA TAC 1380 Cly Clu Cly Tyr 460 1381 GAC AGA AAG CCG GTG AAA GGT GAC TTC TAC CTC TCC GAT GAC GAG CTG 461 Asp Arg Lys Pro Val Lys Gly Asp Phe Tyr Leu Ser Asp Asp GAA CTC ATA AAA 1440 Giu Leu Giu Leu Lys 480 1441 ACC GTC TCG AAA GAA TTC CAC GAT CAG GGT AAG AAA GTT GTG GTT CTT 481 Thr Val Ser Lys Glu Phe His Asp Gln Gly Lys Lys Val Val CTG AAC ATC GGA 1500 Leu Asn lic Gly 500 1301 AGT CCC ATC GAA GTC GCA AGC TGG AGA GAC CTT GTG GAT GGA ATT CTT 501 Ser Pro Ile Glu Val Ala Ser Trp Arg Asp Leu Val Asp Gly Ile CTC TGG CAG ٧ai Τm Gin 1561 GCG GGA CAG GAG ATG GGA AGA ATA GTG GCC GAT GTT CTT GTG GGA AAG 521 Ala Gly Gin Glu Met Gly Arg lie Val Ala Asp Val Leu Val Gly Lya ATT AAT CCC TCC 1620 Pro Scr 540 1621 GGA AAA CTT CCA ACG ACC TTC CCG AAG GAT TAC TCG GAC GTT CCA TCC 541 Gly Lys Leu Pro Thr Thr Phe Pro Lys Amp Tyr Ser Amp Val Pro Ser TGG ACG TTC CCA 1680 Trp Thr Pro 560 1681 GGA GAG CCA AAG GAC AAT CCG CAA AGA GTG GTG TAC GAG GAA GAC ATC 561. Gly Glu Pro Lys Asp Asn Pro Gin Arg Val Val Tyr Glu Glu Asp lic TAC GTG GGA 1740 Tyr Vat , Tyr 1741 AGG TAC TAC GAC ACC TTC GGT GTG GAA CCT GCC TAC GAA TTC GGC TAC Arg Tyr Tyr Asp Thr Phe Gly Val Glu Pro Ala Tyr Glu Phe Gly Tyr GGC CTC TCT TAC 1800 Gly 600 ىما 1301 ACA ANG TIT GAN TAC ANA GAT TIA ANA ATC GCT ATC GAC GGT GAG ACG The Lys Phe Giu Tyr Lys Asp Leu Lys lie Als lie Asp Gly Giu CTC AGA CTG TCG 1860 Arg V٠ Ser 620 1861 TAC ACG ATC ACA AAC ACT GGG GAC AGA GCT GGA AAG GAA GTC TCA CAG Tyr Thr lie Thr Asn Thr Gly Asp Arg Ala Gly Lys Glu Val Ser στc TAC ATC 1920 *** Val Tyr 640 Lys 1921 GCT CCA AMA GGA AMA ATA GAC AMA CCC TTC CAG GAG CTG AMA GCG TTT 641 Ala Pro Lys Gly Lys ilc Asp Lys Pro Phe Gin Giu Leu Lys CAC ACA IORA His Lys Thr 660 Lys 1981 CTT TTG AAC CCG GGT GAA TCA GAA GAA ATC TCC TTG GAA ATT CCT CTC 661 Leu Leu Ain Pro Gly Glu Ser Glu Glu Ile Ser Leu Glu Ile AGA GAT \Box GCG 2040 Pro Arg Αso Leu Ala 680 2041 AGT TTC GAT GGG AAA GAA TGG GTT GTC GAG TCA GGA GAA TAC GAG GTC 681 Ser Phe Asp Gly Lys Glu Trp Val Val Glu Ser AGG CTC COT GCA 2100 Gly Glu Tyr Glu Arg Val Ciy Ala 700 2101 TCT TCG AGG GAT ATA AGG TTG AGA GAT ATT TTT CTG GTT GAG GGA GAG AAG 701 Ser Ser Arg Asp lie Arg Leu Arg Asp lie Phe Leu Val Glu Gly Glu AGA Arg L, y s 2161 CCA TGA 2166 721 Pro End 722

Figure 56(Continued)

PCT/US97/22623

20

1140 380

400

1260 420

1320 440

THERMOCOCCUS AEDII12RA GLYCOSIDASE (188/C) COMPLETE GENE SEQUENCE - 9/95 ATG ATC CAC TGC CGG GTT AAA GGG ATT ATA TGT GAG GCT CGC GGC ATA ACC ATC ACA ATA Het Ile His Cys Pro Vel Lys Gly Ile Ile Ser Glu Ala Arg Gly Ile Thr Ile Thr Ile GAT TTA AGT TIT CAA GGC CAA ATA MAT AAT TTG GTG AAT GCT ATG ATT GTC TIT CCG GAG Asp Leu Ser Phe Gin Cly Gin Ile Asn Asn Leu Val Asn Ala Met Ile Val Phe Pro Giu 120 40 121 THE THE CHE THE GGA ACC GET ACA TET THE CAT CAG ATE GAG GGA GAT AND THE AND 41 Phe Phe Leu Phe Gly Thr Ala Thr Ser Ser His Gln Ile Glu Gly Asp Asn Lys Trp Asn 180 60 181 GAC TOG TGG TAT TAT GAG GAG ATA COT ANG CTC CCC TAC ANA TCC GGT ANA GCC TGC ANT Asp Trp Trp Tyr Tyr Glu Glu Ile Gly Lys Leu Pro Tyr Lys Ser Gly Lys Ala Cys Asn 240 241 CAC TOG GAG CTT TAC AGG GAA GAT ATA GAG CTA ATG GCA CAG CTC GGC TAC AAT GCC TAC 81 His Trp Glu Leu Tyr Arg Glu Asp Ile Glu Leu Het Ala Gln Leu Gly Tyr Asn Ala Tyr 300 CGC TIT TCG ATA GAG TCG AGC CGT CTC TTC CCG GAA GAG GGC AAA TTC AAT GAA GAA GCC 100 301 Arg Phe Ser Ile Glu Trp Ser Arg Leu Phe Pro Glu Glu Gly Lys Phe Asn Glu Glu Ala 360 161 TTC AAC CGC TAC CGT GAA ATA ATT GAA ATC CTC CTT GAG AAG GGG ATT ACT CCA AAC GTT 120 Phe Asn Arg Tyr Arg Glu Ile Ile Glu Ile Leu Leu Glu Lys Gly Ile Thr Pro Asn Val 140 ACA CTG CAC CAC TTC ACA TCA CCG CTG TGG TTC ATG CGG AAG GGA GGC TTT TTG AAG GAA Thr Leu His His Phe Thr Ser Pro Leu Trp Phe Het Arg Lys Gly Gly Phe Leu Lys Glu 480 150 GAA AAC CTC AAG TAC TGG GAG CAG TAC GTT GAT AAA GCC GCG GAG CTC CTC AAG GGA GTC Glu Asn Leu Lys Tyr Trp Glu Gln Tyr Val Asp Lys Ala Ala Glu Leu Leu Lys Gly Val 541 ANG CIT GTA GCT ACA TTC AND GAG CCG ATG GTC TAT GTT ATG ATG GGC TAC CTC ACA GCC 181 Lys Leu Val Ala Thr Phe Asn Glu Pro Het Val Tyr Val Het Het Gly Tyr Leu Thr Ala 600 200 501 THE TOG CCG CCC TTC ATC ANG AGT CCC TIT ANA GCC TIT ANA GTT GCC GCA ANC CTC CTT Tyr Trp Pro Pro Phe Ile Lys Ser Pro Phe Lys Ala Phe Lys Val Ala Ala Asn Leu Leu 201 660 220 661 AMG GCC CAT GCA ATG GCA TAT GAT ATC CTC CAT GGT AAC TIT GAT GTG GGG ATA GTT AAA Lys Ala His Ala Het Ala Tyr Asp Ile Leu His Gly Asn Phe Asp Val Gly Ile Val Lys 720 AND ATO COD ATA ATG CTC COT GOA AGO AND AGA GAG ANA GAC GTA GAA GOT GOO CAA AAG Asn Ile Pro Ile Met Leu Pro Ala Ser Asn Arg Glu Lys Asp Val Glu Ala Ala Gln Lys 780 260 781 GCG GAT AND CTC TIT AND TGG AND TTC CTT GAT GCA ATA TGG AGC GGA ANA TAT ANA GGA 261 Ale Asp Asn Leu Phe Asn Trp Asn Phe Leu Asp Ale Ile Trp Ser Gly Lys Tyr Lys Gly GCT TIT GGA ACT TAC ANA ACT CCA GAA AGC GAT GCA GAC TIC ATA GGG ATA AAC TAC TAC Als Phe Gly Thr Tyr Lys Thr Pro Glu Ser Asp Als Asp Phe Ile Gly Ile Asn Tyr Tyr 281 900 300 901 ACA GCC AGC GAG GTA AGG CAT AGC TGG AAT CCG CTA AAG TIT TTC TTC GAT GCC AAG CTT Thr Ala Ser Glu Val Arg His Ser Trp Asn Pro Leu Lys Phe Phe Phe Asp Ala Lys Leu 320 961 GCA GAC TTA AGC GAG AGA AAA ACA GAT ATG GGT TGG AGT GTC TAT CCA AAG GGC ATA TAC 321 Ale Asp Leu Ser Glu Arg Lys Thr Asp Het Gly Trp Ser Val Tyr Pro Lys Gly Ile Tyr 1020 340 1021 GAN GCT ATA GCA ANG GTT TON CAC THE GGN ANG CON ATG THE ATC ACG GAN AND GGG ATA 141 Glu Ala Ile Ala Lys Val Ser His Tyr Gly Lys Pro Het Tyr Ile Thr Glu Asn Gly Ile 360 1081 GCT ACC TTA GAC GAT GAG TGG AGG ATA GAG TTT ATC ATC CAG CAC CTC CAG TAC GTT CAC

Phe Lys Arg Arg Pro Arg Lys Ser Ala Tyr Ile Tyr Gly Gly Ile Ala Arg Gly Lys Lys 1321 ATA AAA GAC GAA CTG CTG GCA AAG TAT GGG CTT CCG GAG CTA TGA 441 The Lys Asp Glu Leu Leu Ala Lys Tyr Gly Leu Pro Glu Leu End

Figure 6

361 Ala Thr Leu Asp Asp Glu Trp Arg Ile Glu Phe Ile Ile Gln His Leu Gln Tyr Val His

1141 AMA GCC TTA AAC GAT GGC TTT GAC TTG AGA GGC TAC TTC TAT TGG TCT TTT ATG GAT AAC 181 Lys Ala Leu Asn Asp Gly Phe Asp Leu Arg Gly Tyr Phe Tyr Trp Ser Phe Het Asp Asn

1201 TTC GAG TGG GCT GAG GCT TTT AGA CCA CGC TTT GGG CTG GTC GAG GTG GAC TAC ACG ACC Phe Glu Trp Ala Glu Gly Phe Arg Pro Arg Phe Gly Leu Val Glu Val Asp Tyr Thr Thr

1261 TTC AAG AGG AGA CCG AGA AAG AGT GCT TAC ATA TAT GGA GAA ATT GCA AGG GAA AAG AAA

10/46

THERMOCOCCUS CHITONOPHAGUS GLYCOSIDASE - 22G COMPLETE SEQUENCE - 9/95

7 - 77/35	
1 TTC CTT CCA GAG AAC TTT CTC TGG GGA GTT TCA CAG TCC GGA TTC CAG TTT GAA ATG GGG 1 Het Leu Pro Glu Asn Phe Leu Trp Gly Val Ser Gln Ser Gly Phe Gln Phe Glu Het Gly	
61 GAC AGA CTG AGG AGG CAC ARM GAT	
Asp itp Trp Tyr Trp Val Arg Asp Ct.	
121 TAT AAT ATC AAA AAA GGA CTA GTA AGT GGG GAT CTT CCC GAA GAC GGT ATA AAT TCA TAT 41 Tyr Asn lle Lys Lys Gly Leu Val Ser Gly Asp Leu Pro Glu Asp Gly Ile Asn Ser Tyr	
181 GAA TTA TAT CAG ACA CAG CAA	60
181 GAA TTA TAT GAG AGA GAC CAA GAA ATT GCA AAG GAT TTA GGG CTC AAC ACA TAT AGG ATC 61 Glu Leu Tyr Glu Arg Asp Gln Glu Ile Ala Lys Asp Leu Gly Leu Asn Thr Tyr Arg Ile	240 80
491 GGA ATT GAA TCC ACC ACC ACC	
THE VAL ASD VAL CITY TO STATE OF	300 100
301 ATT GAT GAG TCT TAC GGG TTG GTA AAG GAT GTG AAG ATT TCT AAA GAC GCA TTA GAA AAA 101 Ile Asp Glu Ser Tyr Gly Leu Val Lys Asp Val Lys Ile Ser Lys Asp Ala Leu Glu Lys	360
JOI CTT GAT GAA ATC CCT AND GAA ACC	120
361 CTT GAT GAA ATC GCT AAC CAA AGG GAA ATA ATA TAT TAT AGG AAC CTA ATA AAT TCC CTA 121 Leu Asp Glu Ile Ala Asn Gln Arg Glu Ile Ile Tyr Tyr Arg Asn Leu Ile Asn Ser Leu	420 140
421 AGA AAG AGG GGT TTT AAG GTA AGA GTA	
and had his pie Thr Leu Pro Ile Tro Leu	480 160
481 CAT GAT CCT ATC G14 TCT 101 T11 101 T11	
161 His Asp Pro Ile Glu Ser Arg Glu Lys Ala Leu Thr Asn Lys Arg Asn Gly Trp Val Ser	540 180
541 GAA AGG AGT GTT ATA GAG GTT AGA ANA	100
181 Glu Arg Ser Val Ile Glu Phe Ala Lys Phe Ala Ala Tyr Leu Ala Tyr Lys Phe Gly Asp	600 200
601 ATA GTA GAC ATG TGG ACC ACA TOTAL AND ALL COLORS	200
	660
661 GCC CCA TAC TCA GCA TTC CCC CCC CCA CCA CCA CCC	220
The same of the sa	720 240
721 CTA CAT ATG ATA AAC GCC CAT CCT TTA CCC TATA	
All Det Ale lyr Arg Het Ile Lys Lys Phe Asp Arg Lys	780 260
781 AAA GCT GAT CCA GAA TCA AAA GAA CCA GCT GAA ATA GGA ATT ATA TAC AAT AAC ATC GGC 8 261 Lys Ala Asp Pro Glu Ser Lys Glu Pro Ala Clu Tla Clu	340
2 of the Gly 12 of the Gly 12 of the Tyr Asn Asn Ile Gly 2	80
841 GTC ACA TAT CCG TTT AAT CCG AAA GAC TCA AAG GAT CTA CAA GCA TCC GAT AAT GCC AAT 9	00
Asp Set Bys Asp Leu Gin Ala Ser Asp Asn Ala Asn]	00
901 TTC TTC CAC AGT GGG CTA TTC TTA ACG GCT ATC CAC AGG GGA AAA TTA AAT ATC GAA TTT 9	60
The Led The Ald The His Arg Gly Lys Leu Asn Ile Glu Phe 3	20
961 GAC GGA GAG ACA TIT GIT TAC CIT CCA TAT ITA AAG GGC AAT GAT TGG CTG GGA GTG AAT 121 ASP Gly Glu Thr Phe Val Tyr Lau Pro Ty	020
of the life Lea Lys Gly Asn Asp Trp Lea Gly Val Asn 3.	40
1021 TAT TAT ACA AGA GAA GTC GTT AAA TAC CAA GAT CCC ATG TTT CCA AGT ATC CCT CTC ATA 11	080
The strands are the Pro Ser Ile Pro Leu Ile 3	60
1081 AGC TTC AAG GGC GTT CCA GAT TAT GGA TAC GGA TGT AGA CCA GGA ACG ACG TCA AAG GAC 11361 Ser Phe Lys Gly Val Pro Asp Tyr Gly Tyr Gly Tyr Asp Tyr Gly Tyr Gly Tyr Asp Tyr Gly	140
of the control of the	10
1141 CGT AAT CCT GTT AGT GAC ATT CGA TGG GAG TO THE	200
The Gld was Tyr Pro Lys Gly Met Tyr Asp Ser Ile 40	
1201 GTA GCT GCC AAT GAA TAT GCA CTT GCT GTA GTA GTA	
401 Val Ala Ala Asn Glu Tyr Gly Val Pro Val Tyr Val Thr Glu Asn Gly Ile Ala Asp Ser 42	160 10
1261 AAA GAT GTA TTA AGG CCC TAT TAG ATG CCA TOT CLG	
421 Lys Asp Val Leu Arg Pro Tyr Tyr Ile Ala Ser His Ile Glu Ala Het Glu Glu Ala Tyr 44	20

Figure 7a

WO 98/24799

PCT/US97/22623

441	Glu	AAT Asn	Gly	TAT	Asp	Val	Arg Arg	GIA	Tyr	: TT/	CAC Him	TGG Trp	GCA Ala	TTA Leu	ACC Thr	GAT Amp	AAT Asn	TAC	GVV	TGG Trp	1 180 460
1381	CCC .	TTA	GGC	TTC	AGA	ATC	400	-				_									1440
1441 481	AAA (CCC	AGG	**	AAG	ACT	CTA	ACA	~	-											1500
	AGC A	uc	ATC	ACC	***	CAG	ATC	TTE	~.~					36							- · · ·

Figure 7b(Continued)



PYROCOCCUS FURIOSUS GLICOSIDASE - 7G1 COMPLETE GENE SEQUENCE - 10/95

COMPLETE GENE SEQUENCE - 10/95	
ATG TTC COT CAR As a	
1 Met Phe Pro Glu Lys Phe Leu Trp Gly Val Ala Gln Ser Gly Phe Gln Phe Glu Met Gly 61 GAT AAA CTC AGG AGG AAT ATT COS AST AND COS	
bys the Leu Trp Gly Val Ala Gin Ser Gly Pho Gly Law ATG GGG	60
61 GAT ANA CTC ACC ACC	20
21 ASP LYS LEG AND AND GOO CITY CITY AND THE ASP TEP TEP HIS TEP VAL AND AND LYS	• •
and Arg Asn Ile Asp The Asn Th	120
121 ACA AAT ATA GAG AAA GGC CTC GTT AGT GGA GAT CTT CCC GAG GAG GGG ATT AAC AAT TAC	120
AL ATA GAG AAA GGC CTC GTT AGT GGA GAT GGT	40
41 Thr Ash Ile Glu Lye Gly Leu Val Ser Gly Asp Leu Pro Glu Gly Ile Ash Ash Tac 181 GAG CTT TAT GAG AAG GAC CAT GAG ASP CEU Pro Glu Gly Ile Ash Ash Tyr	
der Gly Asp Leu Pro Glu Gly Ila Ann IAC	180
181 GAG CTT TAT GAG AAG GAC CAT GAG ATT GCA AGA AAG CTG GGT CTT AAT GCT TAC AGA ATA 61 Glu Leu Tyr Glu Lys Asp His Glu Ile Ala Arg Lys Leu Gly Leu Asp Ala ATA	60
61 Glu Leu Tyr Glu Lys Asp His Glu Ile Ala Arg Lys Leu Gly Leu Ash Ala Tyr Arg Ile 241 GGC ATA GAG TGG AGG AGA ATA TTC GGC TTS TTC AGG TGG TGT AND Ala Tyr Arg Ile	
AND ALS DIE THE ALE AND LOUGH THE TAC AGA ATA	240
741 CCC NEW COLO	80
81 GIV II. GIV TON CON ATA TTC CCA TGG CCA ACG ACA TTT ATT	
The Ser Arg He Phe Pro Trp Pro The The The CAT CIT GAT TAT AGC	300
301 Tam sam can and Tur Ser	
301 TAT AAT GAA TCA TAT AAC CTT ATA GAA GAT GTA AAG ATC ACC AAG GAC ACT TTG GAG GAG 101 Tyr Ash Glu Ser Tyr Ash Leu Ile Glu Asp Val Lys Ile Thr Lys Asp Thr Leu Glu Glu 361 TTA GAT GAG ATC GCC AAC AAG ACC GAG GAG	100
TYP ASH GIU Ser Tyr Ash Leu Ile Gly Ash Val ANG AIC ACC AAG GAC ACT TTG GAC CAC	
261 may only the Charles of the Char	360
JOI TTA GAT GAG AIC GCC AAC ANG AGG GAG GAG	150
361 TTA GAT GAG ATC GCC AAC AAG AGG GAG GTC GCC TAC TAT AGG TCA GTC ATA AAC AGC CTG 121 Leu Asp Glu Ile Ala Asn Lys Arg Glu Val Ala Tyr Tyr Arg Ser Val Ile Asn Ser Leu 421 AGG AGG AAG GGG TTT AAG CTG AND CTG	
ALL AGC CTG	120
421 AGG AGG AAG GGG TTT AAG GTT ATA GTT AAT CTA AAT CAC TTC ACC CTT CCA TAT TGG TTG 41 ATG Ser Lys Gly Phe Lys Val Ile Val Am Lou Am Mis Phe The Lou Par Tag TTG 4	140
AND AND AND GOG TIT AND GIT ATA GIT ANT CTA ANT CAC TITC ACC CIT CCA TAT TGG TITG ASS CAT GAT CCC ATT GAG GCT ACC CIT AND Leu Ann His Phe Thr Leu Pro Tyr Trp Leu 1	
THE DAY WAL THE WAL AND DRU AND UNA BAR MED CAT COA TAT TGG TTG	80
48: CAT GAT CCC ATT GAG GCT AGG GAG AGG GCG TTA ACT AAT AAG AGG AAC GGC TGG GTT AAC 5	60
161 Kis Asp Pro Ile Glu Ala Arg Glu Arg Ala Leu Thr Asn Lys Arg Asn Gly Trp Val Asn 1	
AND AND PRO ITS GIV ALS ARE GIV ARE ALL AND AND AND AND GOT TOO GIT AND	40
541 CCA AGE AGE COM TOTAL THE SECRET ASE LYS AND ASE GIV TOP Val Ase	
541 CCA AGA ACA GTT ATA GAG TTT GCA AAG TAT GCC GCT TAC ATA GCC TAT AAG TTT GGA GAT 6181 Pro Arg Thr Val lie Glu Phe Ala Lys Tyr Ala Ala Tyr Ile Ala Tyr TAG TTT GGA GAT 6	80
FOO Arg Thr Val lie Giu Phe Ala Lue Mus GCC GCT TAC ATA GCC TAT AAG TTT GCD GTT	
181 Pro Arg Thr Val 11e Glu Phe Ala Lys Tyr Ala Ala Tyr 11e Ala Tyr Lys Phe Gly Asp 20	00
601 ATA GTG GAT ATG TGG AGC ACC TOTAL AND	00
601 ATA GTG GAT ATG TGG AGC ACG TTT AAT GAG CCT ATG GTG GTT GTT GAG CTT GGC TAC CTA GGC TAC CTA GTG GTG GTT GTT GAG CTT GGC TAC CTA GGC TAC CTA GTG GTG GTG GTG GTG GTG GTG GTG GTG G	
THE PAGE AST GLU PRO MET VAL VAL CITY GGC TAC CTA 66	50
661 GCC CCC The men and the Gly Tyr Leu 22	20
221 Ala Pro Tyr Ser Gly Phe Pro Pro Gly Val Leu Asn Pro Glu Ala Ala Lys Leu Ala Ile 24	
ory one pro Pro Gly Val Leu Ash Pro Glu Bla AAG CTG GCG ATA 72	0
721 CTT CAC AND ADD ADD ADD ADD ADD ADD ADD ADD ADD	
241 Lau His ALA AAT GCA CAT GCT TTA GCT TAT AGG CAG ATTA TAG	
721 CTT CAC ATG ATA AAT GCA CAT GCT TTA GCT TAT AGG CAG ATA AAG AAG TTT GAC ACT GAG 78 241 Leu His Het Ile Asn Als His Als Leu Als Tyr Arg Gln Ile Lys Lys Phe Asp Thr Glu 26 781 AAA GCT GAT AAG GAT TCT his side and also the control of the control	٥
781 AAA GCT GAT DAG CON DOOR TO THE LYS LYS Phe Asp Thr Glu 26	
781 AAA GCT GAT AAG GAT TCT AAA GAG CCT GCA GAA GTT GGT ATA ATT TAC AAC AAC ATT GGA 84	•
261 Lys Ala Asp Lys Asp Ser Lys Glu Pro Ala Glu Val Gly Ile Ile Tyr Asn Asn Ile Gly 28 841 GTT GGT TAT GGC ANG GREAT GGA 84	_
At the Art GGR 84	
841 GTT GCT TAT CCC AAG GAT CCG AAC GAT TCC AAG GAT GTT AAG GCA GCA GAA AAC GAC AAC 900	0
261 Val Ala Tyr Pro Lys Asp Pro Asn Asp Ser Lys Asp Val Lys Ala Ala Glu Asn Asp Asn 300	
AND AND SET LYS ASP Val LYS ALL CAN ARC GAC ARC 900	0
901 TTC TTC CAC TCA GGG CTG TTC TTC GAG GCC ATA CAC AAA GGA AAA CTT AAT ATA GAG TTT 960	כ
301 Phe Phe His san GUG CIG TTC TTC GAG GCC ATA CAC ARA GGS ARA GGS	
Ser Gly Leu Phe Phe Glu Ala Ile His Lvs Gly Iva ATA ATA GAG TTT 960)
301 Phe Phe His Ser Gly Leu Phe Phe Glu Ala Ile His Lys Gly Lys Leu Asn Ile Glu Phe 961 GAC GGT GAA ACG TTT ATA CAR GAG TTT 960	
961 GAC GGT GAA ACG TTT ATA GAT GCC CCC TAT CTA AAG GGC AAT GAC TGG ATA GGG GTT AAT 102 321 Asp Gly Glu Thr Phe 11e Asp Ala Pro Tyr Leu Lys Gly Asp Asp Tro	•
The big Giu The Phe Ile Asp Ala Pro Tur Lon Add GGC AAT GAC TGG ATA GGG GTT AAT 100	
321 Asp Gly Glu Thr Phe Ile Asp Ala Pro Tyr Leu Lys Gly Asn Asp Trp Ile Gly Val Asn 340	
1021 TAC TAC ACA AGG GAA GTA GTT ACG TAT CAG GAA CCA ATG TTT CCT TCA ATC CCG CTG ATC 108	,
TYP TYP The Arg Glu Val Val The Ton CAS GAR CCA ATG TTT CCT TCR ATG CCT	_
341 Tyr Tyr Thr Arg Glu Val Val Thr Tyr Gln Glu Pro Het Phe Pro Ser Ile Pro Leu Ile 360	
1081 ACC TIT AAG GOA GIT CAR GO TID GO	ı
361 Thr Phe Lys Gly Val Gla GGA TAT GCC TAT GCC TGC AGA CCT GGA ACT GTG	
1081 ACC TTT AAG GGA GTT CAA GGA TAT GGC TAT GCC TGC AGA CCT GGA ACT CTG TCA AAG GAT 114	0
1141 GAC AGA CCC CTC AGG PLA 380 380	
381 ASB ACC SIC AGC GAC ATA GGA TGG GAA CTC TAT CCA CAC	
And Ard Pro Val Ser Asp Ile Gly Trp Glu Lau TVI CAA GAG GGG ATG TAC GAT TCA ATA 120	С
381 Asp Arg Pro Val Ser Asp lie Gly Trp Glu Leu Tyr Pro Glu Gly Met Tyr Asp Ser lie 400	-
1201 GTT GAR GCT CAC AAG TAC GGC GTT CCA GTT TAC GTG ACG GAG AAC GGA ATA GCG GAT TCA 1261 401 Val Glu Ala His Lys Tyr Gly Val Pro Val Tyr Val Thr Glu ATA GCG GAT TCA 1261	
401 Val Glu Ala His Lys Tyr Gly Val Pro Val Tyr Val Thr Glu Ash Gly Ile Ala Asp Ser 420	^
tal Fro Val Tyr Val Thr Glu Ash Glv Tla Al-	
420 All Gly Ite Ala Amp Ser 420	

Figure 8a

1261 421	-	•			,		. , .	. , .	* * *	~14	261	u13	TIG	Lys	Met	lle	Clu	Lys	Ala	TTY Phe	1320 440
1321	G A G Glu	GAT Asp	GCG	TAI	GAA Glu	GIT Val	Lys	CGC GGC	TAC Tyr	TTC Phe	KT 2 CYC	TGG Trp	GCA Ala	TTA Leu	ACT The	و د/	AAC Aan	TTC Phe	GAG Glu	TGG	1380
1381 461			•		,		~9	1114	GI y	red	LAE	CIU	AT 1	ya U	Leu	Ile	Thr	Lys	Glu	λra	1440
1441 481			-		•						AL Y	GAG Glu	ATA Ile	GTA Val	GCC GCC	AAT neA	AAT Asn	GGT Gly	GTT Val	ACG Th:	1500
1501 501	AAA	AAG	ATT	GAA	GAG	GRA	~~~	~~~				15 51	33								

Figure 8b(Continued)

Bankia gouldi endoglucamase (37071)

(3/024)
9 18 27 36 45 54
ATG AGA ATA CGT TTA GCG ACG CTC GCG CTC TCC CGA CGG
Met Arg Ile Arg Leu Ala Thr Leu Ala Leu Cys Ala Ala Leu Ser Pro Val Thr
63
TIT GCA GAT AAT GTA ACC GTA CAA ATC GAC GCC GAC GGC GGT AAA AAA CTC ATC
Phe Ala Asp Asn Val Thr Val Gln Ile Asp Ala Asp Cly Cly Lys Lys Leu Ile
117 126 135 144 153
AGC CGA GCC CTT TAC GGC ATC AND AND TOO AND COLORS 153 162
Ser Arg Ala Leu Tyr Gly Met Asn Asn Ser Asn Ala Glu Ser Leu Thr Asp Thr
•••
171 180 189 198 207 216
GAC TGG CAG CGT TTT CGC GAT GCA GGT GTG CGC GTG CGG GAA AAT GGC GGC Asp Trp Gln Arg Phe Arg Asp Ala Clv Vol Arg Atg CTG CGG GAA AAT GGC GGC
Asp Trp Gln Arg Phe Arg Asp Ala Gly Val Arg Met Leu Arg Glu Asn Gly Gly
225 234 243 252
AAC AAC AGC ACC AAA TAT AAC TOG CAA CTTC CAG CTTC ACC ACC ACC ACC AAA TAT AAC TOG CAA CTTC CAG CTTC ACC ACC ACC ACC ACC ACC ACC ACC AC
Asn Asn Ser Thr Lys Tyr Asn Trp Gln Leu His Leu Ser Ser His Pro Asp Trp
279 288 297 306 315 324
and and the the eee feet life life and the are
Tyr Asn Asn Val Tyr Ala Gly Asn Asn Asn Trp Asp Asn Arg Val Ala Leu Ile
333 342 351 360
CAG GAA AAC CTG CCC CCC CAC ACC ATT TOC CCC CAC ACC ACC ATT TOC CCC CAC ACC ACC ACC ACC ACC ACC ACC A
Gln Glu Asn Leu Pro Gly Ala Asp Thr Met Trp Ala Phe Gln Leu Ile Gly Lys
• • •
387 396 405 414 423 432
GTC GCG GCG ACT TCT GCC TAC AAC TTT AAC GAT TGG GAA TTC AAC CAG TCG CAA
Val Ala Ala Thr Ser Ala Tyr Asn Phe Asn Asp Tro Glu Phe Asn Gln Ser Gln
441 450 459 468 477 496
TGG TGG ACC GGC GTC GCT CAG ANT CTC GCT GGG GGG GGG
Trp Trp Thr Gly Val Ala Gln Asn Leu Ala Gly Gly Gly Glu Pro Asn Leu Asp
ADE
495 504 513 522 531 540
GGC GGC GGA GCG CTG GTT GAA GGA GAC CCC AAT CTC TAC CTC ATG GAT TGG
Gly Gly Gly Glu Ala Leu Val Glu Gly Asp Pro Asn Leu Tyr Leu Het Asp Trp
549 558 567 576 505 594
TCG CCA GCC GAC ACT GTG GCT ATT CTC CAG GAG GCG
Ser Pro Ala Asp Thr Val Gly Ile Leu Asp His Trp Phe Gly Val Asn Gly Leu
603
GCC GTG CGG CGT CGC AAA GCC AAA TAC TGG AGT ATG GAT AAC GAG CCC GGC ATC
Gly Val Arg Arg Gly Lys Ala Lys Tyr Trp Ser Net Asp Asn Glu Pro Gly Ile
657 666 678 604
TGG GTT GGC ACC CAC GAC GAT GTTA GTTA GT
Trp Val Gly Thr His Asp Asp Val Val Lys Glu Gln Thr Pro Val Glu Asp Phe
The same of the sa

Figure 9a

Bankia gouldi endoglucanese (37021) (continued)

720 729 CTG CAC ACC TAT TTC GAA ACC GCC AAA AAA GCC CGC GCC AAA TTT CCC GGT ATT Leu His Thr Tyr Phe Glu Thr Ala Lys Lys Ala Arg Ala Lys Phe Pro Cly Ile 783 AAA ATC ACC GGT CCG GTG CCC GCT AAT GAG TGG CAG TGG TAT GCC TGG GGC GGT Lys Ile Thr Gly Pro Val Pro Ale Asn Glu Trp Gln Trp Tyr Ale Trp Gly Gly 828 837 TTC TCG GTA CCC CAG GAA CAA GGG TTT ATG AGC TGG ATG GAG TAT TTC ATC AAG Phe Ser Val Pro Gln Glu Gln Gly Phe Met Ser Trp Met Glu Tyr Phe Ile Lyr 873 882 891 CGG GTG TCT GAA GAG CAA CGC GCA AGT GGT GTT CGC CTC GAT GTA CTC GAT Arg Val Scr Glu Glu Gln Arg Ala Ser Gly Val Arg Leu Leu Asp Val Leu Asp 936 945 CTG CAC TAC TAC CCC GGC GCT TAC AAT GCG GAA GAT ATC GTG CAA TTA CAT CGC 954 Leu His Tyr Tyr Pro Gly Ala Tyr Asn Ala Glu Asp Ile Val Gln Leu His Arg 990 999 ACG TTC TTC GAC CGC GAC TTT GTT TCA CTG CAT GCC AAC GGG GTG AAA ATG GTA 1008 Thr Phe Phe Asp Arg Asp Phe Val Ser Leu Asp Ala Asn Gly Val Lyo Het Val 1044 1053 1062 GAA GGT GGC TOG GAT GAC AGC ATC AAC AAG GAA TAT ATT TTC GGG CGA GTG AAC Glu Gly Gly Trp Asp Asp Ser Ile Asn Lys Glu Tyr Ile Phe Gly Arg Val Asn 1089 GAT TOG CTC GAG GAA TAT ATG GGG CCA GAC CAT GGT GTA ACC CTG GGC TTA ACC 1116 Asp Trp Leu Glu Glu Tyr Met Gly Pro Asp His Gly Val Thr Leu Gly Leu Thr 1143 1152 1161 GAA ATG TGC GTG CGC AAT GTG AAT CCG ATG ACT ACC GCC ATC TGG TAT GCC TCC 1170 Glu Met Cys Val Arg Asn Val Asn Pro Met Thr Thr Ala Ile Trp Tyr Ala Ser 1206 1215 1224 ATG CTC GGC ACC TTC GCG GAT AAC GGC GTC GAA ATA TTC ACC CCA TGG TGC TGG Met Leu Gly Thr Phe Ala Asp Asn Gly Val Glu Ile Phe Thr Pro Trp Cys Trp 1251 1260 1269 AAC ACC GGA ATG TGG GAA ACA CTC CAC CTC TTC AGC CGC TAC AAA CCT TAT 127 B Asn Thr Gly Met Trp Glu Thr Leu His Leu Phe Ser Arg Tyr Asn Lys Pro Tyr 1314 1323 1332 CGG GTC GCC TCC AGC TCC AGT CTT GAA GAG TTT GTC AGC GCC TAC AGC TCC ATT Arg Val Ala Ser Ser Ser Ser Leu Glu Glu Phe Val Ser Ala Tyr Ser Ser Ile 1368 1377 1386 AMC GAA GCA GAA GAC GCC ATG ACG GTA CTT CTG GTG AAT CGT TCC ACT ACC GAG Asn Glu Ala Glu Asp Ala Met Thr Val Leu Leu Val Asn Arg Ser Thr Ser Glu

Figure 9b(Continued)

Bankia gouldi endoglucanase (37GP1) (continued)

1413 1422 1431 1440 1449 1458
ACC CAC ACC GCC ACT GTC GCT ATC GAC GAT TTC CCA CTG GAT GGC CCC TAC CGC
Thr His Thr Ala Thr Val Ala Ile Asp Asp Phe Pro Leu Asp Gly Pro Tyr Arg

1467 1476 1485 1494 1503 1512
ACC CTG CGC TTA CAC AAC CTG CCG GGG GAG GAA ACC TTC GTA TCT CAC CGA GAC
Thr Leu Arg Leu His Asn Leu Pro Gly Glu Glu Thr Phe Val Ser His Arg Asp

1521 1530 1539 1548 1557 1566
AAC GCC CTG GAA AAA GGT ACA GTG CGC GCC AGC GAC AAT ACG GTA ACA CTG GAG
AEN Ala Leu Glu Lys Gly Thr Val Arg Ala Ser Aep Aen Thr Val Thr Leu Glu

1575 1584 1593 1602 1611
TTG CCC CCT CTG TCC GTT ACT GCA ATA TTG CTC AAG GCC CGG CCC TAA 3*
Leu Pro Pro Leu Ser Val Thr Ala Ile Leu Leu Lys Ala Arg Pro ***

Figure 94 (Continued)

Theresitoga maritima Alpha-qalactosidade Complete Gone Sequence (LC + 3)

5' GTG ATC TGT GTG GAA ATTA TITC GGA ANG ACC TTC ACA CAG GGA ANA TTC GTT CTC
Val Ile Cys Val Glu Ile Phe Gly Lys Thr Phe Arg Glu Gly Arg Phe Val Leu
ANA GAG ANA ANC TITE ACA CIT GAG THE GGG GTG GAG ANG ATA CAC CIT GGC TGG
Lys Glu Lys Asn Phe Thr Val Glu Phe Ala Val Glu Lys Ile His Leu Gly Trp
AAG ATC TCC GGC AGG GTG AAG GGA AGT CCG GGA AGG CTT GAG GTT CTT CGA ACG
Lys Ile Ser Gly Arg Val Lys Gly Ser Pro Gly Arg Leu Glu Val Leu Arg Thr
ANA GCA CCG GNA ANG GTA CTT GTG ANC ANC TGG CAG TCC TGG GGA CCG TGC AGG
Lys Ala Pro Glu Lys Val Leu Val Asn Asn Trp Gln Ser Trp Gly Pro Cys Arg
OTG GTC GAT GCC TIT TCT TTC AAA CCA CCT GAA ATA GAT CCG AAG TGG AGA TAC
Val Val Asp Ala Phe Ser Phe Lys Pro Pro Clu Ile Asp Pro Asm Trp Ary Tyr
ACC GCT TCG GTG GTG CCC GAT GTA CTT GAA AGG AAC CTC CAG AGC GAC TAT TTC
Thr Ala Ser Val Val Pro Asp Val Lou Glu Arg Asm Leu Gln Ser Asp Tyr Phe
333 342 351 360 369 378 GTG GCT GAA GAA GGA AAA GTG TAC GGT TTT CTG AGT TCG AAA ATC GCA CAT CCT
Val Ala Glu Glu Gly Lys Val Tyr Gly Phe Leu Ser Ser Lys Ile Ala His Pro
387 396 405 414 423
Phe Phe Ala Val Glu Asp Gly Glu Leu Val Ala Tyr Leu Glu Tyr Phe Asp Val
441 450 459 468 479
GAG TIC GAC GAC TIT GIT CCT CIT GAA CCT CIC GIT OIA CIC GAG GAT CCC AAC
Glu Phe Amp Amp Phe Val Pro Leu Glu Pro Leu Val Val Leu Glu Amp Pro Am
ACA CCC CITI CTT CTG GAG AAA TAC GCG GAA CTC GTC GGA ATG GAA AAC AAC GCG
The Pro Leu Leu Clu Lys Tyr Ala Clu Leu Val Cly Met Glu Asn Asn Ala
AGA GTT: CCA ANA CAC ACA CCC ACT CGA TGG TGG TGG TAG CAT TAG TTG CTT
Arg Val Pro Lyu His The Pro The Gly Trp Cyr Ser Trp Tyr His Tyr Phe Leu

Figure 10a

Thermotoga maritima Alpha-galactosidane Complete Gune Sequence (2 of 7)

· · · · · · · · · · · · · · · · · · ·
603 612 621 630 639 648 GAT CTC ACC TOG GAA GAG ACT CTC AAG AAC CTC AAG CTC OCG AAG AAT TTC CCC
Asp Leu Thr Trp Glu Glu Thr Leu Lys Asn Leu Lys Leu Ala Lys Aon Phe Pro
657 666 675
THE CAR AIR GAC CAC TAC CAR ANG CAC ATA GGT GAC TOG CTC
Phe Glu Val Phe Gln Ile Asp Asp Ala Tyr Glu Lys Asp Ile Gly Asp Trp Leu
711 720 729 738 747 756 OTG ACA AGA GGA GAC TIT CCA TCG GTG GAA GAG ATG GCA AAA OTT ATA OCG GAA
Val Thr Arg Gly Asp Phe Pro Ser Val Glu Glu Met Ala Lys Val Ile Ala Glu
765 774 783 783
AAC GOT TTC ATC CCG GGC ATA TGG ACC GCC CCG TTC AGT GTT TCT GAA ACC TCG
Asn Gly Phe Ile Pro Gly Ile Trp Thr Ala Pro Phe Ser Val Ser Glu Thr Ser
819 828 837 846 855 864 GAT GTA TTC AAC GAA CAT CCG GAC TGG GTA GTG AAG GAA AAC GGA GAG CCG AAG
Asp Val Phe Asn Glu His Pro Asp Trp Val Val Lys Glu Asm Gly Glu Pro Lys
ATG GCT TAC AGA AAC TOG AAC AAA AAG ATA TAC GCC CTC GAT CTT TCG AAA GAT
Met Ala Tyr Arg Asn Trp Asn Lys Lys Ile Tyr Ala Leu Asp Leu Ser Lys Asp
927 936 945 954 952 952
CAG GIT CTG AAC TGG CIT TTG GAT CTC TTG TGA TGT CTG AGA AAG ATG GGC TAC
Glu Val Leu Asn Trp Leu Phe Asp Leu Phe Ser Ser Leu Arg Lys Met Gly Tyr
AGG TAC TTC AAG ATC GAC TIT CTC TTC GCG GGT GCC GTT CCA GGA GAA AGA AAA
Arg Tyr Phe Lys Ile Asp Phe Leu Phe Ala Gly Ala Val Pro Gly Glu Arg Lys
1035 1044 1053 1062 1071 1000
AND AND ACA COA ATT CAG GCG TTC AGA ANA GGG ATT GAG ACG ATC AGA ANA
Lys Asn Ile Thr Pro Ile Gln Ala Phe Arg Lys Gly Ile Glu Thr Ile Arg Lys
, 1089 1098 1107 1116 1125 1134 GCC GCG GCA GCA GCA GAA GAA GAT TCT TTC ATC CTC GCA TGC GCC TCT CCC CTT CTT CCC GCA
Ala Val Gly Glu Amp Ser Phe Ile Leu Gly Cys Gly Ser Pro Leu Leu Pro Ala
1141
TTG CCA TCC CTC CAC CCC ATG AGG ATA GGA CCT CAC ACT CCG CCG TTC TCG GGA
Val Gly Cys Val Asp Cly Het Arg Ile Gly Pro Asp Thr Ala Pro Phe Trp Gly

Figure 10 (Continued)

Thermutoga maritima Alpha-qalactoridane Complete Gone Sequenca (3 5 1 3)

1197 1206 1215 1224 1233 GAA CAT ATA GAA GAC AAC CO'A OCT CUC OCT GCA ACA TOG OCG CTG AGA AAC OCC Glu His Tie Glu Asp Asn Cly Ala Pro Ala Ala Arg Trp Ala Lou Arg Asn Ala 1260 1269 1278 ATA ACG ACG TAC TIC ATC CAC CAC ACG TIC TOG CTG AAC GAC CCC CAC TOT CTG Ile Thr Arg Tyr Phe Met His Asp Arg Phe Trp Leu Asm Asp Pro Asp Cys Leu 1305 1314 1323 1332 ATA CTG AGA GAG GAG AAA ACG GAT CTC ACA CAG AAG GAA AAG GAG CTC TAC TCG The Lau Arg Glu Glu Lys Thr Asp Leu Thr Gln Lys Glu Lys Glu Leu Tyr Ser 1368 THE ACG TOT GOA GTG CTC GAC AND ATC ATC ATA GAA AGG GAT GAT CTC TCG CTC 1386 . 1404 Tyr Thr Cys Gly Val Leu Asp Asn Met Ile Ile Glu Ser Asp Asp Leu Ser Leu 1422 1431 1440 GTC AGA GAT CAT GGA AAA AAG GTT CTC AAA GAA ACG CTC GAA CTC CTC GGT GGA Val Ary Asp His Gly Lys Lys Val Leu Lys Glu Thr Leu Glu Leu Leu Gly Gly 1476 AGA COA COG GTT CAA AAC ATC ATG TOG CAG GAT CTG AGA TAC GAG ATC GTC TOG 1485 1494 Arg Pro Arg Val Gln Asn Ile Met Ser Glu Asp Leu Arg Tyr Glu Ile Val Ser 1521 1548 TOT GGC ACT CTC TCA CCA AAC GTC AAG ATC GTG GTC GAT CTG AAC AGC AGA GAG Ser Gly Thr Leu Ser Gly Asn Val Lys Ile Val Val App Lon Face Cam lang Glu 1584 1593 THE CAE CITY GAN ANN GAN GON AND THE THE CITY ANN AND AGE OTH ANN AGE Tyr His Leu Glu Lys Glu Gly Lys Ser Ser Leu Lys Lys Arg Val Val Lys Arg 1629 1638 GRA GAC GCA AGA AAC TTC TAC TTC TAC GAA GAG GGT GAG AGA GAA TGA 3. 1647 Glu Asp Gly Arg Asn Phe Tyr Phe Tyr Glu Glu Gly Glu Arg Glu ...

Figure 10c(Continued)

Thermotoga maritima β-mannanase (saper) (GGPA)

			۵			18			27			36			45			54
ς.	ATG	സ്ത	ATT	GGT	GGC	CYC	GAC	TCC	TGG	AGC	CCG	TCA	GTA	TCG	GCG	GAA	TTC	CLI
,																		
	Met	Gly	Ile	Gly	Gly	λsp	qeA	Ser	Trp	Ser	Pro	Ser	Val	Ser	Ala	Glu	Phe	Leu
		_																
			63			72			81			90	. ~	~ ~	99			108
	TTA								GIT	CIC	TIT	GCA	AGT	GAC	GAG.	TIC	GIG	***
													Ser					
	Leu	Leu	116	AGT	GIU	Dea	Ser	146	141				•••	,				2,0
			117			126			135			144			153			162
	GTG	GAA	AAC	GGA	λλλ	TIC	GCT	CTG	AAC	GGA	$\lambda\lambda\lambda$	GAA	TTC	λGλ	TTC	ATT	GGA	λGC
	Val-	Glu	Asn	Gly	Lys	Phe	λla	Leu	λsn	Gly	ŗys	Glu	Phe	Arg	Phe	Ile	Gly	Ser
												198			207			216
	AAC		171			180	m> c	110	189	110	GGA		ATA			GTT	CTG	
	AAC	AAC	TAC	TAC	ATG													
	λsn	3 8 7	TVT	TVI	Met	His	TYI	Lys	Ser	nak	Gly	Xet	Ile	λsp	Ser	Val	Leu	Glu
	,,,,,,,,,,	<i>,</i> ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	.,-	- , -				•										
			225			234			243			252			261			270
	AGT	GCC	AGA	GAC	ATG	CCT	ATA	λλG	ac	CIC	λGA	ATC	TGG	GGT	IIC	CTC	CAC	GGG
												710	~		Dho.	7.00		Clv
	Ser	Ala	Arg	Asp	Met	GIA	He	rya	VAI	Leu	AFG	114,	Trp	GLY	511A	Den	لإحم	GIA
			279			288			297			306			315			324
	GAG	ACT	TAC	TGC	λGλ	GAC	AAG	AAC		TAC	λTG	CAT	CCI	GAG	ထင	CCT	CII	TTC
	Glu	Ser	Tyr	Cys	Arg	Asp	Lys	yen	Thr	IXI	Met	His	Pro	Glu	Pro	Gly	Val	Phe
												360			369			378
			333			342	m-c		351	CAG	AGC.		TTC	GAA		حيرد	GAC	
	GGG	CIG	CCA	GAA	GGA	ATA	100											
	Gly	Val	Pro	Glu	Glv	Ile	Ser	Asn	Ala	Gln	Ser	Gly	Pbe	Glu	Arg	Leu	Asp	Tyr
	4-1		•••	•														
			387			396			405			414			423			432
	ACA	GTT	CCC	χχχ	GCG	λλλ	CYY	CLC	GGT	ATA	XXX		GTC	ATT	GIT	CIT	GTG	AAC
									~~~	73.0	7.45	T 011	Val		7/27	1.00	Val	Aen
	Thr	Val	Ala	Lys	YTZ	Lys	GIU	Leu	GIA	116	Lys	Deu	***	116	143	<b>5</b> 60	*44	~
			441			450			459			468			477			486
	AAC	TGG	GAC	GAC	TTC	GGT	GGA	ATG	AAC	CAG	TAC	GTG	AGG	TGG	TIT	GGA	GGA	ACC
	λεη	Trp	Asp	Asp	Phe	: Gly	Gly	Met	λεπ	Gln	TY	Val	Arg	Trp	Phe	Gly	GJA	Thr
									E13			522	,		531			540
			495			504		Can	513 GAG		. ATY			GAG			AAG	TAC
	CAT	CYC	GAC	. GA1														
	Ris	His	AST	λsı	Phe	тут	Arg	aA ;	Glu	Lys	Ile	e Lye	Glu	Glu	Ty	Lys	Lys	Tyr

Figure 11a

	•	Ther	moto	ga	meri	Ltim	- β		BAB		(10)	<del></del>	(c	onti	.DD.O	4) (	رہے رہے	12)
		549			558			56	7		576	5		585	5		594	
GTC	TC	TI	, CLC	CT	\ XX	CA:	CI	C AA'	ד אכי	C TAC	acc	GG	GT		- ኮ ጥል/	: AG		
ATT	Sei	Phe	: Leu	, Val	. Asr	His	Va.	l Ası	n Thi	ר דאז	Thr	: Gly	/ Val	. Pro	Ty:	Ar	Glu	
		603	}		612	!		623	,		630	ı		636				
GAG	ccc	. ACC	ATC	ATG	GCC	TGG	GAG	cm:	C GC	. AAC	GAA	ccc		635 TCT	י יראר	~	648 GAC	
Glu	Pro	Thr	Ile	Met	λla	Trp	Glu	Leu	λla	Asn	Glu	Pro	Arg	Cys	Glu	The	Asp	
		657												_				
AAA	TCG			300	666	~~~	C10	675			684			693			702	
			<b>XX</b> C			G11		100	GIG	AAG	GAG	ATG	AGC	TCC	TAC	ATA	λλG	
Lys	Ser	Gly	λsn	Thr	Leu	Val	Glu	Trp	Val	Lvs	Glu	Met	Ser	Ser	~~~	71-		
								·		•					-71	116	rys	
100		711			720			729			738			747			756	
AGT	-16	GAT	CCC	AAC	CAC	CTC	GIG	CCI	GIG	GGG	CYC	GAA	CCA	TTC	TTC	<b>A</b> GC	AAC	
Ser	Leu	Asp	Pro	) en	Hie	Leu	V= 1	31-	17-1									
		.~,			****	neu	ATT	ALA	Val	GIY	ASP	Glu	GΙΆ	Phe	Phe	Ser	Asn	
		765			774			783			792			801			810	
TAC	Gλλ	CCY	TTC	XXX	CCT	TAC	CCT	GGA	GAλ	GCC	GAG	TGG	CCC	TAC	AAC	GGC	TGG	
										~								
Tyr	GIU	CTA	Pne	rys	PTO	TYT	Gly	Gly	Glu	λla	Glu	TTP	Ala	ŢYI	λsn	Gly	Trp	
		819			828			837			846			055				
TCC (	GGT	GTT	GAC	TGG		AAG	CTC	CII	TCG	ATA	GAG	ACG	GTG	855 GAC	رخلعك	~~~	864	
Ser (	Gly	Val	yeb .	Lrp	Lys	Lys	Leu	Leu	Ser	Ile	Glu	Thr	Val	λsp	Phe	Gly	Thr	
		873																
TTC (	CAC		TAT		882 TCC	CAC	ביניאד	891	~~	»~	900	<b>~</b> ~ ~		909			918	
Phe i	His	Leu	Tyr .	PTO	Ser	His	Trp	Gly	Val	Ser	Pro	Glu	Asn	Tvr	Ala	Gla	<u>т</u>	
																		-
GGA (	300	927	m		936 533	C) C	~ . ~	945			954			963			972	
GGA (						GAC	CAC	ATA	AAG	ATC	GCA.	AAA	GAG	ATC	GGA .	AAA	CCC	
Gly A	Ala	Lys	Trp :	Ile	Glu .	Asp	His	Ile	Lvs	Ile .	Ala i	Lve	Glu					
						•						-, -		116	GIÀ	PAR	Pro	
		981			990			999		1	800		1	017		1	026	
GTT (	STT	CIC	GAA (	GAA '	TAT	GGA .	ATT	CCY .	λλG	AGT	GCG	CCA	GTT	AAC	AGA .	ACG	GCC	
 Val v								 D										
Val V		Jeu	ora (	J1 u	AYL '	o t A	TTE	rio	rys	ser.	VT9	PTO	Val	Asn	Arg	Thr	Ala	
		035			044		1	053		1	062		1	071		1	080	
ATC 1	CAC	AGA	CIC .	rgg .	AAC (	GAT	CTG	GTC	TAC	GAT	CTC	GGT	GGA	GAT	GGA	ece ,	ATG	
									~									
Ile 7	УY	Arg	Leu '	. <del>Q.</del> 1	Asn,	Asp	Leu	Val	Tyr	Asp	Leu	Gly	Gly	Asp	Gly	Ala	Met	

Figure 11b(Continued)

Thermotoga maritima β-mannanase (mac) (continued) (66P2) 1098 TTC TGG ATG CTC GCG GGA ATC GGG GAA GGT TCG GAC AGA GAC GAG AGA GGG TAC --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Phe Trp Met Leu Ala Gly Ile Gly Glu Gly Ser Asp Arg Asp Glu Arg Gly Tyr 1143 1152 TAT CCG GAC TAC GAC GGT TTC AGA ATA GTG AAC GAC AGT CCA GAA GCG GAA --- --- --- --- --- --- --- --- --- --- ---Tyr Pro Asp Tyr Asp Gly Phe Arg Ile Val Asn Asp Asp Ser Pro Glu Ala Glu 1197 1206 1215 CTG ATA AGA GAA TAC GCG AAG CTG TTC AAC ACA GGT GAA GAC ATA AGA GAA GAC Leu Ile Arg Glu Tyr Ala Lys Leu Phe Asn Thr Gly Glu Asp Ile Arg Glu Asp 1251 1260 1269 ACC TGC TCT TTC ATC CTT CCA AAA GAC GGC ATG GAG ATC AAA AAG ACC GTG GAA --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Thr Cys Ser Phe Ila Leu Pro Lys Asp Gly Met Glu Ile Lys Lys Thr Val Glu 1305 1314 GTG AGG GCT GGT TTC GAC TAC AGC AAC ACG TTT GAA AAG TTG TCT GTC AAA 1323 Val Arg Ala Gly Val Phe Asp Tyr Ser Asn Thr Phe Glu Lys Leu Ser Val Lys 1368 GTC GAA GAT CTG GTT TTT GAA AAT GAG ATA GAG CAT CTC GGA TAC GGA ATT TAC Val Glu Asp Leu Val Phe Glu Asn Glu Ile Glu His Leu Gly Tyr Gly Ile Tyr 1422 1431 GGC TTT GAT CTC GAC ACA ACC CGG ATC CCG GAT GGA GAA CAT GAA ATG TTC CTT Gly Phe Asp Leu Asp Thr Thr Arg Ile Pro Asp Gly Glu His Glu Met Phe Leu 1467 1476 GAA GGC CAC TIT CAG GGA AAA ACG GTG AAA GAC TCT ATC AAA GCG AAA GTG GTG 1485 Glu Gly His Phe Gln Gly Lys Thr Val Lys Asp Ser Ile Lys Ala Lys Val Val 1521 1530 AAC GAA GCA CGG TAC GTG CTC GCA GAG GAA GTT GAT TTT TCC TCT CCA GAA GAG 1539 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Asn Glu Ala Arg Tyr Val Leu Ala Glu Glu Val Asp Phe Ser Ser Pro Glu Glu 1584 1593 GTG AAA AAC TGG TGG AAC AGC GGA ACC TGG CAG GCA GAG TTC GGG TCA CCT GAC Val Lys Asn Trp Trp Asn Ser Gly Thr Trp Gln Ala Glu Phe Gly Ser Pro Asp

Figure 110 (Continued)

			The	Fact	oga	24	riti	Lma	β- <b>=</b> :	anna	nase	u u	i Geo	<del>)</del> (	cont	inu	•d)	(6GP
A7	m.		162	29	AC G	16 GT G	38 AG G	TG (	16 Ga a	47 AT G	GA G	16 CA C	56 TG C	AG (~				_
11	.e	Glu	Tr	 Τρ λ:	n G	ly G	lu V	al (	ly A	sn G	ly A	la L	eu G	in Le	eu A	sn V	al L	 ys Leu
cc	c (		168 AA		EC 67	16 C T	92 3G G	AA G	17 33. G	01 TG AC	EA GT	A GO	10 2 <b>A</b> AC	G A	G T	rc g≱	UA AC	1728 3A CTC
Pr	۰ (	ly	Ly	s Se	rλs	p Ti	no G	lu G	lu Va	al Ar	g Va	1 11	a Ar	g Ly	s Ph		u Ar	g Leu
TC	A G	Άλ 	173 TG:	r GA	G AT	174 C C1	CO	G T	AC GA	C AT	C TA	CAT	4 T CC	X AA	C GI	C GA	G GG	1782 A CTC
Ser	r G						u Gl	ע ש	/T As	p Il	e Ty	r Il	e Pr	As:	n Va	1 Gl	u G1;	y Leu
AAG	G			110	AGO	3 CC	G TA	C GC	180 G GT	T CIK	3 77(	cc	GGG	TGO	182	AAC	AT	1836 A GGC
		1	845			1854	ı		105	,								Gly
CTC	ري  اد		ATG  Met	λλC 	AAC 	GCC	AAC	GT	" GAJ	AGT	. ecc	GAG	ATC	ATC	1881 ACT	TIC		1890 GGA
		1	399			1908			1917			1076						Gly
Lys	Gl	_ ·	YY 	AGA Arg	AGA  Arg	Phe	CAT His	CIV	AGA	ATT	GAG  Glu	TTC	GAC	AGA	ACA	GCG	GGG	GTG
		19	53			1962			1071									
Lys									Gly									
TTC		20	07		;	2015			2025		-	0024						
									Lye							٠ ر		

Figure 11d (Continued)

# ARPII la β-mannosidase (63GB1)

5' ATG CTA CCA GAA GAG TTC CTA TGG GGC GTT GGG CAG TCA GGC TTT CAG TTC GAA
Met Leu Pro Glu Clu Pro
Met Leu Pro Glu Glu Phe Leu Trp Gly Val Gly Gln Ser Gly Phe Gln Phe Glu
01 77
ATG GGC GAC AAG CTC AGG AGG CAC ATC GAT CCA AAT ACC GAC TGG TGG AAG TGG
Met Gly Asp Lys Leu Arg Arg His Ile Asp Pro Asn Thr Asp Trp Trp Lys Trp
11/ 196
GTT CGC GAT CCT TTC AAC ATA AAA AAG GAG CTT GTG AGT GGG GAC CTT CCC GAG
Val Arg Asp Pro Phe Asn Ile Lys Lys Glu Leu Val Ser Gly Asp Leu Pro Glu
1/1 100
CAC GGC ATC AAC AAC TAC GAA CTT TTT GAA AAC GAT CAC AAG CTC GCT AAA CCC
Asp Gly Ile Asn Asn Tyr Glu Leu Phe Glu Asn Asp His Lys Leu Ala Lys Gly
225 234 242
CTT GGA CTC AAC GCA TAC AGG ATT GGA ATA GAG TGG AGC AGA ATC TTT CCC TGG
Leu Gly Leu Asn Ala Tyr Arg Ile Gly Ile Glu Trp Ser Arg Ile Phe Pro Trp
279 700
CCG ACG TGG ACG GTC GAT ACC GAG GTC GAG TTC GAC ACT TAC GGT TTA GTA AAG
Pro The Trp The Val Asp The Clu Val as
Pro Thr Trp Thr Val Asp Thr Glu Val Glu Phe Asp Thr Tyr Gly Leu Val Lys
GAC GTT AAG ATA GAC AAG TCC ACC CTT GCT GAA CTC GAC AGG CTG GCC AAC AAG
Asp Val Ive Tie Ass the Ass
Asp Val Lys Ile Asp Lys Ser Thr Leu Ala Glu Leu Asp Arg Leu Ala Asn Lys
387 306
GAG GAG GTA ATG TAC TAC AGG CGC GTT ATT CAG CAT TTG AGG GAG CTC GGC TTC
Glu Glu Val Met Tyr Tyr Arg Arg Val Ile Gln His Leu Arg Glu Leu Gly Phe
441 454
ANG GTC TTC GTT ANC CTC ANC CAC TTC ACG CTT CCA ATA TGG CTC CAC GAC CCG
Lys Val Phe Val Asn Leu Asn His Phe Thr Leu Pro Ile Trp Leu His Asp Pro
495 504
ATA GTG GCA AGG GAG AAG GCC CTC ACA AAC GAC AGA ATC GGC TGG GTC TCC CAG
Ile Val Ala Arg Glu Lys Ala Leu Thr Asn Asp Arg Ile Gly Trp Val Ser Gln
And Asp Arg Ile Gly Trp Val Ser Gln

Figure 12a

# AMPII la β-mannosidase (63GB1) (continued)

		549			558			56	7		576	5		585	5		594
λG	G AC	CIM	CIL	CAC	TT	CCC	: AAC	TA:	r cc	r GC	TAC	: ATC	: GC	CA'	- ר ככי	: (-14	יים. ייני
												·					
Arg	Thi	. Val	. Val	. Glu	Phe	<b>λ</b> 14	Lys	יעד י	: Ala	Ala	Tyz	Ile	λla	His	. Al.	1.61	
											-						. Gry
		603			612			621	L		630			639	•		648
GAC	CTC	GTG	GAC	YCY	TGG	λGC	. YCC	TTC	: AAC	ေယ	CCT	ATG	GTA	GTT	GIV	GAG	
						~											
Asp	Leu	Val	λsp	Thr	Trp	Ser	Thr	Phe	λsn	Glu	Pro	Met	Val	Val	Val	Glu	Len
		657			666			675			684			693			702
GGC	TAC	CIC	CCC	ccc	TAC	TCA	CCA	TIT	ccc	CCG	GGA	GTC	λTG	AAC	CCC	GAG	GCC
GIY	Tyr	Leu	YTA	Pro	TYT	Ser	Gly	Phe	Pro	Pro	Gly	Val	Met	Asn	Pro	Glu	λla
~~~		711			720			729			738			747			756
	AAG	CTG	فالبافا	ATC	CTC	AAC	ATG	ATA	AAC	CCC	CAC	GCC	TTG	GCX	TAT	λλG	ATG
11.	7.00	7	81-	71-													
7.14	Lys	Deu	AIG	716	rea	ASD	Met	110	Asn	Ala	His	λla	Leu	λla	Tyr	Lys	Met
		765			774			783			700						
ATA	AAG		مكلمك	GAC		110	130	703	C 2 M	~1~	792			801			810
							~~		GAT	GAG	GAT	AGC	AAG	TCC	CCT	CCC	GYC
Ile	Lys	Àrσ	Phe	σaλ	The	Lvs	Lvs	112) an	Glu) ===	C	7				
	-4-	•				-3-	-,-		ىرىب	914	vab	JEL	rys	Ser	PTO	YTA	Asp
		819			828			837			846			855			064
GTT	GGC	λTλ	ATT	TAC	AAC	λλC	ATC		GTT	GCC	TAC	CCT	222	CAC			864
							~										
Val	Gly	Ile	lle	Tyr	Asn	λsn	Ile	Gly	Val	λla	Tyr	Pro	Lvs	λερ	PTO	À STO) en
				•				-			-			,		7-2-1	, wy
		873			882			891			900			909			918
CCC	AAG	GAC	CTT	XXX	GCA	GCC	GAA	XXC	GAC	AAC	TAC	TTC	CAC	AGC	GGA	CTG	TTC
Pro	Lys	Asp	Val	Lys	Ala	Ala	Glu	λsn	qzλ	Asn	Tyr	Phe	His	Ser	Gly	Leu	Phe
															-		
		927			936			945			954			963			972
Tip	GAT	GCC	YŢC	CXC	λλG	CCT	AAG	CIC	AAC	ATA	GAG	TTC	CYC	GCC	GAA	AAC	TTT
Db -																	
Pne	λsp	VTG	IIS	Hls	ГÄЗ	Gly	Lys	Leu	λsn	Ile	Glu	Phe	Ąsp	Gly	Glu	Yau	Phe
CT'A		981			990			999		1	008		_ 1	.017		1	026
	λλλ	GFF	AUA	CAC	CTA	***	GGC	AAT	CAC	TGG	ATA	GGC	CIC	AAC	TAC	TAC	ACC
Ua 1	Tare	Val	1	u: -	T 4	 *		·									
***	Lys	Val	~. 9	шз	red	Lys	GIA	ASD	АБР	TIP	110	GIÀ	Leu	λan	Tyr	TYT	Thr
	1	.035		1	044		,	053		1	חבי			^~-		_	
CGC	GAG		GTT			TCG	GAG	ברר היים	220	ملحد 1	CCA	مت و	3772	071	~~~	1	080
												VO.1.	~~~	CCC	CIC	ATA	TCC
Arg	Glu	Val	Val	λrα	Tvr	Ser	Glu	Pro	Lvs	Phe	Pro	Ser	T1 e	Dec		71-	
-				- ,										FIO	neu	7 T G	201

Figure 12b(Continued)

APPII 1a β -mannosidase (63GB1) (continued)

(continued)
1089 1098 1107 1116 1125 113. TTC AAG GGC GTT CCC AAC TAC GGC TAC TCC TGC AGG CCC GGC ACG ACC TCC GCC Phe Lys Gly Val Day
and the Ash Tyr Gly Tyr Ser Cys Arg Pro Gly The The San 13
GAT GGC ATG CCC GTC AGC GAT ATC GGC TGG GAA GTC TAT CCC CAC GCL ATC
Act Flo Val Ser Asp Ile Gly Trp Glu Val Tyr Pro Gln Gly Ile m
GAC TCG ATA GTC GAG GCC ACC AAG TAC AGT GTT CCT GTT TAC GTC ACC ACC AAG TAC AGT GTT CCT GTT TAC GTC ACC ACC ACC ACC ACC ACC ACC ACC ACC A
The Val Git Ala The Lys Tyr Ser Val Pro Val Tyr Val The Give Ac-
GGT GTT GCG GAT TCC GCG GAC ACG CTG AGG CCA TAC TAC ATA GTC ATA GTC ATA GTC ATA GTC ATA GTC ACG CTG ATA GTC AT
ory var Ala Asp Ser Ala Asp Thr Leu Arg Pro Tyr Tyr Ile Val Ser Hie Val
TCA AAG ATA GAG GAA GCC ATT GAG AAT GGA TAC CCC GTA AAA GGC TAC ATG
Ser Lys Ile Glu Glu Ala Ile Glu Asn Gly Tyr Pro Val Lys Gly Tyr Met Tyr
1359 1368 1377 1386 1395 1404 TGG GCG CTT ACG GAT AAC TAC GAG TGG GCC CTC GGC TTC AGC ATG AGG TTT GGT
Ala Leu Gly Phe Ser Met Arg Phe Gly
CTC TAC AAG GTC GAC CTC ATC TCC AAG GAG AGG ATC CCG AGG GAG ACA AGG ATC CCG AGG GAG ACA AGG ATC CCG AGG GAG ACA AGG GAG AGG ACA AGG GAG AGG GAG AGG AGG GAG AGG AG
Leu Tyr Lys Val Asp Leu Ile Ser Lys Glu Arg Ile Pro Arg Glu Arg Ser Val
1467 1476 1485 1494 1503 1512 GAG ATA TAT CGC AGG ATA GTG CAG TCC AAC GGT GTT CCT AAG GAT ATC AAA GAG
did lie Tyr Arg Arg Ile Val Gln Ser Asn Gly Val Pro Lys Asp Ile Lys Gly
GAG TTC CTG AAG GGT GAG GAG AAA TGA 3'
Glu Phe Leu Lys Gly Glu Glu Lys ***

Figure 12C(Continued)

OC1/4V Endoglucanase (33GP1)

5' ATG GTA GAA AG GAG TA
5' ATG GTA GAA AGA CAC TTC AGA TAT GTT CTT ATT TGC ACC CTG TTT CTT GTT ATG
Met Val Glu Arg His Phe Arg Tyr Val Leu Ile Cys Thr Leu Phe Leu Val Met
6.3
CTC CTA ATC TCA TCC ACT CAG TGT GGA ANA AAT GAA CCA AAC AAA AGA GTG AAT
Leu Leu Ile Ser Ser Thr Gln Cys Gly Lys Asn Glu Pro Asn Lys Arg Val Asn
117 126
AGC ATG GAA CAG TCA GTT GCT GAA AGT GAT AGC AAC TCA GCA TTT GAA TAC AAC
Ser Met Glu Gln Ser Val Ala Glu Ser Asp Ser Asn Ser Ala Phe Glu Tyr Asn
171
ANA ATG GTA GGT ANA GGA GTA ANT ATT GGA ANT GCT TTA GAN GCT CCT TTC GAN
Lys Met Val Gly Lys Gly Val And The Gi
Lys Met Val Gly Lys Gly Val Asn Ile Gly Asn Ala Leu Glu Ala Pro Phe Glu
225 234 243 252 261 270 GGA GCT TGG GGA GTA AGA ATT GAG GAT GAA TAT TTT GAG ATA ATA
GIV Ala Tro GIV Val And The Till Gall ATA ATA AAG AAA AGG
Gly Ala Trp Gly Val Arg Ile Glu Asp Glu Tyr Phe Glu Ile Ile Lys Lys Arg
279 288 297 306 315 324
GGA TTT GAT TCT GTT AGG ATT CCC ATA AGA TGG TCA GCA CAT ATA TCC GAA AAG
Gly Phe Asp Ser Val Arg Ile Pro Ile Arg Trp Ser Ala His Ile Ser Glu Lys
333 342 351 360 369 378
CCA CCA TAT GAT ATT GAC AGG AAT TTC CTC GAA AGA GTT AAC CAT GTT GTC GAT
Pro Pro Tyr Asp Ile Asp Arg Asn Phe Leu Glu Arg Val Asn His Val Val Asp
387 396
AGG GCT CTT GAG AAT AAT TTA ACA GTA ATC ATC AAT ACG CAC CAT TTT GAA GAA
Arg Ala Leu Glu Asn Asn Leu Thr Val Ile Ile Asn Thr His His Phe Glu Glu
441 450 450
CTC TAT CAA GAA CCG GAT AAA TAC GGC GAT GTT TTG GTG GAA ATT TGG AGA CAG
Leu Tyr Gln Glu Pro Asp Lys Tyr Gly Asp Val Leu Val Glu Ile Trp Arg Gln
495 504 513
ATT GCA AAA TTC TTT AAA GAT TAC CCG GAA AAT CTG TTC TTT GAA ATC TAC AAC
Ile Ala Lys Phe Phe Lys Asp Tyr Pro Glu Asn Leu Phe Phe Glu Ile Tyr Asn

Figure 13A

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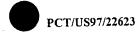
Figure 13b(Continued)



Thermotoga maritima Pullulanase (6GP3)

9 5' ATG GAT CTT ACA AAG	18 GTG GGG ATC	27	36	4 5 5
Met Asp Leu Thr Lys	Val Gly Tla	The Web	TG AAC GAG	TGG CAG GCA AA
Met Asp Leu Thr Lys	72		eu Asn Glu 1	inp Gln Ala Ly:
GAC GTG GCA AAA GAC	AGG TTC ATA	GAG ATA AAA G	90 AC GGA AAG G	99 108 CT GAA GTG TGG
Asp Val Ala Lys Asp	Arg Phe Ile	Clu Ila Lys A	sp Cly Lys A	la Glu Val Trp
ATA CTC CAG GGA GTG	AA GAG ATT	135 1. TTC TAC GAA AJ	14 II UA CCA GAC AG	53 162 CA TCT CCC AGA
Ile Leu Glm Gly Val G	lu Glu Ile	Phe Tyr Glu Ly	Pro Asp Th	
	80	.		
Ile Phe Phe Ala Gin a		anc and GIG AT	C GAG GCT TT	T CTG ACC AAT
Ile Phe Phe Ala Gln A		usn Lys Val II	e Glu Ala Ph	e Lau Thr Asn
COT CTG GAT ACG AAA AA	S4 2 NG AAA GAA C	143 25:	26	1 270
Pro Val Asp Thr Lvs Lv			NCT GTT GA	C GGA AAA GAG
Pro Val Asp Thr Lys Ly		eu Phe Lys Val	Thr Val Asp	Gly Lys Glu
ATT CCC GTC TCA AGA GT	8 2 GGAA AAG G	97 306 CC GAT CCC ACC	319	324
Ile Pro Val Ser Arg Va	 1 Glu Lve 1		GAC	GTG ACG AAC
	_		Asp Ile Asp	Val Thr Asn
TAC GTG AGA ATC GTC CT	2 I TCT GAA TO	51 360 ℃ CTG AAA GAA	369 GAA GAC 0000	378
Tyr Val Arg Ile Val Le	 1 Ser Glu Se			AGA AAA GAC
387 390			Glu Asp Leu	Arg Lys Asp
GTG GAA CTG ATC ATA GA	A GGT TAC AA)5 414 A CCG GCA AGA	423	432
Val Glu Leu Ile Ile Glu	Gly Tyr Tay		AIG	ATG GAG ATC
			Val Ile Met	Met Glu Ile
CTG GAC GAC TAC TAT TAC	45 GAT GGA GA	9 468 G CTC GGA GCC	477	486
Leu Asp Asp Tor Tor Tor			GIA TAT TCT	CCX GAG AAG
Leu Asp Asp Tyr Tyr Tyr		u Leu Gly Ala	Val Tyr Ser	Pro Glu Lys
ACG ATA TTC AGA GTC TGG	TCC CCC CT	522	531	540
Thr Ile Phe ive Val man			GTA AAG GTG	CTT CTC TTC
Thr Ile Phe Arg Val Trp	Ser Pro Va	l Ser Lys Trp	Val Lys Val	Leu Leu Phe

Figure 14a



Thermotoga maritime Pullulanese (5GP3) (continued)
549 559 559
549 558 567 576 585 594 AAA AAC GGA GAA GAC ACA GAA CCG TAC CAG GTT GTG AAC ATG GAA TAC AAG GGA
Lys Asp Gly Gly Asp The Cas The CAS GIT GTG AAC ATG GAA TAC AAG GGA
Lys Asn Gly Glu Asp Thr Glu Pro Tyr Gln Val Val Asn Met Glu Tyr Lys Gly
The sal val Ash Met Glu Tyr Lys Cly
603 612 621 630 639 640
AAC GGG GTC TGG GAA GCG GTT GTT GAA GGC GAT CTC GAC GGA GTG TTC TAC CTC
Asn Gly Val Trp Glu Ala Wal Val and
Asn Gly Val Trp Glu Ala Val Val Glu Gly Asp Leu Asp Gly Val Phe Tyr Leu
657 666 674
TAT CAG CTG GAA AAC TAC GGA AAG ATC AGA ACA ACC GTC GAT CCT TAT TCG AAA
THE COLUMN TOTAL T
Tyr Gln Leu Glu Asn Tyr Gly Lys Ile Arg Thr Thr Val Asp Pro Tyr Ser Lys
711 700
GCG GTT TAC GCA AAC AAC CAA CAA CAA GCG AAC AAC A
GCG CTT TAC GCA AAC AAC CAA GAG AGC GCC GTT GTG AAT CTT GCC AGG ACA AAC
Ala Val Tyr Ala Asn Asn Gln Glu Ser Ala Val Val Asn Leu Ala Arg Thr Asn
765 774 783 792 801 810
CCA GAA GGA TGG GAA AAC GAC AGG GGA CCG AAA ATC GAA GGA TAC GAA GAC GCG
Pro Glu Gly Tro Glu Are han han and gland and
Pro Glu Gly Trp Glu Asn Asp Arg Gly Pro Lys Ile Glu Gly Tyr Glu Asp Ala
819 828 627
ATA ATC TAT GAA ATA CAC ATA GCG GAC ATC ACA GGA CTC GAA AAC TCC GGG GTA
The The Man class to the term of the term
Ile Ile Tyr Glu Ile His Ile Ala Asp Ile Thr Gly Leu Glu Asn Ser Gly Val
873 882 893
ANA NAC ANA GGC CTC TAT CTC GGG CTC ACC GAN GAN NAC ACG ANN GGN CCG GGC
THE GAN MAC ACG AAA GGA CCG GGC
Lys Asn Lys Gly Leu Tyr Leu Gly Leu Thr Glu Glu Asn Thr Lys Gly Pro Gly
927
927 936 945 954 963 972
GGT GTG ACA ACA GGC CTT TCG CAC CTT GTG GAA CTC GGT GTT ACA CAC GTT CAT
Gly Val Thr Thr Gly Leu Ser His Leu Val Glu Leu Gly Val Thr His Val His
981 990 999 1008 1017 1026
TA CIT CCT TIC TIT GAT TIC TAC ACA CCC CAC CAA COC CAA
le Leu Pro Phe Phe Ann Phe True
le Leu Pro Phe Phe Asp Phe Tyr Thr Gly Asp Glu Leu Asp Lys Asp Phe Glu
1035 1044 1052
AG TAC TAC AAC TGG GGT TAC GAT CCT TAC CTG TTC ATG GTT CCG GAG GGC AGA
THE ALG GIT CUU GAG GGC AGA
ys Tyr Tyr Asn Trp Gly Tyr Asp Pro Tyr Leu Phe Met Val Pro Glu Gly Arg
and the state of t

Figure 14b(Continued)



Thermotoga maritima Pullulanase (6GP3) (continued)

1098 TAC TCA ACC GAT CCC AAA AAC CCA CAC ACG AGA ATC AGA GAA GTC AAA GAA ATG 1107 --- --- --- --- --- --- --- --- --- --- --- --- --- ---Tyr Ser Thr Asp Pro Lys Asn Pro His Thr Arg Ile Arg Glu Val Lys Glu Met 1152 1161 GTC AAA GCC CTT CAC AAA CAC GGT ATA GGT GTG ATT ATG GAC ATG GTG TTC CCT --- --- --- --- --- --- --- --- --- --- --- --- --- ---Val Lys Ala Leu His Lys His Gly Ile Gly Val Ile Met Asp Met Val Phe Pro 1197 1206 1215 CAC ACC TAC GGT ATA GGC GAA CTC TCT GCG TTC GAT CAG ACG GTG CCG TAC TAC --- --- --- --- --- --- --- --- --- --- --- --- --- ---His Thr Tyr Gly Ile Gly Glu Leu Ser Ala Phe Asp Gln Thr Val Pro Tyr Tyr 1260 1269 TTC TAC AGA ATC GAC AAG ACA GGT GCC TAT TTG AAC GAA AGC GGA TGT GGT AAC --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Phe Tyr Arg Ile Asp Lys Thr Gly Ala Tyr Leu Asn Glu Ser Gly Cys Gly Asn 1305 1314 GTC ATC GCA AGC GAA AGA CCC ATG ATG AGA AAA TTC ATA GTC GAT ACC GTC ACC 1323 --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Val Ile Ala Ser Glu Arg Pro Met Met Arg Lys Phe Ile Val Asp Thr Val Thr 1368 1377 TAC TGG GTA AAG GAG TAT CAC ATA GAC GGA TTC AGG TTC GAT CAG ATG GGT CTC Tyr Trp Val Lys Glu Tyr His Ile Asp Gly Phe Arg Phe Asp Gln Met Gly Leu 1413 1422 ATC GAC AAA AAG ACA ATG CTC GAA GTC GAA AGA GCT CTT CAT AAA ATC GAT CCA 1431 Ile Asp Lys Lys Thr Met Leu Glu Val Glu Arg Ala Leu His Lys Ile Asp Pro 1476 1485 ACT ATC ATT CTC TAC GGC GAA CCG TGG GGT GGA TGG GGA GCA CCG ATC AGG TIT Thr Ile Ile Leu Tyr Gly Glu Pro Trp Gly Gly Trp Gly Ala Pro Ile Arg Phe 1530 GGA AAG AGC GAT GTC GCC GGC ACA CAC GTG GCA GCT TTC AAC GAT GAG TTC AGA 1539 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Gly Lys Ser Asp Val Ala Gly Thr His Val Ala Ala Phe Asn Asp Glu Phe Arg 1584 1593 GAC GCA ATA AGG GGT TCC GTG TTC AAC CCG AGC GTC AAG GGA TTC GTC ATG GGA 1602 --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- --- ---Asp Ala Ile Arg Gly Ser Val Phe Asn Pro Ser Val Lys Gly Phe Val Met Gly

Figure 14C(Continued)

Thermotoga maritima Pullulanase (6GP3) (continued)

(GD3) (Continued)
1629 1638
1629 1638 1647 1656 1665 1674
THE AND AIR AAA AGG GGT GTT GGA AGC ATA AAC THE
Gly Tyr Gly Lys Glu Thr Lys Ile (Im Ann C)
Gly Tyr Gly Lys Glu Thr Lys Ile Lys Arg Gly Val Val Gly Ser Ile Asn Tyr
1 / 7 7
1683 1692 1701 1710 1719 1728
GAC GGA AAA CTC ATC AAA AGT TTC GCC CTT GAT CCL CLL CLL CLL CLL CLL CLL CLL CLL CL
GAC GGA ANA CTC ATC ANA AGT TTC GCC CTT GAT CCA GAA GAA ACT ATA AAC TAC
Asp Gly Lys Leu Ile Lys Ser Phe Na You
Asp Gly Lys Leu Ile Lys Ser Phe Ala Leu Asp Pro Glu Glu Thr Ile Asn Tyr
1737 1746 454
GCA GCG TGT CAC GAC AAC CAC ACA CTG TGG TGG 1764 1773 1782
THE THE THE THE THE TAG GAC AND AND THE CITY GCC GCC AND
Ala Ala Cys His Asp Asn His Thr Leu Trp Asp Lys Asn Tyr Leu Ala Ala Lys
of the Asp Ash His Thr Leu Trp Asp Lys Ash Tyr Leu Ala Ala Luc
1791 1800 1000
1791 1800 1809 1818 1827
THE MAN AND ANA ANG GAN TOG ACC GAN GAN GAN CTG AND ANG CCC CO
Ala Asp Lys Lys Glu Tro Thr Glu Glu Glu Lys
Ala Asp Lys Lys Glu Trp Thr Glu Glu Glu Leu Lys Asn Ala Gln Lys Leu
of the bys Ash Ala Gin Lys Leu
1845 1854 1863 1872 1881 1890
Ala Giv als Tie Law to Ten Con Got GTT CCT TTC CTC CAC GGA GGG CAG
Ala Gly Ala Ile Leu Leu Thr Ser Gln Gly Val Pro Phe Leu His Gly Gly Gln
Sel din diy val Pro Phe Leu His Gly Gly Gln
1899 1909
GAC TTC TGC AGG ACC AND THE 1917 1926 1935 1944
GAC TTC TGC AGG ACG AAT TTC AAC GAC AAC TCC TAC AAC GCC CCT ATC TCG
Asp Phe Cys Arg The The The Table 12
Asp Phe Cys Arg Thr Thr Asn Phe Asn Asp Asn Ser Tyr Asn Ala Pro Ile Ser
1953 1962 1971 1980 1989 1989
THE GOT THE GAT THE GAX AGA AND CITY CAS THE AND AND AND
Ile Asn Gly Phe Asn Tor Gly has your
Ile Asn Gly Phe Asp Tyr Glu Arg Lys Leu Gln Phe Ile Asp Val Phe Asn Tyr
2007 2016 2025 2034 2043
LAC ANG GGT CTC ATA AAA CTC ACL ALL CLA CLA CLA CLA CLA CLA CLA CL
CAC ANG GGT CTC ATA ANA CTC AGA ANA GAA CAC CCT GCT TTC AGG CTG ANA ANC
His Lys Gly Leu Ile Lys Leu Arg Lys Glu His Pro Ala Phe Arg Leu Lys Asn
and all all all all all all all all all al
2061 2020
GCT GAA GAG ATC AAA AAA CAC CTG GAA TTT CTC CCG GGC GGG AGA AGA ATA GTT
THE CAS GAR THE CTC CCC GGC AGA AGA ATA GITT
Ala Glu Glu Ile Lye Lye Lye Lye Can Glu
Ala Glu Glu Ile Lys Lys His Leu Glu Phe Leu Pro Gly Gly Arg Arg Ile Val
2115 2124 2122
2115 2124 2133 2142 2151 2160
THE CAL GLA GGT GGT GAT CCC TGG AAA CAC ATC CTC CTC
Ala Pho Man
Ala Phe Met Leu Lys Asp His Ala Gly Gly Asp Pro Trp Lys Asp Ile Val Val

Figure 14d(Continued)



Thermotoga maritima Fullulanase (6GP3) (continued)

2169 2178 2187 2196 2205 2214
ATT TAC AAT GGA AAC TTA GAG AAG ACA ACA TAC AAA CTG CCA GAA GGA AAA TGG

TILE Tyr Asn Gly Asn Leu Glu Lys Thr Thr Tyr Lys Leu Pro Glu Gly Lys Trp

Asn Val Val Asn Ser Gln Lys Ala Gly Thr Glu Val Ile Glu Thr Val Glu

GGA ACA ATA GAA CTC GAT CCG CTT TCC GCG TAC GTT CTG TAC AGA GAG TGA 3'
Gly Thr Ile Glu Leu Asp Pro Leu Ser Ala Tyr Val Leu Tyr Arg Glu ***

Figure 140(Continued)

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Figure 15a Thermotoga maritima MSB8 (Clone # 6GP2) Glycosidase

CTT TTA TTG ATC GTT GAG CTC TCT TTC GTT CTC TTT GCA AGT GAC GAG TTC Leu Leu Leu Ile Val Glu Leu Ser Phe Val Leu Phe Ala Ser Asp Glu Phe

GTG AAA GTG GAA AAC GGA AAA TTC GCT CTG AAC GGA AAA GAA TTC AGA TTC Val Lys Val Glu Asn Gly Lys Phe Ala Leu Asn Gly Lys Glu Phe Arg Phe

ATT GGA AGC AAC AAC TAC TAC ATG CAC TAC AAG AGC AAC GGA ATG ATA GAC Ile Gly Ser Asn Asn Tyr Tyr Met His Tyr Lys Ser Asn Gly Met Ile Asp

AGT GTT CTG GAG AGT GCC AGA GAC ATG GGT ATA AAG GTC CTC AGA ATC TGG Ser Val Leu Glu Ser Ala Arg Asp Met Gly Ile Lys Val Leu Arg Ile Trp

GGT TTC CTC GAC GGG GAG AGT TAC TGC AGA GAC AAG AAC ACC TAC ATG CAT Gly Phe Leu Asp Gly Glu Ser Tyr Cys Arg Asp Lys Asn Thr Tyr Met His

CCT GAG CCC GGT GTT TTC GGG GTG CCA GAA GGA ATA TCG AAC GCC CAG AGC Pro Glu Pro Gly Val Pne Gly Val Pro Glu Gly Ile Ser Asn Ala Gln Ser

GGT TTC GAA AGA CTC GAC TAC ACA GTT GCG AAA GCG AAA GAA CTC GGT ATA Gly Phe Glu Arg Leu Asp Tyr Thr Val Ala Lys Ala Lys Glu Leu Gly Ile

AAA CTT GTC ATT GTT CTT GTG AAC AAC TGG GAC GAC TTC GGT GGA ATG AAC Lys Leu Val Ile Val Leu Val Asn Asn Trp Asp Asp Phe Gly Gly Met Asn

CAG TAC GTG AGG TGG TTT GGA GGA ACC CAT CAC GAC GAT TTC TAC AGA GAT Gln Tyr Val Arg Trp Phe Gly Gly Thr His His Asp Asp Phe Tyr Arg Asp

GAG AAG ATC AAA GAA GAG TAC AAA AAG TAC GTC TCC TTT CTC GTA AAC CAT Glu Lys Ile Lys Glu Glu Tyr Lys Lys Tyr Val Ser Phe Leu Val Asn His

GTC AAT ACC TAC ACG GGA GTT CCT TAC AGG GAA GAG CCC ACC ATC ATG GCC Val Asn Thr Tyr Thr Gly Val Pro Tyr Arg Glu Glu Pro Thr Ile Met Ala

TGG GAG CTT GCA AAC GAA CCG CGC TGT GAG ACG GAC AAA TCG GGG AAC ACG Trp Glu Leu Ala Asn Glu Pro Arg Cys Glu Thr Asp Lys Ser Gly Asn Thr

CTC GTT GAG TGG GTG AAG GAG ATG AGC TCC TAC ATA AAG AGT CTG GAT CCC Leu Val Glu Trp Val Lys Glu Met Ser Ser Tyr Ile Lys Ser Leu Asp Pro

AAC CAC CTC GTG GCT GTG GGG GAC GAA GGA TTC TTC AGC AAC TAC GAA GGA Asn His Leu Val Ala Val Gly Asp Glu Gly Phe Phe Ser Asn Tyr Glu Gly

TTC AAA CCT TAC GGT GGA GAA GCC GAG TGG GCC TAC AAC GGC TGG TCC GGT Phe Lys Pro Tyr Gly Gly Glu Ala Glu Trp Ala Tyr Asn Gly Trp Ser Gly

GTT GAC TGG AAG AAG CTC CTT TCG ATA GAG ACG GTG GAC TTC GGC ACG TTC Val Asp Trp Lys Lys Leu Leu Ser Ile Glu Thr Val Asp Phe Gly Thr Phe

CAC CTC TAT CCG TCC CAC TGG GGT GTC AGT CCA GAG AAC TAT GCC CAG TGG His Leu Tyr Pro Ser His Trp Gly Val Ser Pro Glu Asn Tyr Ala Gln Trp

GGA GCG AAG TGG ATA GAA GAC CAC ATA AAG ATC GCA AAA GAG ATC GGA AAA Gly Ala Lys Trp Ile Glu Asp His Ile Lys Ile Ala Lys Glu Ile Gly Lys

CCC GTT GTT CTG GAA GAA TAT GGA ATT CCA AAG AGT GCG CCA GTT AAC AGA Pro Val Val Leu Glu Glu Tyr Gly Ile Pro Lys Ser Ala Pro Val Asn Arg

ACG GCC ATC TAC AGA CTC TGG AAC GAT CTG GTC TAC GAT CTC GGT GGA GAT Thr Ala Ile Tyr Arg Leu Trp Asn Asp Leu Val Tyr Asp Leu Gly Gly Asp

GGA GCG ATG TTC TGG ATG CTC GCG GGA ATC GGG GAA GGT TCG GAC AGA GAC Gly Ala Met Phe Trp Met Leu Ala Gly Ile Gly Glu Gly Ser Asp Arg Asp

GAG AGA GGG TAC TAT CCG GAC TAC GAC GGT TTC AGA ATA GTG AAC GAC GAC Glu Arg Gly Tyr Tyr Pro Asp Tyr Asp Gly Phe Arg Ile Val Asn Asp Asp

AGT CCA GAA GCG GAA CTG ATA AGA GAA TAC GCG AAG CTG TTC AAC ACA GGT Ser Pro Glu Ala Glu Leu Ile Arg Glu Tyr Ala Lys Leu Phe Asn Thr Gly

GAA GAC ATA AGA GAA GAC ACC TGC TCT TTC ATC CTT CCA AAA GAC GGC ATG Glu Asp Ile Arg Glu Asp Thr Cys Ser Phe Ile Leu Pro Lys Asp Gly Met

GAG ATC AAA AAG ACC GTG GAA GTG AGG GCT GGT GTT TTC GAC TAC AGC AAC

Figure 15b (continued)



Glu Ile Lys Lys Thr Val Glu Val Arg Ala Gly Val Phe Asp Tyr Ser Asn

ACG TTT GAA AAG TTG TCT GTC AAA GTC GAA GAT CTG GTT TTT GAA AAT GAG Thr Phe Glu Lys Leu Ser Val Lys Val Glu Asp Leu Val Phe Glu Asn Glu

ATA GAG CAT CTC GGA TAC GGA ATT TAC GGC TTT GAT CTC GAC ACA ACC CGG Ile Glu His Leu Gly Tyr Gly Ile Tyr Gly Phe Asp Leu Asp Thr Thr Arg

ATC CCG GAT GGA GAA CAT GAA ATG TTC CTT GAA GGC CAC TTT CAG GGA AAA Ile Pro Asp Gly Glu His Glu Met Phe Leu Glu Gly His Phe Gln Gly Lys

ACG GTG AAA GAC TCT ATC AAA GCG AAA GTG GTG AAC GAA GCA CGG TAC GTG Thr Val Lys Asp Ser Ile Lys Ala Lys Val Val Asn Glu Ala Arg Tyr Val

CTC GCA GAG GAA GTT GAT TTT TCC TCT CCA GAA GAG GTG AAA AAC TGG TGG Leu Ala Glu Glu Val Asp Phe Ser Ser Pro Glu Glu Val Lys Asn Trp Trp

AAC AGC GGA ACC TGG CAG GCA GAG TTC GGG TCA CCT GAC ATT GAA TGG AAC Asn Ser Gly Thr Trp Gln Ala Glu Phe Gly Ser Pro Asp Ile Glu Trp Asn

GGT GAG GTG GGA AAT GGA GCA CTG CAG CTG AAC GTG AAA CTG CCC GGA AAG Gly Glu Val Gly Asn Gly Ala Leu Gln Leu Asn Val Lys Leu Pro Gly Lys

AGC GAC TGG GAA GAA GTG AGA GTA GCA AGG AAG TTC GAA AGA CTC TCA GAA Ser Asp Trp Glu Glu Val Arg Val Ala Arg Lys Phe Glu Arg Leu Ser Glu

TGT GAG ATC CTC GAG TAC GAC ATC TAC ATT CCA AAC GTC GAG GGA CTC AAG Cys Glu Ile Leu Glu Tyr Asp Ile Tyr Ile Pro Asn Val Glu Gly Leu Lys

GGA AGG TTG AGG CCG TAC GCG GTT CTG AAC CCC GGC TGG GTG AAG ATA GGC Gly Arg Leu Arg Pro Tyr Ala Val Leu Asn Pro Gly Trp Val Lys Ile Gly

CTC GAC ATG AAC AAC GCG AAC GTG GAA AGT GCG GAG ATC ACT TTC GGC Leu Asp Met Asn Asn Ala Asn Val Glu Ser Ala Glu Ile Ile Thr Phe Gly

GGA AAA GAG TAC AGA AGA TTC CAT GTA AGA ATT GAG TTC GAC AGA ACA GCG Gly Lys Glu Tyr Arg Arg Phe His Val Arg Ile Glu Phe Asp Arg Thr Ala

Figure 15C(continued)



GGG GTG AAA GAA CTT CAC ATA GGA GTT GTC GGT GAT CAT CTG AGG TAC GAT Gly Val Lys Glu Leu His Ile Gly Val Val Gly Asp His Leu Arg Tyr Asp

GGA CCG ATT TTC ATC GAT AAT GTG AGA CTT TAT AAA AGA ACA GGA GGT ATG Gly Pro Ile Phe Ile Asp Asn Val Arg Leu Tyr Lys Arg Thr Gly Gly Met

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END

Figure 15d(continued)

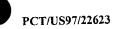


Figure No. 15 Thermotoga maritima MSB8(6gb4)

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6																							
2	, ,	ha c	1 c	3GG /	ACT	GTG	CCX	A GG	G G	TT G	TC C	AG G	CA G	AT (TG	GTC	AG	A A	AA G	GT C	TT C	TT CCA	100
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																							40
12:	1 C	AC C	CG T	AC C	TT	GGG	ATG	AA	C GA	A GA	AT C	C T	C A	AG G	AA 2	מדמ	GA:	A (" N	c			G ATC	
4:	L H:	is P	ro T	yr V	al (Gly	Met	As	n Gl	u As	p Le	u Ph	ie L	/8 G	lu 7	lle.	G1,	1 GA	- A	A GA	LG TO	G ATC P Ile	180
														_			010		P AL	g GI	u Tr	p Ile	60
181	. TA	C G	G A	GG G	AG 1	TTC	GAG	TTO	. AA	A GA	A (2)	тст	יא ביי					•				C GTT	
61	ту	r Gl	u A	rg G	lu F	he	Glu	Phe	Lv	s G1	1) Aa	n Va	1 7.	- 01	AG G	GG	GAA	. CG	r Gr	C GA	T CT	C GTT u Val	240
												P 14	LLy	8 G.	tu G	ΤĀ	Glu	Arg	y Va	l As	p Le	u Val	80
241	TT	T GA	.G GC	C G	וכ פ	: ממ	A CC	صننب	mar														
81	Ph	e Gl	u G1	v va	ם בי	an 7	rh-	7.00	TCC	j GA'	r GT	r ta	CT	G AA	C G	GT (GTT	TAC	CT	GGJ	A AG	CACC	300
				,	^		1111	ren	Ser	AS	Va.	l Ty:	r Le	u As	n G	ly '	Val	Tyr	Let	Gly	/ Sei	C ACC	100
301																							
101	GL	A GA	C AT	G TI	C A	TC (BAG	TAT	CGC	TTC	GA:	GTO	: AC	3 AA	C GI	rg 1	TG	ААА	GAA	AAG	AAT	CAC	350
	GI	ı As	p me	c Ph	e I.	le G	lu	Tyr	Arg	Phe	: Asp	Val	Thi	As	n Va	l I	eu	Lys	Glu	Lys	Asn	CAC His	120
361	CTC	AA(GT	G TA	C AT	CA A	AA '	TCT	CCC	ATC	AGA	GTT	CCC	AA	A AC	тс	TC	GAG	CAG	ממ	TAC	GGG	420
121	Leu	Lys	Va.	l Ty	r Il	le L	ys s	Ser	Pro	Ile	Arg	Val	Pro	Lys	s Th	r L	eu	Glu	Gln	Asn	Tyr	Gly	420 140
																				•			
421	GTC	CTC	GG	GG	r cc	T G	AA (SAT	CCC	ATC	AGA	GGA	TAC	ATI	A AC	A A	n n	ccc	~~~	TAT		_	
141	Val	Leu	Gly	/ G1	y Pr	0 G	lu 2	\sp	Pro	Ile	Arg	Gly	Tyr	Ile	Are	a T	ve	27.a	CAG	TAT Tyr	TCG	TAC	480
											_	•	•				, .	~==	GIH	ryr	ser	Tyr	160
401	GGA	TGG	GAC	TG	G GG	T G	CC A	GA	ATC	GTT	מים	NCC.	CCM	3 mm			_			TAC			
161	Gly	Trp	Asp	Tr	Gl	y Al	la A	ıra	Ile	Val	Thr	200	Clin	All	160	ei A⊿	AA I	CCC	GTC	TAC Tyr	CTC	GAG	540
						-		-			****	361	GLY	116	TI	e r	ys I	Pro	Val	Tyr	Leu	Glu	180
541	GTG	TAC	AGG	GC	CG	Tr (~1	- Tree	'N.C.	~~~														
181	Val	Tyr	Aro	Ala	Ar	n 1.e		7-	GAT	TCA	ACG	GCT	TAT	CTG	TTC	G G	AA (CTT	GAG	GGG	AAA	GAT	600
		•			* ***	9 110	- 4	±11 1	Asp	ser	Thr	Ala	Tyr	Leu	Leu	ı Gi	lu I	ieu .	Glu	GGG	Lys	Asp	200
601	GCC	C TTTT																					
201	Ala	Tan	010	AGG	GT	G AA	AC G	GT :	TTC	GTA	CAC	GGG	GAA	GGA	AAT		rc a	ATT	GTG	GAA	GTT	TAT	660
	,,,,,,	Den	val	Arg	Va.	l As	n G	ly i	Phe	Val	His	Gly	Glu	Gly	Asn	ı Le	eu J	le	Val	GAA Glu	Val	Tyr	220
661	GTA	AAC	GGT	GAA	. AA	G AI	'A G	GG (GAG	TTT	CCT	GTT	CTT	GAA	AAG	A.	ac r	GA	440	AAG	CTC	ም ምረ	720
221	val	Asn	Gly	Glu	Ly	B Il	e G	ly (Glu	Phe	Pro	Val	Leu	Glu	Lys	. A.s	n c	ilv	Gin	AAG Lys	Leu	Dha	720
																							240
721	GAT	GGA	GTG	TTC	CAC	CT	G A	AA (SAT	GTG	AAA	CTA	TCC	ፐስጥ						GGG			
241	Asp	Gly	Val	Phe	His	s Le	u L	ys #	\sp	Val	Lvs	Leu	Trn	Tur	000	TO		VAC	GTG	GGG Gly	AAA	CCG	780
		•							•		., .	4	٠.٢	TAL	PIO	T	pΑ	sn	Val.	Gly	Lys	Pro	260



781 TAC CTG TAC GAT TTC GTT TTC GTG TTC AND GAG TTC	
781 TAC CTG TAC GAT TTC GTT TTC GTG TTG AAA GAC TTA AAC GGA GAG ATC TAC AGA GAA GA 261 Tyr Leu Tyr Asp Phe Val Phe Val Leu Luc Lac Lac Lac Lac Lac Lac Lac Lac Lac La	A 840
261 Tyr Leu Tyr Asp Phe Val Phe Val Leu Lys Asp Leu Asn Gly Glu Ile Tyr Arg Glu Gl	u 280
841 AAG AAA ATC GGT TTG AGA AGA AGA	
841 AAG AAA ATC GGT TTG AGA AGA GTC AGA ATC GTT CAG GAG CCC GAT GAA GAA GGA AAA ACC	900
281 Lys Lys Ile Gly Leu Arg Arg Val Arg Ile Val Gln Glu Pro Asp Glu Glu Gly Lys Thi	300
901 TTC ATA TTC GAA ATC ANG COT CO.	
901 TTC ATA TTC GAA ATC AAC GGT GAG AAA GTC TTC GCT AAG GGT GCT AAC TGG ATT CCC TCA	960
301 Phe Ile Phe Glu Ile Asn Gly Glu Lys Val Phe Ala Lys Gly Ala Asn Trp Ile Pro Ser	320
	•
961 GAA AAC ATC CTC ACG TGG TTG AAG GAG GAA GAT TAC GAA AAG CTC GTC AAA ATG GCA AGG	1020
321 Glu Asn Ile Leu Thr Trp Leu Lys Glu Glu Asp Tyr Glu Lys Leu Val Lys Met Ala Arg	340
	510
1021 AGT GCC AAT ATG AAC ATG CTC AGG GTC TGG GGA GGA GGA ATC TAC GAG AGA GAG ATC TTC	1080
341 Ser Ala Asn Met Asn Met Leu Arg Val Trp Gly Gly Gly Ile Tyr Glu Arg Glu Ile Phe	360
	500
TOTAL CIC IGI GAT GAA CTC GGT ATC ATG GTG TCC GAG	1140
361 Tyr Arg Leu Cys Asp Glu Leu Gly Ile Met Val Trp Gln Asp Phe Met Tyr Ala Cys Leu	1140 380
	200
THE COUNTY CAT CAT CAG THE AGA ANA CHE COUNTY	1200
381 Glu Tyr Pro Asp His Leu Pro Trp Phe Arg Lys Leu Ala Asn Glu Glu Ala Arg Lys Ile	400

AGA TAC CAT CCC TCC ATT CTC TCC TCC	1260
401 Val Arg Lys Leu Arg Tyr His Pro Ser Ile Val Leu Trp Cys Gly Asn Asn Glu Asn Asn	1260 420
	420
1261 TGG GGA TTC GAT GAA TGG GGA AAT ATG GCC AGA AAA GTG GAT GGT ATC AAC CTC GGA AAC	1220
421 Trp Gly Phe Asp Glu Trp Gly Asn Met Ala Arg Lys Val Asp Gly Ile Asn Leu Gly Asn	1320 440
	440
THE CIC TIC GAT TIT CCT GAG ATT TCT CCG GALL	1380
441 Arg Leu Tyr Leu Phe Asp Phe Pro Glu Ile Cys Ala Glu Glu Asp Pro Ser Thr Pro Tyr	460
	400
1381 TGG CCA TCC AGT CCA TAC GGC GGT GAA AAA GCG AAC AGC GAA AAG GAA GGA GAC AGG CAC 461 Trp Pro Ser Ser Pro Tyr Gly Gly Gly	1446
461 Trp Pro Ser Ser Pro Tyr Gly Gly Glu Lys Ala Asn Ser Glu Lys Glu Gly Asp Arg His	1440
	480
1441 GTC TGG TAC GTG TGG AGT GGC TGG ATG AAC TAC GAA AAC TAC GAA AAA GAC ACC GGA AGG	
481 Val Trp Tyr Val Trp Ser Gly Trp Met Asn Tyr Glu Asn Tyr Glu Lys Asp Thr Gly Arg	1500
	500
1501 TTC ATC AGC GAG TTT GGA TTT CAG GGT GCT CCC CAT CCA GAG ACG ATA GAG TTC TTT TCA	
501 Phe Ile Ser Glu Phe Gly Phe Gln Gly Ala Pro His Pro Glu Thr Ile Glu Phe Phe Ser	1560
	520
1561 AAA CCC GAG GAA AGA GAG ATA TTC CAT CCC GTC ATG CTG AAG CAC AAC AAA CAG GTG GAA	
521 Lys Pro Glu Glu Arg Glu Ile Phe His Pro Val Met Leu Lys His Asn Lys Gln Val Glu	1620
Figure 16b(continued)	540
- B Tob(Continued)	

162	1 G	GA (CAG	GA.	A A	A T	rg at	rc ad	G T	C A	та т	דר מ	GA B	ייית	~~~								GAC	
543	1 G	ly c	ln	Gli	u Az	g Le	u Il	Le Ai	ra Ph		1 n		\	W.	1 1-1	GG	A A	G TO	IA TE	AA G	AT T	TC	GAC	1680
						•					ie P	16 G	TA W	sn	Phe	Gl	y Ly	a C	ra Ly	/S A	sp P	he	GAC Asp	560
1681	AC	ST T	TT	GTC	3 ТА	ידי די.	יה דר		~ ~-			_												
561	. Se	er P	he	Va 1	75.	- 7-			G CT	CAA	AC CA	NG G	CG G	AG (GCG	AT	CAA	G TT	C GG	T GI	T G	AA	CAC	1740
				742	y	r ne	u se	r GI	n Le	u As	in Gl	[א ת	a G	lu A	Ala	Ile	e Ly	s Ph	e Gl	y Va	1 G	lu	His	580
1741	TG	G C	GA .	AGC	A.C.	ממ כי	מיחיים	~ ,,																
581	Tr	מ מ	ra (Sar	h w		- m		A AC	ع و و	C GG	C GC	T C	rc I	TC	TGG	CA	TT	AA C	C GA	C AC	3C	TGG	1800
		,	. 5 .	367	MI.	, Ly	3 1y	r Ly	Th:	r Al	a Gl	y Al	a Le	u P	he	Trp	Glr	Phe	Ası	l As	⊃ Se	r ·	Trp	600
1801	cc																							
601	7			rrc	AGC	TGC	TCC	: GCJ	GTO	GA:	TAC	TT	C AA	A A	GG	CCC	AAA	GCI	CTC	TAC	מידי י	٠.	יי איז	
801	PEC	o Va	T P	he	Ser	Trp	Ser	Ala	Val	Asp	туг	Phe	Ly	s A	rg	Pro	Lys	Ala	Leu	Tur	T-1.	_ 1	 	1860
																	•			- , -	. у.		ŊΈ	620
1861	GCG	AG.	A A	GA.	TTC	TTC	GCT	GAA	GTT	СТА	ccc	GTT	ידידיי	. az	\C 1									
621	Ala	Ar	3 A	rg	Phe	Phe	Ala	Glu	Val	Leu	Pro	Val	7.01		1		AGA	GAC	AAC	AAA	ATA	ı G	AA	1920
												• • • •	Det	ı Dy	, a 1	-ys	arg	Asp	Asn	Lys	Ile	G	lu	640
1921	CTG	CTO	3 G'	rg	GGT	GAG	CGA	TCT	GAG	GGA	GAC	AAA	ac a	, AC	T (m .c							
641	Leu	Let	ı Va	al (Gly	Glu	Arq	Ser	Glu	G1 v	Aen	Lare	7.50			.10	-	CAG	GCT	TGC	AGC	C	TA	1980
							_			,		L, a	A19	36	r L	eu	ser	Gln	Ala	Сув	Ser	L	eu	660
1981	CGA	GAA	G.	VA (GGG	AGA	AAA	car	ידייף מ	CC 3		~~~												
661	Arg	Glu	Gl	u c	Slv	Ara	Lva	Clu	ATT	N	AAA	GAC	TTA	CAC	G A	AC (GGT	ACT	CCC	AGC	AGA	CC	3G	2040
					,		_, 5	Gry	Ile	Arg	Lys	Asp	Leu	Gli	n A	sn (Gly	Thr	Pro	Ser	Arg	Ar	g	680
2041	TGT	GAG	TT	тс	GT	TGA	20	55																
	Сув						68																	

Figure 16 c(continued)



Figure No. 17a-Bankia gouldí (37gp4)

	1 A'	TG A	AA A	аа а	AT (CTA	СТ	A A	יים ידיי	т-т- л.	* * *	~ ~	~~ ·										
	1 M	et Ly	ys L	vs A	sn i	Len	1.0	n Ma	. D			·	-	ice .	TAT	. CI	'A C	TT	IG T	TT T	TA A	TG CTG	60
				•				G 116	ic Pi	ie r	ys A	rg L	eu T	nr	Tyr	Le	u Pr	:0 L	eu P	he L	eu M	et Leu	20
6:	י רי	יר די	ים מי	T 3 8 7	-m -																		
21	Le	11 Se		4A A		rca 	GTA	A GC	T CA	A TO	T C	CT G	TA G	AA A	AAA	CA	T GG	c co	T T	ra c	AA G1	TT GAC	120
	-		- 2	-u 31	er s	er	va.	LAI	a Gl	n Se	r Pi	co Va	al G	lu I	ys	Hi	s Gl	y Ar	g Le	u G	n Va	1 Asp	40
			_																				
121	. GG	А АА	.C .C.	C AT	TT C	TT	AAT	GC	G TC	T GG	A GA	A A1	TA	G A	.GC	TT	GC	T GG	T AA	C AG	ССТ	C TTT	180
4.1	GI	y AS	n Ar	g II	.e L	eu	Asn	Al	a Se	r Gl	y Gl	u Il	e Th	ır s	er	Leu	Ala	a Gl	y As	n Se	r Le	u Phe	60
181	TG	G AG	T AA	T GC	T G	GA ·	GAC	ACC	TC	GA'	T TT	T TA	T AA	T G	CA	GAA	ACT	GT	r ga	T TT	T TT	A GCA	240
61	Tr	Se	r As	n Al	a G	ly,	Asp	Thi	Sei	. As	Ph	е Ту	r As	n A	la :	Glu	Thr	Va:	i As	o Ph	e Lei	ı Ala	80
							·																
241	GA	L AAC	TG	G AA	TAC	3C :	TCA	CTT	ATI	` AGA	ATZ	A GC	TA 1	G GC	GC (GTA	AAA	GAZ	L AAT	י יוכע	. (3)	GGC	300
81	Glu	Ası	Tr	As:	n Se	er s	Ser	Leu	Ile	Arg	Ile	Ala	Me	t Gl	у 1	Val	Lys	Glu	Asr	Tr	Asn	Gly	100
																	•			•			200
301	GGA	AAT	GG(TA	r at	т	AT	AGT	CCG	CAG	GAG	CAR	GA	A GC	T 2	AAA	ATT	AGA	222	CTT	ATT	CAT	760
101	Gly	Asn	Gl	Ty:	r Il	e A	sp	Ser	Pro	Gln	Glu	Glr	Gli	1 Al	a L		Ile	Ara	Lvs	Val	Ile	Agn	360
																			-, -	• • • •	110	vaħ	120
361	GCA	GCT	ATT	GC1	C AA	.c g	GC	ATA	TAT	GTA	ATA	ATA	GAC	י ידכי	G C	יא כי	n Orr	CNC	~		GAG		
121	Ala	Ala	Ile	Ala	As	n G	ly	Ile	Tyr	Val	Ile	Ile	Asc	Tr	o c	nc.	Thr	UNC	Clu	GCA	GAG Glu	TTA	420
									-					,			****	nib	GIU	ATG	GIU	Leu	140
421	TAC	ACA	GAT	GAG	GC	T G	TT	GAC	ттт	ململمك	ACC	a Ca	a mo								ACT		
141	Tyr	Thr	Asp	Glu	Al	a V	al.	Asp	Phe	Phe	Thr	Ara	Mar	יינט י	4 G	AC	CTA	TAC	GGA	GAT	ACT Thr	CCC	480
										•	••••	A. y	Mec	VI.	1 A	вр	Leu	TYT	GIY	Asp	Thr	Pro	160
481	AAT	GTA	ATG	TAT	GA	ים ב	. مايد	דמד	חממ	CNC	~~	272											
161	AAT Asn	Val	Met	Tyr	Gli	ı I	le '	Tvr	Asn	GAG	Dro	ALA	TAC	CAA	A A	GT	TGG 	CCT	GTT	ATT	AAG	AAT	540
				-1-				• , -	V911	GIL	PLO	116	ıyr	GII	1 5	er	Trp	Pro	Val	Ile	Lys	Asn	180
541	TAT	GCA	GAG	CD D	GT!	A 25-	, 17 -17	~~~	CC#														
181	Tyr	Ala	Glu	Gln	Val	1 A.	11 (SCI Nia	GGT Cl	ATA	CGT	TCT	AAA	GAC	: C	CA (GAT	AAT	TTA	ATA	ATT	GTA	600
	-	-				• •	,	114	GIY	116	Arg	ser	Lys	Asp	P:	ro i	Asp	Asn	Leu	Ile	Ile	Val	200
601	GGT	ם ביי	300					.															
201	Glv	The	AGC	AAT	TAT	r To	er c	CAG	CAA	GTT	GAT	GTA	GCA	TCA	G	CA (GAC	CCA	ATA	TCT	GAT	ACT	660
	,	****	267	MBII	Tyl	. 56	er (Jin	Gln	Val	Asp	Val	Ala	Ser	: A	la	Asp	Pro	Ile	Ser	Asp	Thr	220
661																							
	AAT	GTG	GCA	TAT	ACT	T	TA (CAT	TTT	TAT	GCA	GÇA	TTT	AAC	: co	CG (CAT	GAT	AAC	TTA	AGA	AAT	720
	Asn	AUT	WIF	Tyr	Thi	: Le	eu }	lis	Phe	Tyr	Ala	Ala	Phe	Asn	Pı	ro I	His	Asp	Asn	Leu	Arg	Asn	240
721 241	GTA Val	GCA	CAG	ACA	GC	T	CA C	GAT	AAT	AAT	GTT	GCT	TTG	TTT	G:	TT 2	ACA	GAA	TGG	GGT	ACA	ATT	780
441	Val	Ala	Gln	Thr	Ala	Le	eu A	A sp	Asn	Asn	Val	Ala	Leu	Phe	Va	al :	Thr	Glu	Trp	Gly	Thr	Ile	260

781 TTA BAT AGG GG
781 TTA AAT ACC GGA CAA GGA GAA CCA GAC AAA GAA AGC ACT AAT ACT TGG ATG GCC TTT TTG 840
The Ash The Tro Mer Ala Dan
AAA GGT ATA AGT CAC GCT AAT TOO
281 Lys Glu Lys Gly Ile Ser His Ala Asn Trp Ser Leu Ser Asp Lys Ala Phe Pro Glu Thr 300
300 msp bys Ala Phe Pro Glu Thr
901 GGG TCT GTA GTT CAA GCA GGA CAA GGT GTA TCT GGT TTA ATT AGC AAT AAA CTT ACA GCC 960
JO1 Gly Ser Val Val Gln Ala Gly Gln Gly Val Ser Gly Leu Ile Ser Asn Lys Leu Thr Ala 320
961 TCT GGT GAA ATT GTA AAA AAC ATC ATC CAA AAC TGG GAT ACA GAG ACC TCT ACA GGA CCT 1020
321 Ser Gly Glu Ile Val Lys Asn Ile Ile Gln Asn Trp Asp Thr Glu Thr Ser Thr Gly Pro 340
340
1021 AAA ACA ACA CAA TGT AGT ACT ATA GAA TGT ATT AGA
1021 AAA ACA ACA CAA TGT AGT ACT ATA GAA TGT ATT AGA GCT GCA ATG GAA ACA GCA CAA GCA 1080 341 Lys Thr Thr Gln Cys Ser Thr Ile Glu Cys Ile Arg Ala Ala Met Glu Thr Ala Gln Ala 360
of the Alg Ala Met Glu Thr Ala Gln Ala 360
1081 GGA GAT GAA ATT ATA ATT GCC CCT GGA AAC TAC AAT TTT CAA GAC AAG ATA CAA GGT GCC 1140
361 Gly Asp Glu Ile Ile Ile Ala Pro Gly Asn Tyr Asn Phe Gln Asp Lys Ile Gln Gly Ala 380
380 Ash Tyr Ash Phe Gln Asp Lys Ile Gln Gly Ala
1141 TTT AAC CGT AGT GTT TAG CTT TAT GGT
1141 TTT AAC CGT AGT GTT TAC CTT TAT GGT AGT GCT AAC GGA AAC AGT ACA AAC CCT ATT ATA 1200
381 Phe Asn Arg Ser Val Tyr Leu Tyr Gly Ser Ala Asn Gly Asn Ser Thr Asn Pro Ile Ile 400
1201 TTA AGA GGC GAA AGC GCT ACA ANG GGT AGA
1201 TTA AGA GGC GAA AGC GCT ACA AAC CCT CCT GTT TTC TCA GGA TTA GAT TAT AAC AAT GGC 1260
401 Leu Arg Gly Glu Ser Ala Thr Asn Pro Pro Val Phe Ser Gly Leu Asp Tyr Asn Asn Gly 420
1261 TAC CTA TTA AGT ATT GAA CCT CAM Day
1261 TAC CTA TTA AGT ATT GAA GGT GAT TAT TGG AAT ATT AAA GAT ATA GAG TTT AAA ACT GGG 1320
421 Tyr Leu Leu Ser Ile Glu Gly Asp Tyr Trp Asn Ile Lys Asp Ile Glu Phe Lys Thr Gly 440
1321 TCT AAA GGT ATT GTT CTT GAC AAT TCT AAT GGT AGT AAA TTA AAA AAC CTT GTT CAT 1380 441 Ser Lys Gly Ile Val Leu Asp Asn Ser Asn Gly Ser Lys Leu Lys Asn Leu Val Val His 460
Ash Ser Ash Gly Ser Lys Leu Lys Ash Leu Val Val His 460
1381 GAT ATT GGA GAA GAA GCT ATT CAC TTG CGT GAT GGA TCT AGC AAT AAT AGT ATA GAT GGT 1440
461 Asp Ile Gly Glu Glu Ala Ile His Leu Arg Asp Gly Ser Ser Asn Asn Ser Ile Asp Gly 480
1441 TGC ACT ATA TAC AAT ACA GGT AGA AGA
1441 TGC ACT ATA TAC AAT ACA GGT AGA ACT AAA CCT GGT TTT GGT GAA GGT TTA TAT GTA GGC 1500
481 Cys Thr Ile Tyr Asn Thr Gly Arg Thr Lys Pro Gly Phe Gly Glu Gly Leu Tyr Val Gly 500
1501 TCA GAT AAA GGA CAA CAT GAC ACT TAT GAA AGA GCT TGT AAC AAT AAC ACT ATT GAA AAC 1560
501 Ser Asp Lys Gly Gln His Asp Thr Tyr Glu Arg Ala Cys Asn Asn Thr Ile Glu Asn 520
1561 TGT ACC GTT GGA CCC AAT GTA ACA GCA GAA GGC GTA GAT GTT AAG GAA GGT ACA ATG AAC 1620 521 Cys Thr Val Gly Pro Asn Val Thr Ala Gly Gly Val Arm White Acc 1620
val Asp val Lys Glu Glu The
27 Old Gly Inr Met Asn 540

Figure 17b(continued)

PCT/US97/22623

WO 98/24799

43/46

1621 ACT ATT ATA AGA AAT TGC GTG TTT TCT GCA GAA GGA ATT TCA GGA GAA AAT AGC TCA GAT 541 Thr Ile Ile Arg Asn Cys Val Phe Ser Ala Glu Gly Ile Ser Gly Glu Asn Ser Ser Asp 560 1681 GCT TTT ATT GAT TTA AAA GGA GCC TAT GGT TTT GTA TAC AGA AAC ACG TTT AAT GTT GAT 561 Ala Phe Ile Asp Leu Lys Gly Ala Tyr Gly Phe Val Tyr Arg Asn Thr Phe Asn Val Asp 1740 580 1741 GGT TCT GAA GTA ATA AAT ACT GGA GTA GAC TTT TTA GAT AGA GGT ACA GGA TTT AAT ACA 581 Gly Ser Glu Val Ile Asn Thr Gly Val Asp Phe Leu Asp Arg Gly Thr Gly Phe Asn Thr 1800 600 1801 GGT TTT AGA AAT GCA ATA TTT GAA AAT ACA TAT AAC CTT GGC AGT AGA GCT TCA GAA ATT 601 Gly Phe Arg Asn Ala Ile Phe Glu Asn Thr Tyr Asn Leu Gly Ser Arg Ala Ser Glu Ile 1860 620 1861 TCA ACT GCT CGT AAA AAA CAA GGT TCT CCT GAA CAA ACT CAC GTT TGG GAT AAT ATT AGA 621 Ser Thr Ala Arg Lys Lys Gln Gly Ser Pro Glu Gln Thr His Val Trp Asp Asn Ile Arg 1920 1921 AAC CCT AAT TCT GTT GAT TTT CCA ATA AGT GAT GGT ACA GAA AAT CTA GTA AAT AAA TTC 641 Asn Pro Asn Ser Val Asp Phe Pro Ile Ser Asp Gly Thr Glu Asn Leu Val Asn Lys Phe 660 1981 TGC CCA GAT TGG AAT ATA GAA CCA TGT AAT CCT GTA GAC GAA ACC AAC CAA GCA CCT ACA 661 Cys Pro Asp Trp Asn Ile Glu Pro Cys Asn Pro Val Asp Glu Thr Asn Gln Ala Pro Thr 680 2041 ATA AGC TTC CTA TCT CCT GTT AAC AAT ATT ACT TTA GTT GAA GGT TAT AAT TTA CAA GTT 681 Ile Ser Phe Leu Ser Pro Val Asn Asn Ile Thr Leu Val Glu Gly Tyr Asn Leu Gln Val 2100 700 2101 GAA GTT AAT GCT ACT GAT GCA GAT GGA ACT ATT GAT AAA GTT TAT ATA GAT AAC 701 Glu Val Asn Ala Thr Asp Ala Asp Gly Thr Ile Asp Asn Val Lys Leu Tyr Ile Asp Asn 2160 720 2161 AAT TTA GTT AGG CAA ATA AAT TCT ACT TCA TAT AAA TGG GGC CAT TCT GAT TCT CCA AAT 721 Asn Leu Val Arg Gln Ile Asn Ser Thr Ser Tyr Lys Trp Gly His Ser Asp Ser Pro Asn 2220 740 2221 ACA GAT GAA CTT AAT GGT CTT ACA GAA GGA ACT TAT ACC TTA AAA GCA ATT GCA ACT GAT 741 Thr Asp Glu Leu Asn Gly Leu Thr Glu Gly Thr Tyr Thr Leu Lys Ala Ile Ala Thr Asp 2280 760 2281 AAC GAC GGG GCT TCT ACA GAA ACG CAA TTT ACG TTA ACT GTA ATA ACA GAA CAA AGT CCG 761 Asn Asp Gly Ala Ser Thr Glu Thr Gln Phe Thr Leu Thr Val Ile Thr Glu Gln Ser Pro 2340 780 2341 TCT GAG AAT TGT GAC TTT AAT ACA CCT TCT TCA ACT GGT TTA GAA GAT TTT GAC ATT AAA 781 Ser Glu Asn Cys Asp Phe Asn Thr Pro Ser Ser Thr Gly Leu Glu Asp Phe Asp Ile Lys 2400 800 2401 AAG TIT TCT AAC GIT TIT GAG TTA GGA TCT GGC GGA CCA TCT TTA AGT AAT TTA AAA ACA 2460

Figure 174 (continued)

80	l L	/S P	Phe	Ser	As	n Va	.1 P)	ie G	lu	Leu	G1:	y Se	r G	ly G	lly :	Pro	Ser	Leu	Se:	r As	an I	Leu	Lys	Thr	82
246] 82]	l TI	TA	CT .	ATT	AA	TG	G AA	ТТ	CG	CAA	ተአረ	~ nn	T CC						•					AAC Asn	
2521 841							_	•				Dec	Lys	PE	-0 L	ys I	le 1	rhr	Phe	Glr	ı Pi	ie I	Lys	Asn	2580 860
2581 861							TCT		_			361	Leu	110	e Pr	0 A	sn P	he A	dsp	Gly	As	p T	yr '	Trp	2640 880
881	GTA Val	AC	A TO	CA G	AT.	AAC Asn	GGT Gly	AAT Asn	TT Ph	rr c	TG /	ATG Met	GTA Val	TCI Ser	Ly:	A AC	T A	AT A Sn A	AT sn i	TTT Phe	AC(3 A:	FA 1	TAC Yr	2700 900
	TTT Phe											ys ,	nsn	val	Thr	Pro) Se	r As	n G	ln	Ile	Se	r L	ys	2760 920
										• • •		ys L	eu 1	yr	Pro	Asn	Pro	Al	a L	eu A	\sp	GAJ Glu	A AC	T	2820 940
941]	ATT T	TT he	GTG Val	Se:	C G(CT G	AA c	iAT .sp	GAA Glu	AA Ly	A CI S Le	TA G	CT T	TG (GTG Val	CTT Leu	GTA Val	CC Pro	A G1		70 56				

Figure 17d(continued)

Figure No. 180 Pyrococcus furiosus VC1(7EG1)

leader sequence: amino acids 1-24
_
9 18 27 36 45 54
AIG AGC AAG AAA AAG TTC GTC ATC GTA TCT ATC TTA
Met Ser Lys Lys Phe Val Ile Val Ser Ile Leu Thr Ile Leu Leu Val Gln
and the Leu Val Gln
63 72
GCA ATA TAT TTT GTA GAA AAG TAT CAT ACC TCT GAG GAC AAG TCA ACT TCA AAT
Ala Ile Tyr Phe Val Glu Lys Tyr His Thr Ser Glu Asp Lys Ser Thr Ser Asn
117 126 135 144 153 162
ACC TCA TCT ACA CCC CAA ACA ACA CTT TCC ACT ACT
Thr Ser Ser Thr Pro Pro Gln Thr Thr Leu Ser Thr Thr Lys Val Leu Lys Ile
and the Lys Ile
171 180 189 199
AGA TAC CCT GAT GAC GGT GAG TGG CCA GGA GCT CCT ATT GAT AAG GAT GGT GAT
Arg Tyr Pro Asp Asp Gly Glu Trp Pro Gly Ala Pro Ile Asp Lys Asp Gly Asp
225 234 243 252 261 270
GGG AAC CCA GAA TTC TAC ATT GAA ATA AAC CTA TGG AAC ATT CTT AAT GCT ACT
Gly Asn Pro Glu Phe Tyr Ile Glu Ile Asn Leu Trp Asn Ile Leu Asn Ala Thr
to bed Ash Ale Thr
279 288 297 306 315
GGA TTT GCT GAG ATG ACG TAC AAT TTA ACC AGC GGC GTC CTT CAC TAC GTC CAA
Gly Phe Ala Glu Met Thr Tyr Asp Leu mby C
Gly Phe Ala Glu Met Thr Tyr Asn Leu Thr Ser Gly Val Leu His Tyr Val Gln
333 342 25
342 351 360
CIT GAC AAC ATT GTC TTG AGG GAT AGA AGT AAT TCG GTG
Gln Leu Asp Asn Ile Val Leu Arg Asp Arg Ser Asn Trp Val His Gly Tyr Pro
387 396 405 414 423
GAA ATA TTC TAT GGA AAC AAG CCA TGG AAT GCA AAC TAC GCA ACT GAT GGC CCA
Glu Ile Phe Tyr Gly Asn Lys Pro Trp Asn Ala Asn Tyr Ala Thr Asp Gly Pro
And Ash Tyr Ala Thr Asp Gly Pro
441 450 450
459 468 488
ATA CCA TTA CCC AGT AAA GTT TCA AAC CTA ACA GAC TTC TAT CTA

ATA CCA TTA CCC AGT AAA GTT TCA AAC CTA ACA GAC TTC TAT CTA ACA ATC TCC Ile Pro Leu Pro Ser Lys Val Ser Asn Leu Thr Asp Phe Tyr Leu Thr Ile Ser

TAT AAA CTT GAG CCC AAG AAC GGC CTG CCA ATT AAC TTC GCA ATA GAA TCC TGG
Tyr Lys Leu Glu Pro Lys Asn Gly Leu Pro Ile Asn Phe Ala Ile Glu Ser Trp

549
558
567
576
585
580
580

TTA ACG AGA GAA GCT TGG AGA ACA ACA GGA ATT AAC AGC GAT GAG CAA GAA GTA
Leu Thr Arg Glu Ala Trp Arg Thr Thr Gly Ile Asn Ser Asp Glu Gln Glu Val

603 612 621 630 639 648 ATG ATA TAG TAT GAC GGA TTA CAA CCG GCT GGC TCC AAA GTT AAG GAG Met Ile Trp Ile Tyr Tyr Asp Gly Leu Gln Pro Ala Gly Ser Lys Val Lys Glu

ATT GTA GTC CCA ATA ATA GTT AAC GGA ACA CCA GTA AAT GCT ACA TTT GAA GTA Ile Val Val Pro Ile Ile Val Asn Gly Thr Pro Val Asn Ala Thr Phe Glu Val

711 720 729 738 747 756

TGG AAG GCA AAC ATT GGT TGG GAG TAT GCT GCA TTT AGA ATA AAG ACC CCA ATC

Trp Lys Ala Asn Ile Gly Trp Glu Tyr Val Ala Phe Arg Ile Lys Thr Pro Ile

AAA GAG GGA ACA GTG ACA ATT CCA TAC GGA GCA TTT ATA AGT GTT GCA GCC AAC Lys Glu Gly Thr Val Thr Ile Pro Tyr Gly Ala Phe Ile Ser Val Ala Ala Asn

819 828 837 846 855 864
ATT TCA AGC TTA CCA AAT TAC ACA GAA CTT TAC TTA GAG GAC GTG GAG ATT GGA
Ile Ser Ser Leu Pro Asn Tyr Thr Glu Leu Tyr Leu Glu Asp Val Glu Ile Gly

873 882 891 900 909 918

ACT GAG TTT GGA ACG CCA AGC ACT ACC TCC GCC CAC CTA GAG TGG TGG ATC ACA

Thr Glu Phe Gly Thr Pro Ser Thr Thr Ser Ala His Leu Glu Trp Trp Ile Thr

927 936 945 954

AAC ATA ACA CTA ACT CCT CTA GAT AGA CCT CTT ATT TCC TAA 3'
Asn Ile Thr Leu Thr Pro Leu Asp Arg Pro Leu Ile Ser *

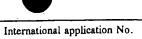
Figure 18b(continued)

INTERNATIONAL SEARCH REPORT

International application No. PCT/US97/22623

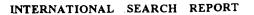
A. CLASSIFICATION OF SUBJECT MATTER IPC(6) :C07H 21/04; C12N 1/20, 1/14, 5/00, 9/38, 9/42; C08B 30/04 US CL :435/207, 209, 252.3, 254.11, 274, 275, 320.1, 325; 536/23.2 According to International Patent Classification (IPC) or to both national classification and IPC B. FIELDS SEARCHED Minimum documentation searched (classification system followed by classification symbols) U.S. : 435/207, 209, 252.3, 254.11, 274, 275, 320.1, 325; 536/23.2 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)									
Please See Extra Sheet.									
C. DOCUMENTS CONSIDERED TO BE RELEVANT									
Category* Citation of document, with indication, where a	ppropriate, of the relevant passages Relevant to claim No.								
 GRABNITZ et al. Structure of the Clostridium thermocellum: Sequence A of Cellulases and β-Glycosidases Included Hydrolase. Eur. J. Biochem. Septem pages 301-309, see entire document. VOORHORST et al. Characterization β-Glucosidase from the Hyperthermocoli. J. Bacteriol. December 1995, Vo. 7111, see entire document. 	nalysis Reveals a Superfamily species II spe								
Further documents are listed in the continuation of Box	C. See patent family annex.								
Special catagories of cited documents: "A" document defining the general state of the art which is not considered to be of particular relevance "B" cartier document published on or after the international filing date "L" document which may throw doubts on priority claim(a) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means									
the priority date claimed	Date of mailing of the international search report								
Date of the actual completion of the international search 26 MARCH 1998	2 1 APR 1998								
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer LISA J. HOBBS, PH.D. Telephone No. (703) 308-0196								

INTERNATIONAL SEARCH REPORT



PCT/US97/22623

R		Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
Th	is inter	mational report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1.		Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2.		Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3.		Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box	k II C	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
Thi	s Inter	mational Searching Authority found multiple inventions in this international application, as follows:
	Ple	ease See Extra Sheet.
1.		As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2.		As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	X	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.: -11, species I-III
4.		No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
R	emark	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.





International application No. PCT/US97/22623

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS and STN (Bioscience and Patent Indexes): Desulfurococc##, Staphylotherm##, Thermatoga, galactosidase#, glucosidase#, beta glucosidase#, beta glucosidase#. Genbank, EMBL, ESTs1-4, STS, N-Geneseq: Seq. ID Nos.: 1-3 and A-Geneseq, PIR, Swissprot: Seq ID Nos.: 15-17.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains claims directed to more than one species of the generic invention. These species are deemed to lack Unity of Invention because they are not so linked as to form a single inventive concept under PCT Rule 13.1. The species are as follows: there are 18 distinct enzymes disclosed in the description, as enumerated in Figs. 1-18 and Table 1.

The claims are deemed to correspond to the species listed above in the following manner: while all the claims form one Group for examination, each of the claims is generic to the 18 enzyme species disclosed.

The species listed above do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, the species lack the same or corresponding special technical features for the following reasons: each enzyme is a different product, thus has the special technical feature of the recited enzyme, which the other species lack.